Full Length Research Paper

# Influence of potassium solubilizing bacteria on crop productivity and quality of tea (*Camellia sinensis*)

B. Bagyalakshmi<sup>1</sup>\*, P. Ponmurugan<sup>1</sup> and S. Marimuthu<sup>2</sup>

<sup>1</sup>Department of Biotechnology, K. S. Rangasamy College of Technology, Tiruchengode -637 215, Namakkal District, Tamil Nadu, India.

<sup>2</sup>R&D Centre, Parry Agro Industries, Valparai – 642125, Tamil Nadu, India.

Accepted 6 June, 2012

A field experiment was conducted to study the efficacy of indigenous potassium solubilizing bacteria (KSB) in combination with various dosages of potash fertilizers along with recommended dose of nitrogen (N) and phosphorus (P) fertilizers in tea plants. Soil and leaf samples were drawn from the respective plots and they were subjected for the analysis of various parameters related to nutrients and quality aspects. Among various treatments, plants treated with  $N_{100}$   $P_{100}$   $K_{75}$  + KSB concentration formulation was found to be the best in terms of high chlorophyll, carotenoid, N, P and K contents in the crop shoots followed by other treatments. Significantly higher yield of the green leaf was achieved by the same treatment. K content in soil and also in crop shoots was greatly improved due to the application of KSB along with possible reduced doses of potash source. On the other hand, N<sub>100</sub> P<sub>100</sub> K<sub>25</sub> + KSB formulation and untreated control plots have exhibited least green leaf yield and nutrient status of soil and crop shoots. The biochemical parameters, total polyphenols, catechins, amino acids and sugars were significantly at higher level in plants after imposing treatments in combination with KSB. Biometric parameters such as plucking surface of the tea bush canopy, plucking points per unit area, internodal length, leaf moisture and dry matter contents were analyzed and found to be high in tea plants treated with  $N_{100} P_{100} K_{75}$  + KSB combination. Banji content was significantly reduced invariably in all the treatments except the untreated control plots. Evaluation of KSB population in soil revealed that wherever KSB was incorporated, there was a significant increase in population level and has coincided with dehydrogenase enzyme activity. The flush shoots of tea comprising of three leaves and a bud were subjected to manufacture black tea, and it was revealed that almost all the tea quality parameters such as theaflavin, thearubigin, highly polymerized substances, total liquor colour, caffeine, briskness, colour and flavour indexes were greatly improved in KSB treated plants, which in turn improve the quality as well. This finding confirms that the influence of indigenous potassium solubilizing bacteria upon potassium nutrient exhibited improvement in the productivity and nutrient uptake in plants and retained in soil including quality parameters in tea plantations.

Key words: Tea, potassium solubilizing bacteria, biofertilizer, nutrient status, crop productivity, tea quality.

# INTRODUCTION

Tea (*Camellia sinensis*) is an important commercial crop in many subtropical and tropical areas of the world. Tea, owing to its favorable benefits on human health, currently enjoys a great popularity among other beverages worldwide (Liang et al., 2007). The crop shoots are plucked at regular intervals (10 to 15 days) and removed a certain amount of various elements from the plant-soil system.

Essential nutrients have to be supplemented through various sources of fertilizer for higher productivity and maintenance of soil health. Next to nitrogen (N), potassium (K) is another major essential nutrient and it is being taken up by the crop plants in quite large quantities similar to or more than nitrogen. It has been reported that potassium and magnesium are required in large quantities, and they are both involved in almost all

<sup>\*</sup>Corresponding author. E-mail: bagyalakshmi09@gmail.com. Tel: +91 4288 274741 – 44. Fax: +91 4288 274745.

biological reactions in crop plants (Sedaghathoor et al., 2009). Potassium promotes root growth, stronger stem, and increases resistance to cold and water stress. It directly connects with improvement of the quality of crop, reduces pest and disease incidence by enhancing crop resistance as well. Although K is not a constituent of any organic molecule or plant structure, it is involved in numerous biochemical and physiological processes and pivotal role to plant growth, yield, quality and stress (Cakmak, 2005). In tea plantations, both seedlings and clones of tea are being cultivated under diverse soil types, areas with abundant rainfall and various agro climatic conditions. Due to perennial habitat of the tea plant and continuous application of inorganic NPK fertilizers (5-6 splits), the tea plantation soils are highly weathered, low in base status and are generally acidic in reaction.

There is a mutual relationship between soil microflora and minerals possessed in soil environment and these interactions have been extensively studied to produce technologies such as bio mineralization, bioremediation and bio hydrometallurgy (Rawlings, 2002).

Moreover, quality of made tea depends on organic and inorganic composition of harvested shoots, which are changed into the substances, these are responsible for taste, flavor and colour of made tea. The nutrients absorbed by crop shoots may influence the quality attributes in made tea. In which, potassium is the major nutrient element which influences on the quality of marketable tea (Venkatesan et al., 2006). Also, the important quality components of black tea were shown to be improved with increased K application in Chinese cultivation practices (Ruan and Hardter, 2001).

In this regard, supplement of balanced nutrition of tea is of particular importance to secure good harvested fresh leaves as a prerequisite for tea of superior quality. In recent years, the tea plantation mainly depends on synthetic inorganic source of fertilizers for higher yield and productivity. Deep dependency of such inorganic fertilizer remains demand, in future, is simultaneously harmful to soil health and environment. There are reports available for application of plant growth promoting bacteria and metals/minerals solubilizing bacteria to enhance the productivity and reduce banji contents, without affecting the quality of black tea (Mandal, 2007).

Experiments were conducted to evaluate the potential of potassium solubilizing bacteria (KSB) in soils planted with egg plant which revealed that KSB enhanced the plant growth and soil nutrient level significantly (Han and Lee, 2005). Similarly, *Frateuria aurantia* belonging to the family Pseudomonaceae obtained from the agricultural soils of Coimbatore region of Tamil Nadu, India was solubilized with K considerably, and this promoted the crop yield (Ramarethinam and Chandra, 2006). This solubilization effect is generally due to the production of certain organic acids and enzymes by KSB. In addition, they are also known to produce amino acids, vitamins and growth promoting substances like indole-3-acetic acid (IAA) and gibberellic acid (GA<sub>3</sub>) which help in better growth of the plants (Ponmurugan and Gopi, 2006). To keep this in mind, a study was undertaken to assess the influence of indigenous KSB in combination with various doses of potash fertilizers and the recommended dosage of nitrogen and phosphorus (P) fertilizers for improving yield potential and quality of tea.

