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Screening cotton varieties (*Gossypium hirsutum* L.) for heat tolerance under field conditions

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Fifteen upland cotton *Gossypium hirsutum* L., were evaluated for heat tolerance based on agronomical and physiological characteristics under field conditions where temperature exceeded 40°C in July and August. Genotypes showed statistically significant differences for stomatal conductance, photosynthetic yield, fluorescence, photosynthetically active radiation (PAR), seed cotton yield, plant height, boll weight, seed cotton weight, number of seeds per boll and 100 seed weight. In the study, relative cell injury level (RCIL), ranged from 54.56 to 79.44% and stomatal conductance ranged from 264.86 to 570.50 mol m⁻²s⁻¹. Associations between investigated traits indicated that there were some positive correlations. These data indicated that photosynthetic yield, fluorescence, chlorophyll content (SPAD value), cell membrane thermostability (CMT), plant height and number of bolls per plant can be used for improving seed cotton yield. However, among these measurements CMT, SPAD value and fluorescence were more practical in large breeding trials. The results showed that AGC 375 and AGC 208 cotton varieties had lower relative cell injury level (higher cell membrane thermostability) and higher seed cotton yield and photosynthetic yield than other investigated varieties.

Key words: Cotton, heat tolerance, physiological traits, cell membrane thermostability (CMT).

INTRODUCTION

High temperature stress is one of the most important abiotic stress influencing productivity of cotton. Therefore, identification of cotton cultivars with high- temperature tolerance would be beneficial in both current and future climates (Kakani et al., 2005). The temperature increase

is expected to continue 1.5 to 5.9°C within the next century because of global warming (Hodges and McKinion, 1996). Thus, if global warming occurs as projected, cotton production in the future will be reduced, and breeding heat tolerant cotton cultivars will be necessary. Cultural practices, such as earlier planting, may be used to avoid the flowering of cotton in the high temperatures that occur during mid to late summer (Reddy et al., 2002).

During the growing stage, cotton is sensitive to high temperature stress, especially flowering and boll-formation period. Early studies revealed that maximum number of bolls and squares retain occur at 30/22°C day/night temperatures (Reddy et al., 1992a). Mean maximum temperatures in the range of 27 to 32°C are more desirable during the period of boll development and

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Abbreviations: PAR, Photosynthetically active radiation; CMT, cell membrane thermostability; LEL, leaf electrolyte leakage; CTD, canopy temperature depression, VPD, leaf air vapor pressure deficit; RCIL, relative cell injury level; QTLs, quantitative trait loci.

maturation (Gipson and Joham, 1968; Mauney, 1974; Freeland et al., 2006). Plants gained more total biomass and partitioned more of it to bolls and squares at 30/20°C day/night temperatures than any other temperature regimes examined (Reddy et al., 1991).

Bibi et al. (2008) reported that the optimum temperature for photosynthetic carbon fixation of cotton was approximately 33°C and that photosynthesis in cotton decreased significantly at temperatures of 36°C and above. High temperature environments of 35 to 40°C are frequently associated with cotton sterility and boll retention problems (Reddy et al., 1992b). Number of bolls produced, bolls retained, and percent retention were progressively reduced as time per day at 40°C was increased.

The physiological processes affecting the overall performance of cotton and their associations with high temperature stress are reported by some researchers. According to these researchers, there were strong negative association between photosynthesis and high temperature (Schrader et al., 2004; Bibi et al., 2004a, b), photosynthetically active radiation (PAR) and maximum temperature (Bhardwaj and Sing, 1991). On the other hand, some researchers indicated that there were positive association between photorespiration and high temperature (Krieg, 1986), stomatal conductance and transpiration (Kolb and Robberecht, 1996).

Cell membrane thermostability (CMT) has been proposed by Sullivan (1972), as a means for measuring the amount of electrolyte leakage from leaf disks bathed in deionized water after exposure to heat treatment and this method has been used as a measure of heat tolerance in several crops, including rice, soybean, potato, tomato and cotton (Sing et al., 2007; URL 1). CMT has been used in cotton as an appropriate screening and selection criterion for heat tolerance owing to its ability to discriminate among heat sensitive and heat tolerant genotypes within the species (Malik et al., 1999; Ashraf et al., 1994, Rahman et al., 2004, Azhar et al., 2009).

The use of relatively new physiological techniques such as CMT, leaf electrolyte leakage (LEL), carbon isotope discrimination; ecophysiological based remote sensing/infrared techniques such as canopy temperature depression (CTD), leaf air vapor pressure deficit (VPD), chlorophyll fluorescence; and biochemical parameters such as chlorophyll a and b contents and a:b ratio are gaining popularity to screen efficiently and quickly with reliability (Singh et al., 2007).

Breeding programs with most agricultural crops have successfully increased yields by empirical selection for higher yields, without any explicit selection for yield-related physiological traits (Lu et al., 1998). Selection for higher yields in Pima cotton and bread wheat appear to have generated selection pressures for higher stomatal conductances that are independent of operating pressures for higher photosynthetic rates. Stomatal conductance could be a valuable selection criterion for

higher yields in irrigated crops grown at supra-optimal temperatures (Lu et al., 1998; Ulloa et al., 2000; Rahman, 2005). Chlorophyll fluorescence and membrane leakage were the most sensitive and practical techniques for quantifying tolerance to high temperatures in both controlled and field conditions (Oosterhuis et al., 2009). Photosynthesis was also sensitive but was not practical for screening large numbers of genotypes for temperature tolerance (Bibi et al., 2008).