# MATERIALS AND METHODS

# Selection of indigenous KSB strain for field application

The indigenous potassium solubilizing bacteria (Pseudomonas putida) strain obtained from tea estate of Valparai, Tamil Nadu, India was selected based on prior knowledge of their higher potassium solubilization rate and production of phytohormones (Bagyalakshmi et al., 2009). The selected KSB strain was formulated using talcum powder as carrier for field application with prior mixing with farm yard manure. This bioformulation was incubated for a week for rapid multiplication of KSB, and maintained population level as  $1 \times 10^9$  cfu/g before field application. The biopreparation was applied in soil around the tea bush by placement method (at a depth of 10 cm) at the rate of half kg/bush. The NK fertilizers were applied as per the recommendation by UPASI Tea Research Institution, Valparai, India, in four equal splits during April, May, September and November by broadcasting every year, avoiding very wet and dry periods. Pretreatment yield of the experimental blocks was recorded for three months before the start of the experiment. Applications of KSB bioformulation were carried out twice a year during April/May and September/October for a period of four years.

# **Field experiment**

A field trial was conducted at Parry Agro Tea Industries, Valparai, Tamil Nadu, India during 2008 to 2011 located in the Western Ghats of Southern India (10°30'N, 77°0'E, altitude 1050 m above mean sea level). The field was planted with a drought resistant clone UPASI-9. Experimental plots were laid out as randomized block design and plot size consisted of 20 bushes, and each was replicated three times. There were 8 treatments namely,

- 1. N<sub>100</sub>P<sub>100</sub>K<sub>100</sub> + KSB
- 2. N<sub>100</sub>P<sub>100</sub>K<sub>75</sub> + KSB
- 3. N<sub>100</sub>P<sub>100</sub>K<sub>50</sub> + KSB
- 4. N<sub>100</sub>P<sub>100</sub>K<sub>25</sub> + KSB
- 5. N<sub>100</sub>P<sub>100</sub>K<sub>100</sub> + KSB + VC
- 6. N<sub>50</sub>P<sub>50</sub>K<sub>50</sub> + KSB + VC
- 7.  $N_{100}P_{100}K_{100}$  alone
- 8. Control  $(N_0P_0K_0)$

Urea, rock phosphate and Muriate of Potash (MOP) were used as NPK sources of nutrients respectively as per the recommendations of UPASI. Moreover, vermicompost was also applied along with NPK for better efficiency, and regular culture practices other than manuring were followed as per the standard procedure (Verma and Palani, 1997).

#### Yield assessment

Green leaf yield was recorded at every harvesting regularly, during the course of the experimental period. Yield and yield attributes, particularly the percentage of banji occurrence was monitored. The green leaf yield was expressed into made tea as per the reference of Ponmurugan and Baby (2007).

Productivity index (PI) in response to soil application of KSB bioformulation was calculated according to the method of Sharma and Satyanarayana (1990).

#### Soil analysis

The soil samples were collected periodically and subjected to analyse various physicochemical parameters such as soil pH, EC, total organic carbon (Walkley and Black, 1934), nitrogen (AOAC, 1990), available phosphorus (Jackson, 1973), exchangeable potassium (Murphy and Riley, 1962) and calcium, sodium and magnesium (Bhargava and Raghupathi, 2001). Population level of KSB in the experimental plots were enumerated by pour plate technique using Aleksandrov media (Aleksandrov et al., 1967). Total microbial activity of KSB (soil dehydrogenase) treated soil samples were estimated by following titrimetry method by Dubey and Maheshwari (2009).

#### **Biochemical constituents of green leaves**

Crop shoots consisting of three leaves and a bud from each replicated plot were subjected to analyse various nutrient uptake and quality parameters.

Total chlorophyll and carotenoids (Wellburn, 1994), sugars (Dubois et al., 1956), nitrogen (AOAC, 1990), proteins (Lowry et al., 1951), amino acids (Moore and Stein, 1948), polyphenols (Dev Choudhury and Goswami, 1983) and catechins (Swain and Hillis, 1959) were estimated.

#### Study of tea bush architecture

Biometric observations of bushes were monitored by assessing plucking surface of the bush, number of active plucking points containing three leaves and a bud, internodal length (average 1 to 3 internodes), fresh and dry weight of leaves, were studied by the method adopted by Balasubramanian et al. (2010).

#### Tea quality constituents

A weight of 2.5 kg of flush shoots containing three leaves and a bud was collected and manufactured the black tea in the crush, tear, curl (CTC) miniature unit, and subsequently theaflavins (TF), thearubigins (TR), highly polymerized substance (Thanraj and Shesadari, 1990), and caffeine (Ronald and Ronald, 1991) were analysed. For determining the quality, organoleptic evaluation (liquor colour, briskness, colour and flavor indexes) of made tea were assessed with professional tea taster's to accomplish the market value realization of the product (Willet, 1987). The flavor index was calculated from the flavor profile, which is the ratio of sum of VFC Group II to that of VFC group I (Obanda and Owuor, 1995).

#### Statistical analysis

The data were analysed using SPSS 14.0 version of statistical software package (SPSS, Inc. Chicago, IL). Data obtained were subjected to analysis of variance (ANOVA) and the significant means were segregated by critical difference (CD) at various levels of significance. The standard error (SE) and coefficient of variance (CV) were also calculated (Gomez and Gomez, 1984).

# **RESULTS AND DISCUSSION**

# Nutrient status of tea plantation soil

The results of tea soil nutrient analysis after imposing various doses of NPK and KSB application was outlined in Table 1. It showed that the tea soils were acidic with a pH range of 4.7 to 5.4. The initial pH measured was 5.4 before imposing treatments. Further, EC ranged between 0.2 and 0.3 dSm<sup>-1</sup> in KSB treated soils. The total organic carbon was increased from 5.2% to varying levels (6.8 to 8.2 %). Moreover, it was significantly increased in soil (8.2%) treated with  $N_{100}P_{100}K_{75}$  + KSB combination. NPK contents were also increased significantly (P < 0.05) in the same treatment. But concentration of calcium was found to be low in N100P100K75 + KSB treatment which was recorded as 164.8 mg/kg soil dry weight. Available magnesium and sodium concentration of soil samples determined between 91 to 156 and 20.8 to 37.3 mg/kg soil dry weight, respectively. Exchangeable K content in soil samples was determined between 97.8 and 141.3 mg/kg soil dry weight which increased in KSB treated plots. Apart from the control plot, soil samples collected from N<sub>100</sub>P<sub>100</sub>K<sub>100</sub> alone treated plots had registered least potassium content (116.2 mg/kg) and this was supported by Ranganathan and Natesan (1987).

The observation and maintenance of high acidic pH in the KSB treated soils revealed the influence of bioinoculant like KSB. The aforementioned experiment results showed that the tea plants are well known to grow better in acidic soil environment (Pandey and Palani, 1996). The increased content of magnesium in soil also had its importance in plant growth. Magnesium is the central metal constituent of chlorophyll molecule that regulates photosynthesis activity and acts as an enzyme activation system (Ma et al., 2005).

# Tea canopy architectural analysis

The treatments where N<sub>100</sub>P<sub>100</sub>K<sub>75</sub> along with KSB combination were given, was the most efficient in enhancing the tea bush canopy architecture when compared to other treatments (Table 2). Plucking surface of the bushes determined between 4892.7 and 8614 cm<sup>2</sup>. It was moderate in plants treated with different dosages of NPK along with KSB and vermicompost combinations which ranged between 6954 and 7935.7 cm<sup>2</sup>. The untreated control bushes registered the plucking surface of an average of 4892.7 cm<sup>2</sup> harvestable shoots. The area of area of plucking surface coincided with the number of plucking points in the bushes due to various treatments. There was a strong positive correlation between the results of plucking points per unit area and the observation of internodal length, leaf moisture and dry matter contents. However, these were found to be superior were plants treated with  $N_{100}P_{100}K_{75}$  along with KSB, followed by  $N_{100}P_{100}K_{100}$  along with KSB and

Treatment details	Soil reaction (pH) #	Electrical conductivity (dSm <sup>-1</sup> )	Organic carbon (%)	Nitrogen (%)	Phosphorus (mg/kg)	Potassium (mg/kg)	Calcium (mg/kg) #	Magnesium (mg/kg)	Sodium (mg/kg)
N <sub>100</sub> P <sub>100</sub> K <sub>100</sub> + KSB	$5.1 \pm 0.2^{e}$	$0.2 \pm 0.04^{a}$	$7.8 \pm 0.1^{b}$	2.2 ± 0.1 <sup>c</sup>	$12.7 \pm 0.7^{9}$	129.6 ± 2.5 <sup>d</sup>	253.7 ± 11.2 <sup>c</sup>	142.8 ± 9.0 <sup>e</sup>	32.3 ± 1.0 <sup>e</sup>
N <sub>100</sub> P <sub>100</sub> K <sub>75</sub> + KSB	$4.7 \pm 0.2^{a}$	$0.3 \pm 0.02^{b}$	$8.2 \pm 0.1^{f}$	$2.7 \pm 0.1^{f}$	13.5 ± 0.9 <sup>h</sup>	141.3 ± 3.6 <sup>f</sup>	164.8 ± 12.2 <sup>a</sup>	156.0 ± 8.0 <sup>f</sup>	37.3 ± 0.9 <sup>g</sup>
N <sub>100</sub> P <sub>100</sub> K <sub>50</sub> + KSB	$4.9 \pm 0.2^{\circ}$	$0.2 \pm 0.04^{a}$	$7.6 \pm 0.1^{e}$	2.5 ± 0.1 <sup>e</sup>	$11.4 \pm 0.6^{f}$	125.6 ± 6.4 <sup>d</sup>	375.3 ± 10.4 <sup>f</sup>	136.3 ± 5.7 <sup>d</sup>	$34.3 \pm 1.1^{f}$
N <sub>100</sub> P <sub>100</sub> K <sub>25</sub> + KSB	$5.4 \pm 0.2^{f}$	$0.2 \pm 0.01^{a}$	$6.8 \pm 0.2^{b}$	$2.2 \pm 0.2^{c}$	$10.6 \pm 0.7^{c}$	120.3 ± 5.0 <sup>c</sup>	384.0 ± 11.4 <sup>g</sup>	124.8 ± 7.1 <sup>c</sup>	31.3 ± 1.0 <sup>d</sup>
N <sub>100</sub> P <sub>100</sub> K <sub>100</sub> + KSB+VC	$4.8 \pm 0.1^{b}$	$0.3 \pm 0.04^{b}$	$7.6 \pm 0.1^{e}$	2.4 ± 0.1 <sup>d</sup>	$10.7 \pm 0.4^{e}$	134.3 ± 6.4 <sup>e</sup>	196.3 ± 13.1 <sup>b</sup>	151.3 ± 10.1 <sup>f</sup>	$29.6 \pm 0.8^{\circ}$
N <sub>50</sub> P <sub>50</sub> K <sub>50</sub> + KSB+VC	$5.3 \pm 0.1^{d}$	$0.2 \pm 0.02^{a}$	$7.2 \pm 0.2^{d}$	$2.2 \pm 0.1^{\circ}$	$10.8 \pm 0.9^{d}$	121.3 ± 3.7 <sup>c</sup>	324.7 ± 14.0 <sup>d</sup>	104.0 ± 11.8 <sup>b</sup>	$24.7 \pm 0.9^{b}$
$N_{100} P_{100} K_{100}$ alone	$4.8 \pm 0.1^{b}$	$0.2 \pm 0.02^{a}$	$7.1 \pm 0.1^{\circ}$	$2.0 \pm 0.1^{b}$	10.6 ± 0.7 <sup>b</sup>	116.2 ± 9.9 <sup>b</sup>	313.3 ± 13.1 <sup>e</sup>	140.7 ± 11.0 <sup>e</sup>	31.8 ± 1.2 <sup>d</sup>
Control $(N_0 P_0 K_0)$	$5.3 \pm 0.2^{d}$	$0.2 \pm 0.02^{a}$	$5.2 \pm 0.2^{a}$	$1.4 \pm 0.2^{a}$	$10.4 \pm 0.7^{a}$	$97.8 \pm 2.3^{a}$	$420.3 \pm 14.4^{h}$	$91.0 \pm 7.6^{a}$	$20.8 \pm 0.9^{a}$
SE ±	0.13	0.02	0.12	0.08	0.59	4.60	8.97	7.56	0.83
CD at <i>P</i> = 0.05	0.39	0.06	0.27	0.19	1.26	9.68	18.86	21.77	1.76

Table 1. Effect of KSB on nutrient status of tea plantation soils.

VC = Vermicompost. KSB = Potassium solubilizing bacteria. Values are mean of three replications. Standard deviations followed by the same letter are not significantly different (P < 0.05) as determined by Duncan's multiple range test. A higher alphabet indicates the improvements obtained due to treatment. # A lower alphabet indicates the improvement obtained due to treatment.

Table 2. Evaluation of KSB on canopy architectural analysis of tea bush.