Singh et al. (2007) indicated that better understanding of the possible impact of high temperature stress on physiological, morphological, and yield processes would not only help in mitigating the adverse effects of high temperature stress, but also in developing reliable field screening tools, however screening tools used for stress tolerance are difficult to operate and time consuming. One of the most important and economic way to overcome negative effect of heat stress is to identify and/or develop heat tolerant cultivars. Current commercial cotton cultivars do not appear to have much tolerance to high temperatures (Brown and Oosterhuis, 2005). In recent years, some cotton varieties were developed and registered for superior lint yield, fiber length, and competitive fiber strength under heat stress environments (Ulloa et al., 2006).

Most cotton heat tolerance studies have been conducted in the greenhouse and growth chambers. The objective of this study was to determine the effect of high temperature stress on some agronomical and physiological parameters of cotton under field studies and to use efficiently the most appropriate screening tools and physiological traits associated with cotton yield.

MATERIALS AND METHODS

Field experiment was carried out at the GAP International Agricultural Research and Training Center's experimental area during 2010 cotton growing season in Diyarbakır/Turkey. The experimental field is located (37°55'36" N, 40°13'49" E) at 670 m above sea level. Generally, this region is characterized by a semi-arid continental climate with very hot and dry summers and cold, rainy or snowy winters. The long-term (62 years) average annual temperature is 15.8°C, total rainfall is 491 mm and the average relative humidity is about 29.9%.

The meteorological data of the experimental site during the study period have been presented in Figures 1 and 2. Both maximum and mean temperatures during the experiment were higher than that of long term periods, and precipitation were inadequate during sowing time of experiments when compared with long term precipitation at the sowing time (Figure 1). On the contrary, long term precipitations were higher than that of the experiment at harvest time (Figure 2).

The soil is low in organic matter and phosphorus. It has adequate potassium, calcium, and high clay content (49 to 67%) in the 0 to 150 cm profile.

The experiment was laid out as randomized complete block design with four replications. In this study 15 cotton varieties were used as plant material. Seeds of these cotton varieties were planted with a combine cotton drilling machine on 17th May, before sowing total herbicide (200 ml da⁻¹ doses) were applied for weed control and once herbicides at the doses 60 ml/da was applied after emergence for *Sorghum halepense*.

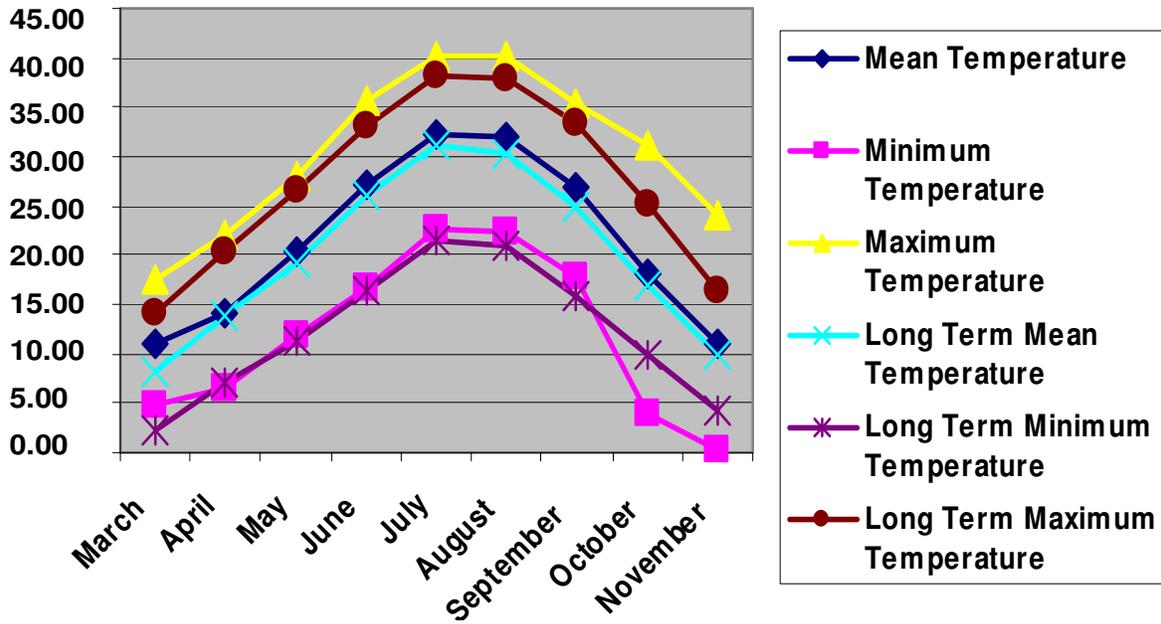


Figure 1. Monthly average and maximum temperature (°C) during experiment and long term period.

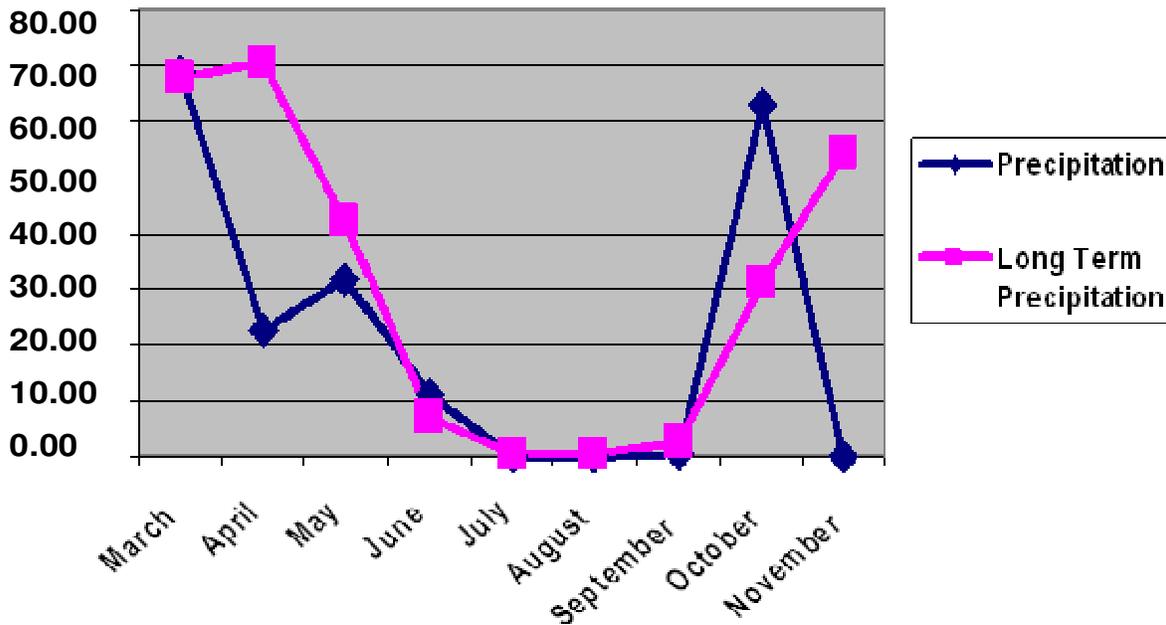


Figure 2. Average monthly precipitation levels (mm) and long term values.

Each plot consisted of four rows of 12 m. The between and within the row spacing was 0.70 and 0.20 m, respectively. All plots were treated with 20-20-0 composite fertilizer to provide 70 kg N ha⁻¹ and 70 kg P₂O₅ ha⁻¹. Just before flowering, 70 kg N ha⁻¹ were applied as ammonium nitrate as an additional N dose. Insects were monitored throughout the experiment and no insect control was necessary during these growing seasons. Plants were grown under recommended cultural practices for commercial production; the experiment was thinned and hoed three times by hand and two times with a machine. Experimental plots were irrigated by drip

irrigation method. Adequate irrigation was provided for minimizing effect of drought stress during cotton growing stage, especially in the reproductive stage. In the study, five plants were selected randomly for observations in each plot. All physiological traits were measured 80 days after planting, the time of peak flowering and fruiting. After all bolls had opened, a 50-boll sample was collected from each plot by hand-harvesting for boll weight, seed cotton weight per boll and number of seeds per boll. Plots were harvested twice by hand and yield of four rows of plot were weighed and calculated for seed cotton yield. First harvest was done on 22

Table 1. Average values of the investigated agronomical, morphological and physiological traits of different 15 cotton varieties.