Treatment details	Plucking surface (cm <sup>2</sup> )	Plucking points (per sq. ft.)	Leaf moisture (%)	Internodal length (cm)	Dry matter (%)
N <sub>100</sub> P <sub>100</sub> K <sub>100</sub> + KSB	$8262.7 \pm 85.9^{f}$	$117.4 \pm 8.0^{f}$	$72.5 \pm 0.5^{e}$	$4.9 \pm 0.2^{e}$	$21.8 \pm 0.8^{e}$
N <sub>100</sub> P <sub>100</sub> K <sub>75</sub> + KSB	8614.0 ± 86.7 <sup>9</sup>	122.9 ± 9.2 <sup>g</sup>	75.1 ± 0.2 <sup>g</sup>	$5.3 \pm 0.1^{f}$	23.6 ± 1.3 <sup>g</sup>
N <sub>10 0</sub> P <sub>100</sub> K <sub>50</sub> + KSB	$7242.0 \pm 89.6^{d}$	113.4 ± 5.4 <sup>e</sup>	$70.4 \pm 1.0^{e}$	$4.8 \pm 0.1^{e}$	$22.7 \pm 1.1^{f}$
N <sub>100</sub> P <sub>100</sub> K <sub>25</sub> + KSB	6942.7 ± 73.5 <sup>d</sup>	$101.4 \pm 6.8^{\circ}$	$68.4 \pm 1.0^{d}$	$3.6 \pm 0.1^{b}$	$21.3 \pm 0.9^{e}$
N <sub>100</sub> P <sub>100</sub> K <sub>100</sub> + KSB + VC	7935.7 ± 75.8 <sup>e</sup>	$116.0 \pm 6.9^{f}$	73.5 ± 1.0 <sup>f</sup>	$4.6 \pm 0.1^{d}$	$22.4 \pm 1.3^{f}$
N <sub>50</sub> P <sub>50</sub> K <sub>50</sub> + KSB + VC	$6954.0 \pm 58.9^{d}$	111.8 ± 7.1 <sup>d</sup>	$68.7 \pm 0.6^{d}$	$4.1 \pm 0.1^{\circ}$	$20.6 \pm 0.6^{d}$
$N_{100} P_{100} K_{100}$ alone	$6681.0 \pm 10.5^{\circ}$	110.3 ± 6.1 <sup>d</sup>	72.7 ± 1.0 <sup>f</sup>	$4.8 \pm 0.1^{e}$	$22.6 \pm 1.2^{f}$
Control ( $N_0 P_0 K_0$ )	4892.7 ± 10.7 <sup>b</sup>	$90.07 \pm 8.4^{b}$	$65.4 \pm 0.8^{\circ}$	$3.6 \pm 0.1^{b}$	$17.5 \pm 0.7^{\circ}$
SE ±	52.78	4.18	0.63	0.08	0.78
CD at P = 0.05	151.9	12.05	1.83	0.24	2.26
CV %	0.93	4.70	1.10	2.31	4.52

Values are mean of three replications. Standard deviations followed by the same letter are not significantly different (P < 0.05) as determined by Duncan's multiple range test. VC = Vermicompost. KSB = Potassium solubilizing bacteria. A higher alphabet indicates the improvements obtained due to treatment.

Treatment details	Biochemical constituents (%)									
	Chlorophyll*	Carotenoid*	Sugar	Nitrogen	Protein	Amino acids	Catechin	polyphenol	Potassium	Phosphorous
N100 P100 K100 + KSB	1.7 ± 0.9 <sup>e</sup>	0.3 ± 0.3 <sup>e</sup>	7.1 ± 0.8 <sup>d</sup>	$6.0 \pm 0.1^{f}$	10.6 ± 0.4 <sup>e</sup>	$1.6 \pm 0.1^{d}$	$19.2 \pm 0.9^{d}$	$27.0 \pm 1.3^{d}$	$2.1 \pm 0.2^{d}$	0.5±0.1°
N100 P100 K75 + KSB	$1.8 \pm 0.2^{f}$	$0.4 \pm 0.1^{f}$	$7.4 \pm 0.6^{f}$	$6.2 \pm 0.3^{e}$	11.4 ± 0.6 <sup>f</sup>	1.6 ± 0.2 <sup>e</sup>	19.5 ± 0.6 <sup>e</sup>	$28.2 \pm 0.3^{e}$	$2.7 \pm 0.4^{f}$	$0.7 \pm 0.2^{f}$
N100 P100 K50 + KSB	$1.4 \pm 0.1^{b}$	0.2 ± 0.2 <sup>c</sup>	$6.2 \pm 0.3^{b}$	5.8 ± 0.1°	10.2 ± 0.7°	1.6 ± 0.1°	18.7 ± 0.7°	26.3 ± 1.1°	1.9 ± 0.1 <sup>b</sup>	$0.5 \pm 0.1^{b}$
N100 P100 K25 + KSB	1.5 ± 0.1°	0.2 ± 0.1 <sup>b</sup>	6.7 ± 0.1°	$5.6 \pm 0.2^{b}$	$10.0 \pm 0.4^{d}$	1.5 ± 0.1 <sup>b</sup>	18.7 ± 0.8°	26.8 ± 1.1°	1.8 ± 0.1ª	0.5 ± 0.1ª
N100 P100 K100 + KSB+VC	1.7 ± 0.1e	0.3 ± 0.1 <sup>d</sup>	7.1 ± 0.1 <sup>d</sup>	$6.0 \pm 0.1^{f}$	$11.4 \pm 0.7^{f}$	1.7 ± 0.3 <sup>f</sup>	18.5 ± 0.3 <sup>b</sup>	28.2 ± 1.1°	2.2 ± 0.2 <sup>e</sup>	$0.6 \pm 0.2^{e}$
N <sub>50</sub> P <sub>50</sub> K <sub>50</sub> + KSB+VC	$1.6 \pm 0.1^{d}$	$0.2 \pm 0.2^{b}$	6.3 ± 0.1 <sup>b</sup>	$5.9 \pm 0.1^{d}$	10.7 ± 1.1°	$1.6 \pm 0.1^{d}$	18.5 ± 0.8 <sup>b</sup>	26.8 ± 0.9°	1.9 ± 0.1 <sup>b</sup>	0.5 ± 0.1°
N100 P100 K100 alone	1.7 ± 0.2 <sup>e</sup>	0.2 ± 0.1°	7.3 ± 0.1°	$5.7 \pm 0.5^{b}$	$10.5 \pm 0.9^{d}$	1.6 ± 0.1°	18.6 ± 0.9°	26.9 ± 0.2°	2.0 ± 0.3°	0.6 ± 0.1 <sup>e</sup>
Control $(N_0 P_0 K_0)$	$0.9 \pm 0.1^{a}$	0.2 ± 0.1ª	5.5 ± 0.1ª	4.7 ± 0.1ª	$8.6 \pm 0.6^{a}$	1.5 ± 0.1ª	17.4 ± 0.9ª	24.5 ± 1.0 <sup>b</sup>	1.8 ± 0.1ª	$0.4 \pm 0.2^{a}$
SE ±	0.10	0.07	0.29	0.12	0.19	0.07	0.60	0.80	0.19	0.04
CD at <i>P</i> = 0.05	0.30	0.21	0.85	0.37	0.57	0.21	1.74	2.30	0.56	0.14

**Table 3.** Estimation of biochemical parameters in tea crop shoots treated with KSB.

Values are mean of three replications. Standard deviations followed by the same letter are not significantly different (P < 0.05) as determined by Duncan's multiple range test. \* mg/g fresh weight of leaves. VC = Vermicompost. KSB = Potassium solubilizing bacteria. A higher alphabet indicates the improvements obtained due to treatment.