| Cultivar | PH | BN | BW | SCW | NSB | 100 SW | CC | LT | SC | RCIL | PAR | FL | PY | SCY |
|-------------------|----------------------|-------|---------------------|---------------------|----------------------|----------------------|-------|-------|-----------------------|-------|----------------------|------------------------|----------------------|------------------------|
| 1. DP 396 | 88.73 ^{b-e} | 18.06 | 6.57 ^{cd} | 5.12 ^{cd} | 33.22 ^{c-g} | 8.50 ^l | 46.96 | 29.36 | 541.85 ^{ab} | 54.73 | 718.30 ^a | 3870.33 ^{f-h} | 47.00 ^{d-f} | 3474.52 ^{a-c} |
| 2. DP 90 | 90.06 ^{b-e} | 16.73 | 6.72 ^{a-d} | 5.23 ^{a-d} | 32.55 ^{d-g} | 9.31 ^{e-g} | 44.93 | 28.73 | 570.50 ^a | 60.16 | 435.80 ^e | 5038.70 ^b | 44.93 ^{ef} | 2997.65 ^{cd} |
| 3. DP 499 | 94.80 ^{a-d} | 14.53 | 6.78 ^{a-c} | 5.32 ^{a-d} | 34.44 ^{a-f} | 9.17 ^{f-h} | 45.06 | 29.36 | 469.61 ^{a-c} | 60.71 | 336.90 ^f | 4867.20 ^{bc} | 51.66 ^{cd} | 3123.41 ^{a-d} |
| 4. STV 453 | 89.60 ^{b-e} | 16.26 | 6.71 ^{a-d} | 5.24 ^{a-d} | 36.00 ^{a-c} | 10.31 ^{a-c} | 46.70 | 26.66 | 291.23 ^{de} | 65.82 | 458.46 ^{de} | 4005.60 ^{e-g} | 47.20 ^{c-f} | 3225.91 ^{a-d} |
| 5. STV 468 | 86.33 ^{c-e} | 15.66 | 5.60 ^f | 4.34 ^e | 30.00 ^g | 8.62 ^{hi} | 45.76 | 28.20 | 382.75 ^{b-e} | 67.93 | 767.40 ^a | 6780.40 ^a | 42.20 ^f | 3079.08 ^{cd} |
| 6. STV 474 | 88.13 ^{b-e} | 15.46 | 6.60 ^{b-d} | 5.16 ^{b-d} | 32.77 ^{c-g} | 9.58 ^{d-f} | 46.93 | 28.10 | 496.60 ^{a-c} | 63.92 | 279.63 ^g | 4092.20 ^{ef} | 50.80 ^{c-e} | 3477.77 ^{a-c} |
| 7. SJ-U 86 | 95.33 ^{a-c} | 15.46 | 6.83 ^{a-c} | 5.36 ^{a-c} | 34.66 ^{a-e} | 9.45 ^{ef} | 45.03 | 26.36 | 461.45 ^{a-c} | 70.71 | 476.80 ^{de} | 3695.50 ^{gh} | 52.93 ^{b-d} | 2572.81 ^{de} |
| 8. AGC 85 | 86.13 ^{de} | 13.60 | 6.44 ^{c-e} | 5.17 ^{b-d} | 36.66 ^{ab} | 8.50 ^l | 44.06 | 27.73 | 264.86 ^e | 55.13 | 288.90 ^{fg} | 3549.40 ^{hi} | 51.53 ^{cd} | 3783.96 ^a |
| 9. AGC 208 | 85.06 ^e | 15.53 | 6.02 ^{d-f} | 4.76 ^{de} | 31.11 ^{fg} | 9.43 ^{e-g} | 46.96 | 26.83 | 403.85 ^{b-e} | 64.72 | 297.76 ^{fg} | 3693.13 ^{gh} | 58.73 ^{ab} | 2750.55 ^d |
| 10. AGC 375 | 95.66 ^{ab} | 19.86 | 6.50 ^{cd} | 5.11 ^{cd} | 35.33 ^{a-d} | 8.85 ^{g-i} | 49.80 | 26.93 | 341.25 ^{c-e} | 54.56 | 242.30 ^g | 3767.33 ^{f-h} | 60.50 ^a | 3771.19 ^{ab} |
| 11. Fiber Max 819 | 96.73 ^{ab} | 16.46 | 5.75 ^{ef} | 4.41 ^e | 30.33 ^g | 9.15 ^{f-h} | 47.23 | 26.96 | 458.85 ^{a-c} | 61.15 | 444.33 ^{de} | 4516.60 ^{cd} | 50.76 ^{c-e} | 3151.54 ^{a-d} |
| 12. Fiber Max 832 | 92.66 ^{b-e} | 16.00 | 7.30 ^{ab} | 5.78 ^a | 37.44 ^a | 9.83 ^{c-e} | 47.13 | 27.40 | 510.23 ^{ab} | 66.42 | 636.73 ^b | 4251.20 ^{de} | 53.50 ^{bc} | 1955.07 ^{ef} |
| 13. Fiber Max 958 | 92.46 ^{b-e} | 16.30 | 7.36 ^a | 5.74 ^{ab} | 33.77 ^{b-f} | 10.12 ^{b-d} | 48.63 | 26.93 | 436.25 ^{a-d} | 60.16 | 321.66 ^f | 2942.50 ^j | 48.86 ^{c-e} | 1794.00 ^f |
| 14. Acala 1517-95 | 97.26 ^{ab} | 16.46 | 6.85 ^{a-c} | 5.34 ^{a-d} | 34.44 ^{a-f} | 10.45 ^{ab} | 42.73 | 25.26 | 471.55 ^{a-c} | 63.18 | 536.46 ^c | 3322.60 ^l | 50.70 ^{c-e} | 3094.84 ^{b-d} |
| 15. Acala 1517-99 | 103.46 ^a | 15.06 | 6.93 ^{a-c} | 5.34 ^{a-d} | 31.88 ^{e-g} | 10.78 ^a | 44.63 | 25.26 | 400.00 ^{b-e} | 79.44 | 501.33 ^{cd} | 4001.20 ^{e-g} | 49.20 ^{c-e} | 3144.04 ^{a-d} |
| Mean | 92.16 | 16.10 | 6.60 | 5.16 | 33.64 | 9.47 | 46.17 | 27.34 | 433.39 | 63.25 | 449.52 | 4159.59 | 50.70 | 3026.42 |
| CV (%) | 5.93 | 15.83 | 6.51 | 6.78 | 6.09 | 3.69 | 5.47 | 10.53 | 22.73 | 21.67 | 7.84 | 5.04 | 7.49 | 13.41 |
| LSD (0.05) | 9.11* | ns | 0.72** | 0.58** | 3.42** | 0.57** | ns | ns | 164.09* | ns | 58.75** | 349.28** | 6.32** | 67.64** |

* and **, Significant at 0.05 and 0.01 level of probability, respectively; PH, plant height; BN, boll number (per/plant); BW, boll weight (g); SCW, seed cotton weight per boll (g); NSB, number of seeds per boll; 100 SW, 100 seed weight (g); CC, chlorophyll content (SPAD value); LT, leaf temperature; SC, stomatal conductance ($\text{mol m}^{-2}\text{s}^{-1}$); RCIL, relative cell injury level (%); PAR, photosynthetically active radiation, FL, fluorescence, PY, photosynthetic yield, SCY, seed cotton yield (kg ha^{-1}).

October, 2010; second harvest was done on 10 November, 2010. The samples were ginned on a laboratory roller gin, and seed samples of 100 seeds were taken from each of the replicates of field plots for 100 seed weight. Statistical analyses were performed using JMP 5.0.1 statistical software (<http://www.jmp.com>) and the means were grouped with LSD (0.05) test.

Measurement of physiological traits

All physiological traits were measured at the first week of August, approximately 80 days after planting, the time of peak flowering and fruiting. Measurements were taken during mid-day on the fifth fully expanded leaf below the terminal with one reading per leaf of the plant according to Johnson and Saunders (2003). Photosynthesis was

measured using the EARS-PPM Plant Photosynthesis System. Photosynthetic yield ($\mu\text{mol CO}_2 \text{ m}^{-2}\text{s}^{-1}$), photosynthetic active radiation (PAR) and fluorescence were measured by this tool. Stomatal conductance ($\text{mol m}^{-2}\text{s}^{-1}$) was measured by a leaf porometer (Model SC-1), leaf temperature was measured by an infrared thermometer (DT-8811H) and leaf chlorophyll contents were measured by a Minolta SPAD-502 chlorophyll meter. Relative cell injury level (RCIL), a measurement for cell membrane thermostability (CMT) under high temperature stress, was determined according to Sullivan (1972).

RESULTS AND DISCUSSION

The analysis of variance of the investigated traits

indicated that significant differences among the varieties were obtained for plant height, boll weight, seed cotton weight per boll, number of seeds per boll, 100 seed weight, stomatal conductance, PAR, fluorescence, photosynthetic yield and seed cotton yield. Non-significant differences were observed for boll number per plant, chlorophyll content (SPAD value), leaf temperature and RCIL as seen in Table 1.

The analysis of variance indicated that there were significant differences ($P \leq 0.05$) among the varieties for stomatal conductance. Among the genotypes screened, DP 90, DP 396, Fiber Max 832, Stoneville 474, DP 499, Acala 1517-95 and

SJ-U 86 had the highest stomatal conductance value. Comparisons of stomatal responses in cultivars with contrasting agronomic characteristics have been reported by Roark and Quisenberry (1977), but no conclusive evidence has emerged on defined relations between stomatal conductance and yield. Recent studies have used such an approach to investigate the relationship between stomatal conductance and yield and revealed that low yielding lines showed a lower stomatal conductance than high yielding advanced lines (Lu et al., 1998).

Lu et al. (1997) observed that an upland cotton cultivar DP 90 showed 25 to 35% higher stomatal conductance, 35 to 50% higher photosynthetic rate, and 45% smaller leaf area than Pima S-6. Furthermore, the higher photosynthetic rate and stomatal conductance of DP 90 leaves were partly related to their sun-tracking ability.

RCIL ranged from 54.56 to 79.44%. Among the cotton cultivars AGC 375, DP 396 and AGC 85 had the lowest relative injury level (higher cell membrane thermostability), while Acala 1517-99, SJ-U 86, Stoneville 468, Fiber Max 832 and Stoneville 453 had the highest. However, cultivar differences were not significant statistically (Table 1). Azhar et al. (2009) used relative cell injury percentage to measure heat tolerance in cotton, and reported that heat tolerant accessions had more stable yield and yielded more seed cotton with better quality fiber than the heat-intolerant accessions across four environments.

Wang (1988), indicated that when plants are under high-temperature stress, the structure of membrane is altered, permeability increases, electrolyte leakage increases, and eventually the cell dies. Rahman et al. (2004) indicated that exposure to high temperature prior to the CMT test produced better distinction between heat-tolerant and heat-susceptible cultivars; however, they cautioned regarding its indirect selection on the basis of seed cotton yield under non-heat stressed environments.

The cultivar effect on photosynthetic yield, fluorescence and PAR are shown in Table 1. There were significant differences between cultivars for fluorescence, photosynthetic yield and PAR at ($P \leq 0.01$) probability level.

The data revealed that the Stoneville 468, DP 90 and DP 499 cotton cultivars showed statistically higher fluorescence value when compared to all the other cultivars. Stoneville 468, DP 396 and Fiber Max 832 showed numerically higher PAR compared to all the other cultivars. AGC 375 and AGC 208 had the highest value for photosynthetic yield.

The cultivars used in the experiment AGC 85, AGC 208, AGC 375 and SJ-U 86 were registered as modern cultivars for heat tolerance and better fiber quality (Ulloa et al., 2006). The other used standard cultivars (Stoneville and DP cultivars) commonly have been planting in this region. The results of this study showed that photosynthesis could be used as indicator of heat tolerance, but these results were inconsistent with Bibi et

al. (2008), who reported that photosynthesis sensitivity was not practical for screening large numbers of genotypes for temperature tolerance. This inconsistency may be stem from differences of the used photosynthesis measurement devises.

Among the varieties, Acala 1517-99 was taller than the other cotton genotypes. The highest boll number were obtained from AGC 375 (19.86 per/plant) and DP 396 (18.06 per/plant) cotton genotypes, but differences between genotypes was non-significant for boll number per plant. Fiber Max 958 and Fiber Max 832 genotypes had the highest value for boll weight and seed cotton weight per boll and Fiber Max 832 also had the highest value for number of seed per bolls.

Comparing the response to high temperature of all the cultivars used in this study revealed that cultivar variation existed for seed cotton yield at ($p \leq 0.01$) probability level, ranging from 1794.00 to 3783.96 kg ha⁻¹. Among the cotton cultivars AGC 85, AGC 375, Stoneville 474, DP 396, Stoneville 453, Fiber Max 819, Acala 1517-99 and DP 499 had the highest seed cotton yield (Table 1). The lowest seed cotton yields were obtained from Fiber Max 958 and Fiber Max 832, respectively. Oosterhuis et al. (2009) screened 134 entries from the Arkansas Variety tests to heat tolerance; only PHY370WR and DP515BGRR exhibited both tolerance to elevated temperatures as well as an ability to recover from the high temperature without any subsequent detrimental effect. In general, the majority of the 134 lines tested to date did not show any appreciable tolerance to high temperatures. They reported that the tolerant lines selected from the study (PHY370WR and DP515BGRR) will be compared with the yields in field tests that experienced heat stress.