 $N_{100}P_{100}K_{100}$  with KSB and vermicompost. The dry matter content was recorded between 22.6 and 23.6% in trial plots and the same was 17.5% in untreated control plots. The application of KSB in soils showed improvement in tea plant growth and also reduced banji content. Similarly, the increasing level on nutrients in soil had increased the yield by improving the yield components like plucking points; plucking surface was reported by Balasubramanian et al. (2010).

#### Biochemical parameters of tea crop shoots

Combined application of KSB formulation with NPK fertilizers resulted in significant increase in chlorophyll, carotenoid and catechins contents when compared to control plot. Chlorophyll is a major green pigment found in green leaves and is undoubtedly determining the photosynthetic efficiency and productivity of plants. Notably K also played an important role in the synthesis of chlorophyll by taking part in various enzyme

activities (Senthurpandian et al. 2008). Since K is found to influence the total chlorophyll and carotenoid contents of the leaves it may also directly and / or indirectly improve crop yield through increased photosynthesis. Similar kind of results was also reported (Jayaganesh et al., 2011). Increase in N, P and K uptake by the plants might be due to the activity of KSB. The results are on par with the findings. (Venkatesan et al. 2005 and 2006). High chlorophyll and carotenoid (1.8 and 0.4 mg/g fresh weight, respectively) contents were recorded by  $N_{100}P_{100}K_{75}$  along with KSB combination (Table 3). Among the nutrient parameters, P and K contents had shown tremendous increase in the case of  $N_{100}P_{100}K_{75}$  with KSB treatment (2.7 and 0.7%, respectively) and N<sub>100</sub>P<sub>100</sub>K<sub>100</sub> with KSB and Vermicompost treated (2.2 and 0.6%, respectively) plots whereas, the N content was recorded higher in  $N_{100}P_{100}K_{100}$  with KSB and Vermicompost, followed by N<sub>100</sub>P<sub>100</sub>K<sub>100</sub> with KSB alone (6.2 and 6.0%) treated plots. The contents of protein, catechin, and polyphenol (11.4, 28.2 and 19.5%, respectively) contents were registered in the plants treated with  $N_{100}P_{100}K_{75}$  with KSB when compared to other treatments. High amino-acid content was observed in the plots wherever KSB was incorporated and this was substantiated with the findings of Ruan et al. (1998). The significant increase in sugar content was observed in the treatment with  $N_{100}P_{100}K_{75}$  (7.4%) followed by  $N_{100}P_{100}K_{100}$  with KSB and Vermicompost (7.1%). Less sugar content was registered in control plots (5.5%).

An integrated application of inorganic manures with KSB that solubilize them might provide faster and continuous supply of K for higher plant growth. Similarly, the use of plant growth promoting rhizobacteria (PGPR) including phosphate solubilizing bacteria (PSB) and KSB as biofertilizers was suggested as a sustainable solution to improve plant nutrient and production. The importance of potassium in the yield potential in tea has been reported earlier by Venkatesan et al.

Treatment details	Green leaf yield (kg made tea ha <sup>-1</sup> )	Productivity index (PI)	Banji content (%) #
N 100 P100 K100 + KSB	$2624.3 \pm 65.2^{e}$	$1.2 \pm 0.1^{e}$	$40.0 \pm 1.0^{b}$
N <sub>100</sub> P <sub>100</sub> K <sub>75</sub> + KSB	2957.0 ± 75.7 <sup>9</sup>	1.6 ± 0.1 <sup>g</sup>	$36.0 \pm 2.0^{a}$
N <sub>100</sub> P <sub>100</sub> K <sub>50</sub> + KSB	$2524.7 \pm 52.8^{d}$	$1.3 \pm 0.1^{f}$	$41.7 \pm 0.6^{b}$
N <sub>100</sub> P <sub>100</sub> K <sub>25</sub> + KSB	2358.0 ± 43.1 <sup>b</sup>	$0.9 \pm 0.1^{b}$	$48.3 \pm 1.5^{\circ}$
N <sub>100</sub> P <sub>100</sub> K <sub>100</sub> + KSB+VC	2562.7±38.9 <sup>d</sup>	$1.2 \pm 0.2^{c}$	$50.0 \pm 1.0^{d}$
N <sub>50</sub> P <sub>50</sub> K <sub>50</sub> + KSB+VC	2451.3 ± 41.1 <sup>c</sup>	$1.2 \pm 0.1^{\circ}$	55.3 ± 4.1 <sup>e</sup>
N <sub>100</sub> P <sub>100</sub> K <sub>100</sub> alone	$2739.3 \pm 59.7^{f}$	$1.5 \pm 0.1^{d}$	$53.7 \pm 5.5^{f}$
Control $(N_0 P_0 K_0)$	1904.3 ± 83.1 <sup>a</sup>	0.7±0.2 <sup>a</sup>	$63.0 \pm 2.0^{9}$
SE ±	41.39	0.05	2.22
CD at <i>P</i> =0.05	119.14	0.16	6.41
CV %	2.06	5.72	5.45

Table 4. Evaluation of KSB on growth parameters of tea plants.

Values are mean of three replications. Standard deviations followed by the same letter are not significantly different (P < 0.05) as determined by Duncan's multiple range test. VC = Vermicompost. KSB = Potassium solubilizing bacteria. A higher alphabet indicates the improvements obtained due to treatment. # A lower alphabet indicates the improvement obtained due to treatment.

(2005). Significant positive effect was observed by (Singh et al., 2010) for quality-related parameters like sugars, amino acids, proteins, polyphenols and caffeine by the inoculation of plant growth promoting fungus. Amino acids are responsible for flavor profile of made tea, it was noticed in Chinese tea gardens (Thomas et al., 2009). Lowest content of amino acids in control plot could be due to the reduction of inorganic K fertilizer or lack of indigenous K solubilizers (Venkatesan and Ganapathy, 2004).

# Evaluation of potassium solubilizing bacteria on growth parameters and enumeration of KSB and dehydrogenase activity in tea soil

Green leaf yield was recorded at every harvesting period and presented as made tea yield/ha (Table 4). Application of indigenous KSB formulation with various doses of potash fertilizers with N and P had enhanced green leaf yield and productivity index (PI) significantly. The higher yield (2957 kg made tea/ha) was observed in  $N_{100}P_{100}K_{75}$  with KSB treatment followed by  $N_{100}P_{100}K_{100}$ (2739 kg made tea/ha) treatment. A drastic reduction in yield potential was noticed in N100P100K25 with KSB formulation and control blocks (2358 and 1904 kg made tea/ha, respectively). Significant improvement of green leaf yield was observed in the experimental plots which may be due to proper fixation of nutrients in the soil through solubilization by producing organic acids by KSB strains. Likewise Sheng et al. (2002) reported that KSB namely Bacillus mucilaginosus is able to solubilize inorganic source of K like muriate of potash and sulphate of potash by means of production of organic acids in order to improve the yield. The plants which receive the highest level of K gave the highest yield in all the years (Natesan et al. 1984). PI in our experiment was positively correlated with the yield of tea plants applied with highest level of K along with KSB.As the yield increases, there was a decrease in percentage of banji content in the plucking surface during harvest.

The highest enzyme activity (41.23 mg of  $CO_2$  evolutions) and population (3.2 × 10<sup>6</sup> cfu/g of soil) was observed in N<sub>100</sub>P<sub>100</sub>K<sub>75</sub> with KSB followed by N<sub>100</sub>P<sub>100</sub>K<sub>100</sub> with KSB combinations and least microbial activity was noticed in control plot (Figures 1 and 2). Significant improvement of dehydrogenase activity and KSB population was observed in KSB incorporated plots due to their native origin of environment and better establishment.

Higher green leaf yield and bush health in terms of number of plucking points, internodal length, and physiological and biochemical constituents might be due to the action of KSB by solubilizing K content in the soils. These parameters were further coincided with higher population of KSB and dehydrogenase activity (Figures 1 and 2). Least KSB population was recorded in control and other treatments due to either lesser/native state of KSB or lack of reduced level of inorganic fertilizers. The population density of KSB was significantly coincided with soil nutrients like total organic carbon, total N and available P contents. Similar observations were reported in tea soils (Bagyalakshmi et al., 2009).

# Quality parameters of black tea

Influence of KSB on TF, TR, highly polymerized substances (HPS), total liquor colour (TLC) and caffeine contents were presented in Table 5. Significant increase in the level of TF and TR contents was observed in  $N_{100}P_{100}K_{75}$  along with KSB treatment (1.3 and 9.9%) and  $N_{100}P_{100}K_{100}$  with KSB and Vermicompost treated blocks (1.2 and 9.5%, respectively). Least contents were observed in the control (0.8% TF and 7.6% TR) plots, where no KSB was incorporated. TF and TR contents



**Figure 1.** Enumeration of KSB Population in tea soil. VC, Vermicompost; KSB, Potassium solubilizing bacteria.



Figure 2. Evaluation of dehydrogenase activity in tea soil. VC, Vermicompost; KSB, Potassium solubilizing bacteria.

were found to be lower in control blocks. In the case of HPS status, significant variation was observed among the treatments studied. There was a drastic improvement in caffeine content in KSB amended plots. Similarly, higher caffeine content (3.9, 2.9 and 2.9%) was observed in the treatment of N<sub>100</sub>P<sub>100</sub>K<sub>75</sub> with KSB, N<sub>100</sub>P<sub>100</sub>K<sub>100</sub> + KSB +Vermicompost and N<sub>100</sub>P<sub>100</sub>K<sub>100</sub> alone, respectively.

Colour, flavour and briskness indexes were significantly higher in the same order of treatments.

The chemical basis of the quality of black tea indicated that TF and TR are the two important factors determining the liquoring characteristics of tea. When observing made tea parameters in order to evaluate the quality attributes, TF and TR ratio was higher with KSB applied plants Table 5. Analysis of quality parameters of black tea of KSB treated tea leaves.

	Made tea characteristics (%)								
Treatment details	Theaflavin	Thearubigin	Highly polymerized Substances	Total liquor colour	Caffeine	Briskness Index	Colour Index	Flavour Index	
N <sub>100</sub> P <sub>100</sub> K <sub>100</sub> + KSB	$1.2 \pm 0.2^{d}$	$9.1 \pm 0.4^{d}$	$9.8 \pm 0.2^{e}$	$3.6 \pm 0.1^{d}$	2.9 ± 0.1 <sup>e</sup>	10.0 ± 0.1 <sup>d</sup>	35.5 ± 1.2 <sup>e</sup>	$23.1 \pm 0.1^{\circ}$	
N <sub>100</sub> P <sub>100</sub> K <sub>75</sub> + KSB	1.3 ±0.1 <sup>d</sup>	$9.9 \pm 0.8^{h}$	$10.5 \pm 0.6^{f}$	3.8 ± 0.1 <sup>e</sup>	$3.9 \pm 0.1^{f}$	$14.0 \pm 0.1^{e}$	$36.6 \pm 0.6^{f}$	$24.1 \pm 0.1^{h}$	
N <sub>100</sub> P <sub>100</sub> K <sub>50</sub> + KSB	1.1 ± 0.1 <sup>c</sup>	$9.1 \pm 0.5^{f}$	$10.2 \pm 0.8^{f}$	$3.3 \pm 0.1^{\circ}$	$2.6 \pm 0.2^{d}$	11.2 ± 0.1 <sup>c</sup>	33.2 ± 1.2 <sup>b</sup>	$23.8 \pm 0.2^{9}$	
N <sub>100</sub> P <sub>100</sub> K <sub>25</sub> + KSB	$1.0 \pm 0.2^{b}$	$9.1 \pm 0.0^{e}$	$8.8 \pm 0.1^{\circ}$	$3.2 \pm 0.1^{b}$	$2.4 \pm 0.2^{b}$	11.9 ± 0.1 <sup>b</sup>	$33.8 \pm 0.3^{\circ}$	$22.3 \pm 0.2^{d}$	
N <sub>100</sub> P <sub>100</sub> K <sub>100</sub> + KSB + VC	1.2 ± 0.1 <sup>c</sup>	$9.5 \pm 0.5^{9}$	9.8 ± 0.1 <sup>e</sup>	$3.6 \pm 0.2^{d}$	2.8 ± 0.1 <sup>c</sup>	$10.1 \pm 0.2^{d}$	$34.1 \pm 0.8^{d}$	$23.4 \pm 0.5^{e}$	
N <sub>50</sub> P <sub>50</sub> K <sub>50</sub> + KSB+VC	$1.1 \pm 0.1^{b}$	$8.9 \pm 0.9^{\circ}$	$9.2 \pm 0.2^{d}$	$3.1 \pm 0.2^{b}$	2.5 ± 0.1 <sup>b</sup>	11.2 ± 0.1 <sup>c</sup>	$33.8 \pm 0.2^{\circ}$	$21.9 \pm 0.9^{b}$	
$N_{100}P_{100}K_{100}$ alone	$0.9 \pm 0.1^{a}$	$8.4 \pm 0.6^{b}$	$8.5 \pm 0.1^{b}$	$3.2 \pm 0.1^{\circ}$	2.9 ± 0.1 <sup>e</sup>	$10.4 \pm 0.1^{d}$	$34.7 \pm 0.3^{d}$	$23.6 \pm 0.5^{f}$	
Control (N <sub>0</sub> P <sub>0</sub> K <sub>0</sub> )	$0.8 \pm 0.2^{a}$	$7.6 \pm 0.3^{a}$	$7.2 \pm 0.1^{a}$	$2.3 \pm 0.2^{a}$	2.1 ± 0.1 <sup>a</sup>	$12.9 \pm 0.1^{a}$	$30.7 \pm 0.7^{a}$	$20.9 \pm 0.3^{a}$	
SE ±	0.06	0.48	0.24	0.12	0.08	0.15	0.43	0.26	
CD at P =0.05	0.20	1.39	0.71	0.35	0.25	0.45	1.27	0.77	