Correlation coefficient between seed cotton yield, yield components and some morphological and physiological traits are presented in Table 2. Seed cotton yield positively correlated with chlorophyll content, fluorescence, photosynthetic yield, plant height and boll number. Seed cotton yield negatively correlated with leaf temperature, stomatal conductance, RCIL, PAR, boll weight, seed cotton weight, number of seeds per boll and 100 seed weight. The results of this study indicated that photosynthesis yield, fluorescence, chlorophyll content, cell membrane thermo stability, plant height and the number of bolls per plant can be used as selection criterion for seed cotton yield. Similar results were also reported by Malik et al. (1999), Rahman, (2005), Cottee et al. (2007), Bibi et al. (2008), Khan et al. (2008), Azhar et al. (2009) and Oosterhuis et al. (2009). Consistent with previous research, cultivar differences in yield were correlated with fruit retention that affected boll number, which reflected that high temperature stress during flowering induced fruit abscission that limited yield (Cottee et al., 2010). In that study, which was carried out under tents and ambient field conditions, stomatal conductance was not found to be correlated with seed cotton yield, however a strong positive correlation was

Table 2. Correlation coefficients between seed cotton yield, yield components and physiological traits.

| Parameter | LT | SC | RCIL | PAR | FL | PY | PH | BN | BW | SCW | NSB | 100 SW | SCY |
|-----------|---------|---------|-----------|---------|----------|-----------|---------|---------|-----------|-----------|----------|----------|----------|
| CC | -0.1518 | -0.0308 | 0.1506 | -0.1801 | -0.0645 | 0.2248 | -0.1751 | 0.0818 | -0.0173 | -0.0270 | 0.0212 | -0.1152 | 0.0241 |
| LT | - | 0.2182 | -0.3833** | 0.0705 | 0.1816 | -0.2546 | -0.1920 | -0.1220 | -0.1700 | -0.1502 | -0.0586 | -0.2625 | -0.0733 |
| SC | - | - | 0.1235 | 0.2051 | 0.0280 | -0.1220 | -0.1225 | -0.1181 | 0.1427 | 0.0964 | -0.0672 | -0.0011 | -0.3106* |
| RCIL | - | - | - | 0.0818 | 0.0672 | 0.1812 | -0.0342 | -0.1082 | 0.0772 | 0.0409 | -0.0207 | 0.1847 | -0.0816 |
| PAR | - | - | - | - | 0.4626** | -0.5200** | 0.0261 | 0.0384 | -0.0973 | -0.1265 | -0.1600 | -0.0338 | -0.1477 |
| FL | - | - | - | - | - | -0.4164** | -0.1195 | -0.0085 | -0.4134** | -0.4375** | -0.3739* | -0.3103* | 0.1546 |
| PY | - | - | - | - | - | - | 0.0740 | 0.1724 | 0.0706 | 0.1010 | 0.3338* | -0.0992 | 0.0212 |
| PH | - | - | - | - | - | - | - | 0.3476* | 0.4478** | 0.4103** | 0.0488 | 0.4956** | 0.0011 |
| BN | - | - | - | - | - | - | - | - | 0.1843 | 0.1820 | 0.0429 | -0.0133 | 0.1461 |
| BW | - | - | - | - | - | - | - | - | - | 0.9903** | 0.5451** | 0.4888** | -0.3024* |
| SCW | - | - | - | - | - | - | - | - | - | - | 0.5927** | 0.4427** | -0.2830 |
| NSB | - | - | - | - | - | - | - | - | - | - | - | 0.0394 | -0.0076 |
| 100 SW | - | - | - | - | - | - | - | - | - | - | - | - | -0.3278* |

*Correlation that is significant between measurements for $p < 0.05$; ** Correlation that is significant between measurements for $p < 0.01$; PH, plant height; BN, boll number (per/plant); BW, boll weight (g); SCW, seed cotton weight per boll (g); NSB, number of seeds per boll; 100 SW, 100 seed weight (g); CC, chlorophyll content (SPAD value); LT, leaf temperature; SC, stomatal conductance ($\text{mol m}^{-2} \text{s}^{-1}$); RCIL, relative cell injury level (%); PAR, photosynthetically active radiation, FL, fluorescence, PY, photosynthetic yield, SCY, seed cotton yield (kg ha^{-1})

reported between stomatal conductance and transpiration rate. Previous studies showed that stomatal conductance can be used as selection criterion for seed cotton yield, however, in this study such a correlation was not obtained. The reason for these differences can be related with measuring time and the quantity of observations. Some literatures have confirmed these results.

According to Radin et al. (1994), the physiological benefits of very high stomatal conductance are unexplained. Ulloa et al. (2000) reported that the strong environmental dependence of stomatal conductance and the large number of required measurements may limit the breeder's use of stomatal conductance as a selection criterion. Therefore, they suggested that quantitative trait loci (QTLs) and molecular markers provide breeders alternatives for selection.

Stomatal conductance at high temperature is positively correlated with stomatal sensitivity to temperature, and independent from photosynthesis (Lu and Zeiger, 1994).