Values are mean of three replications. Standard deviations followed by the same letter are not significantly different (P < 0.05) as determined by Duncan's multiple range test. VC = Vermicompost.KSB = Potassium solubilizing bacteria. A higher alphabet indicates the improvements obtained due to treatment.

(Table 5). This may be due to the K source, and is the most important nutrient to increase the TF value (Venkatesan et al., 2006). The optimum TF/TR ratio has been reported to be around 1:10 (ideal ratio) and any wide deviation in the ratio would lead to quality deterioration. The higher the water extracts content, the higher the quality. All the treatments had above 32%; the limiting factors which influence the cuppage of black tea. The increase in TF could be due to the increase in polyphenols at the optimum ratio, which is the precursor of TF and TR. In case of other parameters, HPS, TLC, polyphenols (PP), catechins (Cat), amino acids (AA) and reducing sugars (RS) contributed to the overall performance of the plants treated with N100P100K75 + KSB. More polyphenols is found in the tea shoots undoubtedly; the quality of the manufactured tea will be greater (Venkatesan et al., 2005) because polyphenols are so important and responsible for all the biochemical reactions which contribute to make a good cup of tea.

In made tea, flavor index (FI) was estimated and observed was the significant increase due to the application of KSB in tea soil, and tea aroma consists of groups of flavor compounds. Higher amount of crude fiber in control and vermicompost treated plots was due to the decreased availability of K in soil. This leads to the accumulation of carbohydrates and cellulose. Our results are on par with the findings of Venkatesan and Ebert (2005).

# Conclusion

Hence, with the existing economics crisis for tea growers, it is necessary to look for KSB biofertilizers integrated with reduced level of K fertilizers which in turn provide high crop yield with low cost. The present research findings provide the information that balanced nutrient application to tea plants and sustain yield were achieved with 75% of K fertilizers along with indigenous KSB.

Moreover, quality of black tea may be improved

by the application of 75% K along with KSB. Thus, the application of KSB can be recommended as a sustainable way to increase yield potential and maintain the soil health in the tea ecosystem.

# ACKNOWLEDGEMENTS

The researchers are thankful to the Principal and the Management, K.S.R. College of Educational Trust, Thiruchengode, Tamil Nadu, and Dr P. Mohankumar, Director and Head of Tea Technology Division, UPASI Tea Research Institute, Valparai, Tamil Nadu, India, are gratefully acknowledged for their encouragement and support for conducting the above the experiment.

#### REFERENCES

Aleksandrov VG, Blagodyr RN, Ilev IP (1967). Liberation of phosphoric acid from apatite by silicate bacteria. Mikrobiol.

Z. 29:111-114.

- AOAC (1990). Official Methods of Analysis of the Association of Official Analytical Chemist. (ed), Helrich, K. 15th Edition, AOAC Inc., Gaithersburg, USA., pp. 1-2.
- Bagyalakshmi B, Nithya B, Ponmurugan P (2009). Studies on potassium solubilizing bacteria from Southern Indian tea soils, in: P.Ponmurugan, B.Nithya (Eds.), Guidelines for Entrepreneurship development programme to biotechnology graduates, Excel India Publishers, New Delhi, pp. 71–76.
- Balasubramanian S, Netto LA, Parathiraj S (2010). Unique graft combination of tea, Cr-6017 / UPASI-9. Curr. Sci. 98:1508-1517.
   Bhargava BS, Raghupathi HB (2001). Analysis of plantmaterials for macro and micronutrients. In H.L.S Tandon. (Ed.), Methods of Analysis of Soils, Plants, Waters and Fertilizer. New Delhi, India: Fertilizer Association and Consultation Organization. pp. 49-82.
- Cakmak I (2005). The role of potassium in alleviating detrimental effects of abiotic stresses in plants J. Plant Nutr. Soil Sci. 168:521–530.
- Dev Choudhury MN, Goswami MR (1983). A rapid method for determination of total polyphenolic matter in tea (*Camellia sinesis* L.). Two and a Bud 30:59-61.
- Dubey RC, Maheshwari DK (2009). "Practical Microbiology". S. Chand and Company LTD, New Delhi, pp. 192- 193.
- Dubois M, Gilles KA, Hamilton JK, Roberts PA, Smith F (1956). Colorimetric method for determination of sugars and relatedsubstances. Determination of concentration of pure sugar solution. Ann. Chem. 28:350–360.
- Gomez KA, Gomez AA (1984). Statistical procedures for agricultural research.2nd edn. John Wiley and Sons Inc., New York, pp. 95-109
- Han HS, Lee KD (2005). Phosphate and potassium solubilizing bacteria effect on mineral uptake, soil availability and growth of egg plant. Res. J. Agric. Biol. Sci. 1(2):176-180.
- Jackson ML. (1973). Soil chemical analysis. Prentice Hall of India Pvt. Ltd. New Delhi, pp. 498-516
- Jayaganesh S, Venkatesan S, Senthurpandian VK (2011) Impact of sources and doses of magnesium fertilizer on biochemical constituents and quality parameters black tea. Food Chem. 90:535-539.
- Liang CAE, Zhi-Xiu Zhou AE, Ya-Jun Y (2007).Genetic improvement and breeding of tea plant (Camellia sinensis) in China: from individual selection to hybridization and molecular breeding. Euphytica 154:239–248.
- Lowry OH, Rosenbrough NJ, Farr AL, Randall RJ (1951). Protein measurements with the folin phenol reagent. J Biol. Biochem. 193:265–275.
- Ma L, Ruan JY, Han W, Shi Y (2005). Effect of magnesium nutrition of the formation and transport of free aminoacids in tea plants. Proceedings of International Symposium on Innovation in Tea science and sustainable development in tea industry,Nov.10-15,Tea Research Institute, Chineese academy of Agricultural Sciences, Hangzhou,China, pp. 151-158.
- Mandal AKA, Kumar RR, Thomas J (2007). An overview of PGPR trials in UPASI TRF. Planters Chron. 103:12-16.
- Moore S, Stein WH (1948). In S. P. Colowick and N. D. Kaplan (Eds.) New York: Academic press. Methods in enzyme, p. 468
- Murphy J, Riley JP (1962). A modified single solution method for determination of phosphate in natural waters. Anal. Chim. Acta 27:31-36.
- Natesan S, Subrahmanya Bhat S, Ranganathan V (1984). Agronomic and cultural factors interacting with potassium manuring in tea. In: Proc. 5thAnnu. Symp. Plantation Crops, Kasaragod, (India). PLACROSYM Standing Committee, Indian Society of Plantation Crops Kasaragod Kerala India.
- Obanda M, Owuor PO (1995) Impact of shoot maturity on chlorophyll content, composition of volatile flavour compounds and plain black tea quality parameters of clonal leaf. J. Sci. Food Agric., 69:529-534.
- Pandey A, Palni LMS (1996). The rhizosphere effect of tea on soil microbes in a Himalayan monsoonal location. Biol. Fertil. Soils 21(3):131-137.
- Ponmurugan P, Baby UI (2007). Evaluation of fungicides and biocontrol agents against Phomopsis canker of tea under field condition. Aust. J. Plant Pathol. 36:68–72.

Ponmurugan P, Gopi C (2006). In vitro production of growth regulators

and phosphate activity by phosphate solubilizing bacteria. Afr. J. Biotechnol. 5:348-350.

- Ramarethinam S, Chandra K (2006). Studies on the effect of potash solubilizing bacteria *Frateuria aurantia* (Symbion-K- Liquid Formulation) on Brinjal (Solanum melongena L) growth and yield. Pestology 11:35-39.
- Ranganathan V, Natesan S (1987). Potassium availability in tea growing soils of South India. Soil testing, plant analysis and fertilizer evaluation for potassium.Publ.No.415: New Delhi, India, p. 368.
- Rawlings DE (2002). Heavy metal mining using microbes. Annu. Rev. Microbiol. 56:65–91.
- Ronald SK, Ronald S (1991) Beverages and chocolate. In Pearson's composition and analysis of foods, 9<sup>th</sup> ed., Longman Scientific & Technical publishers, England, pp. 356 362.
- Ruan J, Hardter R (2001). Productivity and quality response of tea to balanced nutrient management – Examples from China tea gardens, Int. Potash Inst. pp. 118-122.
- Ruan J, Wu X, Hardter R (1998). Effect of potassium, magnesium and sulphur applied in different forms of fertilizers on free amino acids content in leaves of tea (*Camellia sinensis* L). J. Sci. Food Agric. 76:107–113.
- Sedaghathoor S, Torkashv AM, Hashemabadi D, Kaviani B (2009) Yield and quality response of tea plant to fertilizers. Afr. J. Agric. Res. 4(6):568-570.
- Senthurpandian VK, Jayganesh S, Vankatesan S (2008). Influence of excess magnesium on aminoacid synthesis pathway and biochemical parameters of tea leaves. News lett. UPASI Tea Res. Found. 18:4-5.
- Sharma VS, Sathyanarayana N (1990). Productivity index of tea (*Camellia* L.sp.). Proceedings of the national symposium of new trends in tea. Crop improvement of perennial species, (NSTT90), Rubber research institute of India, Kottayam, Kerala, India, pp. 16-16.
- Sheng XF, He LY, Haung WY (2002). The condition of releasing potassium by a silicate dissolving bacterial strain NBT. Agric. Sci. China 1:662-666.
- Singh S, Pandey A, Kumar B, Palni LMS (2010). Enhancement in growth and quality parameters of tea [*Camellia sinensis* (L.) O. Kuntze] through inoculation with arbuscular mycorrhizal fungi in an acid soil. Biol. Fertil. Soils 46:427–433.
- Swain T, Hillis WE (1959). The phenolic constituents of *Prunus domestica*. I.The quantitative analysis of phenolic constituents. J. Sci. Food Agric. 10:63-68.
- Thanaraj SNS, Seshadri R (1990). Inflence of polyphenol oxidase activity and polyphenol content of tea shoot on quality of black tea. J. Sci. Food Agric. 51:57 69.
- Thomas J, Mandal AKA, Rajkumar R, Chordia A (2009). Role of biologically active aminoacid formulations on quality and crop productivity of tea (Camellia sp.). Intell. J. Agric. Res. 4(7):228-236.
- Walkley A, Black CA (1934). An examination of the Degtjareff method for determining soil organic matter and proposed modification of chromic valid titration method. Soil Sci. 37:29-38.
- Wellburn R (1994). The spectral determination of chlorophylls a and b, as well as total carotenoids, using various solvents with spectrophotometers of different resolution. J. Plant Physiol. 144:307– 313.
- Venkatesan S, Ebert G (2005). Influence of SOP and MOP on productivity and quality of tea. Proceedings of International Symposium on Innovation in Tea science and sustainable Development in Tea industry, Nov.10-15, Tea Research Institute, Chinese Academy of Agricultural Sciences, Hangzhou, China, pp. 107-117.
- Venkatesan S, Murugesan S, Senthurpandian VK, Ganapathy MNK (2005). Impact of sources and doses of potassium on biochemical parameters of tea. Food Chem. 90:535-539.
- Venkatesan S, Senthurpandian VK, Murugesan S, Maibaum W, Ganapathy MNK (2006). Quality standards of CTC black teas as influenced by sources of potassium fertilizer. J. Sci. Food Agric. 86:799-803.
- Venkatesan, S, Ganapathy MNK (2004). Impact of nitrogen and potassium fertilizer application on quality of CTC teas. Food Chem. 84:325-328.
- Verma DP, Palani N (1997). Manuring of tea in south India (revised recommendations). In: Handbook of tea culture. Valparai, Tamil

Nadu: UPASI Tea Res. Inst. 11 33. Willet RE (1987). Gas chromatography. John Wiley and Sons. New York, USA., p. 160.