In addition, the differences in stomatal conductance between the low- and high-yielding pima lines is under genetic control (Percy et al., 1996; Ulloa et al., 2000). Rahman (2005) revealed that the selection efficiency of stomatal conductance in segregating populations was likely to be affected by the complexity of its inheritance, environmental dependency, and presence of substantial non-allelic and genotype \times temperature regime interactions. Radin et al. (1994) determined that yield was more closely related to conductance in the afternoon than to that determined in the morning.

The stomatal component of heat resistance is apparently dependent on evaporative cooling.

The result of this study provided evidence that there are cultivar differences with respect to stomatal conductance, photosynthetic yield and seed cotton yield.

Therefore, this information is important in the development of adaptation studies, particularly concerning heat stress. Further confirmation of this approach is needed, that is, correlation between seed cotton yield and stomatal conductance of genotypes under field and controlled conditions.

There are many mechanisms by which plants are able to tolerate higher temperatures. Pollen germination, pollen tube growth and boll retention have been proposed as tools to identify cultivars tolerant to high temperature tolerance (Kakani et al., 2005; Liu et al., 2006). It is estimated that these methods are more sensitive therefore studies are needed for the current and future

climates.

Conclusion

In the cotton growing areas of Turkey, especially Southeastern Anatolia Region, several episodes of temperatures higher than 35°C usually occur in mid-July to mid-August. It is known that optimum temperature of cotton during growth and boll formation period is between 27 and 32°C. If global warming occurs as projected, cotton production in the future will be reduced and most of cotton production in semi-arid regions probably will be more affected. Therefore, this study represents an initial field investigation into the impact of heat stress on cotton yields in this region. The results of this study indicated that photosynthetic yield, fluorescence, chlorophyll content (SPAD value), CMT, plant height and the number of bolls per plant can be used as selection criterion for seed cotton yield in breeding studies for heat tolerance.

REFERENCES

- Ashraf M, Saeed MM, Qureshi MJ (1994). Tolerance to high temperature in cotton (*Gossypium hirsutum* L.) at initial growth stages. *Environ. Exp. Bot.* 34:275-283.
- Azhar FM, Ali Z, Akhtar MM, Khan AA, Trethowan R (2009). Genetic variability of heat tolerance, and its effect on yield and fibre quality traits in upland cotton (*Gossypium hirsutum* L.). *Plant Breed.* 128:356-362.
- Bhardwaj SN, Singh M (1991). Thermal relation of light utilization and biomass production in upland cotton (*Gossypium hirsutum* L.). In "Proceedings of Indo US Workshop on Impact of Global Climatic Changes on Photosynthesis and Plant Productivity, January 8-12, 1991 (I. P. Abrol, Ed.), pp. 651-662. Oxford and IBH Publishing Company Ltd., New Delhi, India.
- Bibi AC, Oosterhuis DM, Gonias ED, Bourland FM (2004a). Evaluation of Techniques for Quantifying the Physiological Response of Cotton to High Temperature. *Summaries of* <http://arkansasagnews.uark.edu/533-4.pdf>
- Bibi AC, Oosterhuis DM, Gonias ED, Bourland FM (2004b). Screening a Diverse Set of Cotton Cultivars for High Temperature Tolerance. <http://arkansasagnews.uark.edu/533-5.pdf>
- Bibi AC, Oosterhuis DM, Gonias ED (2008). Photosynthesis, quantum yield of photosystem II and membrane leakage as affected by high temperatures in cotton genotypes. *J. Cotton. Sci.* 12:150-159.
- Brown RS, Oosterhuis DM (2005). High Daytime temperature stress effects on the physiology of modern versus obsolete cotton cultivars. *Summaries of Cotton Research in 2004. Arkansas Agric. Exp. Station Res. Series* 533:63-67.
- Cottee NS, Tan DKY, Cothren T, Bange M P, Campbell LC (2007). Screening Cotton Cultivars for Thermotolerance under Field Conditions. <http://www.icac.org/meetings/wcrc/wrc4/presentations/data/papers/Paper2234.pdf>
- Cottee NS, Tan DKY, Bange MP, Cothren JT, Campbell LC (2010). Multi-Level Determination of Heat Tolerance in Cotton (*Gossypium hirsutum* L.) under Field conditions. *Crop Sci.* 50:2553-2564
- Freeland Jr TB, Pettigrew B, Thaxton P, Andrews GL (2006). Agrometeorology and cotton production, pp. 1-17. Chapter 13A in *guide to agricultural meteorological practices*, 3rd Ed.
- Gipson JR, Joham HE (1968). Influence of night temperature on growth and development of cotton (*Gossypium hirsutum* L.). I. Fruiting and boll development. *Agron. J.* 60:292-295.
- Hodges HF, McKinion JM (1996). Food and Agriculture in the 21st Century: A Cotton Example. *World Resour. Rev.* 8:80-97, USA.
- Johnson JR, Saunders JR (2003). Evaluation of Chlorophyll Meter for Nitrogen Management in Cotton. <http://msucares.com/nmrec/reports/2002/>
- Kakani VG, Reddy KR, Koti S, Wallace TP, Prasad PVV, Reddy V R, Zhao D (2005). Differences in in vitro Pollen Germination and Pollen Tube Growth of Cotton Cultivars in Response to High Temperature. *Ann. Bot.* 96:59-67.
- Khan AI, Khan IA, Sadaqat HA (2008). Heat tolerance is variable in cotton (*Gossypium hirsutum* L.) and can be exploited for breeding of better yielding cultivars under high temperature regimes. *Pak. J. Bot.* 40(5):2053-2058
- Kolb PF, Robberecht R (1996). High temperature and drought stress effects on survival of *Pinus ponderosa* seedlings. *Tree Physiol.* 16:665-672.
- Krieg DR (1986). Feedback control and stress effects on photosynthesis. *Proceeding of the Beltwide Cotton Conferences*, 227-243. National Cotton Council of America, Memphis, TN.
- Liu Z, Yuan Y, Liu S, Yu X, Rao L (2006). Screening for High-Temperature Tolerant Cotton Cultivars by Testing *In Vitro* Pollen Germination, Pollen Tube Growth and Boll Retention. *J. Integr. Plant Biol.* 48(6):706-714.
- Lu ZM, Zeiger E (1994). Selection for higher yields and heat resistance in pima cotton has caused genetically determined changes in stomatal conductances. *Phyiol. Plant.* 92:273-278.
- Lu Z, Chen J, Percy RG, Zeiger E (1997). Photosynthetic Rate, Stomatal conductance and Leaf Area in Two Cotton Species (*Gossypium barbadense* and *Gossypium hirsutum*) and their Relation with Heat Resistance and Yield. *Austr. J. Plant Phys.* 24:693-700.
- Lu Z, Percy RG, Qualset CO, Zeiger E (1998). Stomatal conductance predicts yields in irrigated Pima cotton and bread wheat grown at high temperatures. *J. Exp. Bot.* 49:453-460.
- Mauney JR (1974). Meteorological factors affecting the commercial production of cotton. Unpublished report. National Cotton Council of America, Memphis, TN.
- Malik MN, Chaudhry FI, Makhdam MI (1999). Cell membrane thermostability as a measure of heat-tolerance in cotton. *Pak. J. Sci. Ind. Res.* 42:44-46.
- Oosterhuis DM, Bourland FM, Bibi AC, Gonias ED, Loka D, Storch D (2009). Screening for Temperature Tolerance in Cotton. *Summaries of Arkansas Cotton Research in 2008, AAES Research Series* 573.<http://arkansasagnews.uark.edu/573-5.pdf>
- Percy RG, Lu Z, Radin JW, Turcotte EL, Zeiger E (1996). Inheritance of stomatal conductance in cotton (*Gossypium barbadense* L.) *Phys. Plant.* 96:389-394.
- Radin JW, Lu Z, Percy RG, Zeiger E (1994). Genetic variability for stomatal conductance in Pima cotton and its relation to improvements of heat adaptation. *Proc. Natl. Acad. Sci. USA.* 91:7217-7221.
- Rahman H, Malik SA, Saleem M (2004). Heat tolerance of upland cotton during the fruiting stage evaluated using cellular membrane thermostability. *Field.Crop Res.* 85:149-158.
- Rahman HU (2005). Genetic analysis of stomatal conductance in upland cotton (*Gossypium hirsutum* L.) under contrasting temperature regimes. *J. Agric. Sci.* 143:161-168.
- Reddy VR, Baker DN, Hodges HF (1991). Temperature effect on cotton canopy growth, photosynthesis and respiration. *Agron. J.* 83:699-704.
- Reddy KR, Hodges HF, McKinion JM, Wall GA (1992a). Temperature effects on pima cotton growth and development. *Agron. J.* 84:237-243.
- Reddy KR, Hodges HF, Reddy VR (1992b). Temperature Effects on Cotton Fruit Retention. *Agron. J.* 84:26-30.
- Reddy KR, Doma PR, Mearns LO, Boone MYL, Hodges HF, Richardson AG, Kakani VG (2002). Simulating the impacts of climate change on cotton production in the Mississippi Delta. *Climate Res.* 3:271-281
- Roark B, Quisenberry JE (1977). Environment and genetic components of stomatal behavior in upland cotton. *Plant Phys.* 59:354-356.
- Schrader SM, Wise RR, Wacholtz WF, Ort DR, Sharkey TD (2004). Thylakoid membrane responses to moderately high leaf

- temperature in Pima cotton. *Plant Cell Environ.* 27:725.
- Singh RP, Prasad PVV, Sunite K, Giri SN, Reddy KR (2007). Influence of High Temperature and Breeding for Heat Tolerance in Cotton: A Review
http://www.gri.msstate.edu/publications/docs/2007/01/3796Advances_in_Agronomy.pdf
- Sullivan CY (1972). Mechanisms of Heat and Drought Resistance in Grain Sorghum and Methods of Measurements. In *Sorghum in Seventies* (Eds.N.G.P. Rao and L.R. House). Oxford and IBH Publishing Co., New Delhi. pp. 247-264
- Ulloa M, Cantrell RG, Percy RG, Zeiger E, Lu Z (2000). QTL Analysis of Stomatal Conductance and Relationship to Lint Yield in an Interspecific Cotton. *J. Cotton Sci.* 4:10-18.
- Ulloa M, Percy RG, Hutmacher R, Cantrell RG (2006). Registration of SJ-U86 Cotton Germplasm Line with High Yield and Excellent Fiber Quality. *Crop Sci.* 46:2336-2338.
- Wang B (1988). Biological free radicals and membrane damage of plant. *Plant Physiol. Commun.* 2:12-16.
- URL 1. The Cell Membrane Stability (CMS) by the electro-conductivity method. <http://www.plantstress.com/methods/index.asp>.