

Full Length Research Paper

Nutrient, benthic algae relationships in the Sinop Karagol-Aksaz Marsh, Turkey

Ridvan Erdal Sivaci

Department of Biology, Faculty of Adiyaman Art and Science, Adiyaman University, Adiyaman, Turkey.

Accepted 3 April, 2013

Investigation of nutrient and composition of benthic algae from chosen random sampling station on Karagöl-Aksaz Marsh were researched between November 2005 and November 2006. In addition, some physical and chemical parameters (soluble reactive phosphorus, total phosphorus, NH_4 , NO_3 , Cl, chlorophyll-*a* (Chl-*a*), carotenoid, pH, O_2) were measured. Benthic algae diversity was calculated by using the Shannon-Weaver (H') and evenness (J) index. (H') indices varied from 1.03 to 1.48, and (J) from 0.42 to 0.67. Cluster analysis produced the three major groups, A-B swamp-wet and C swamp-dry, and occurs at the highest level of similarity. The analyses were recognized at the 0.4 dissimilarity level. A total of 55 benthic algae taxa were identified in Kargöl-Aksaz Marsh lake. Although the species of genus *Gomphosphaeria* spp., *Dictyosphaerium* spp., *Navicula* spp., and *Gomphonema* spp. constituted the main composition of the benthic flora, the species belonging to genus *Sellaphora* spp., *Frustulia* spp., *Pinnularia* spp. and *Stauroneis* spp. were found to be less numerous. The flora as a whole increased in March, April and May (swamp-wet) and decreased in July and August (swamp-dry).

Key words: Marsh, benthic algae, cluster, diversity, nutrient.

INTRODUCTION

Marsh is a type of wetland which is an important ecological unit. Hydrological cycle of a Marsh is variable and distinguishes itself from other ecosystems. It can vanish from reconstruction by geology, meteorological movements and human impact (Fleeger et al., 2008). Marsh landscapes include unvegetated mudflats, a creek-Marsh ecotone between vegetated and non-vegetated sediments and a densely vegetated high Marsh platform. The gradient of elevation/inundation in Marshes vary with biotic factors (inundation, aerial exposure, flow, light, and sediment chemistry) (Menéndez and Sanmarti, 2007). Soluble organic matter in Marsh is a considerable energy source for microorganisms (Newell and Porter, 2000).

Nutrient load imported to Marsh areas has increased annually with the increase of waste from human activity, agricultural runoff and industrial production, especially drought. Water is the essential element that dominates in Marsh habitats. Marshes and other wetland ecosystems undergo cyclical periods of flooding, drought, and deterioration. Water levels in Marsh environments are continuously fluctuating and individual Marshes have unique water level patterns. Generally, primer production in the Marsh has three components; benthic algae, planktonic algae and angiosperms. It has long been recognized that algae distributions in Marshes are strongly affected by water chemistry and different water cyclical periods. Seasonal succession of algae is



Figure 1. The location of Karagöl-Aksaz Marsh in Turkey.

conditioned by the joint action of biotic and abiotic factors of environment in Marsh (Danilov, 2002). Therefore, many algal including diatoms species are indicators of Marsh conditions reflecting the influence of hydrological cycle. Diatoms constitute an important group of algae. The cell wall is silicified to form a frustule, comprising two valves. They have been recorded and classified for over 200 years (Horton et al., 2006). One of the important factors affecting the distribution algae (especially diatoms) of the water chemistry is known to be a long time. Therefore, algae are precious indicators of water quality characteristics including pH, nutrient status, and salinity. As an advantage over other bioindicators, they have shorter generation times, showing a rapid response to environmental changes and therefore become early warning indicators of changes in nutrient status (Smol and Stoermer, 2010).

MATERIALS AND METHODS

Description of the study site

Anatolian region for more than the total size of 2,000,000 ha (2,155,045 ha), 135 units of international importance are wetlands. These fields are required to enter into this important wetland in the Sinop region, which is our study area is swampy Karagöl-Aksaz. Karagöl-Aksaz Marshes in the Sinop Black Sea shore line which exists at the interface of freshwater through inflow salt water entering the Karasu stream from the Black Sea. The Marsh surrounds Aksaz lake, a shallow tidal lake that has developed in effect of a sandy barrier Black Sea beach since 4,000 or 5,000 years ago. The crest of the barrier beach is approximately +2 m above the elevation of the Marsh, which is drained by means of a culvert through the barrier (Özener, 1998) (Figure 1). This study aims to determine the benthic algae composition and relations

amongst nutrient, water regime and diversity of Karagöl-Aksaz lake. Measurements were taken from January, 2005 to March, 2006 in sampling periods set at random according to the hydro-period changes. Sediment samples were collected from random stations by means of a glass pipe 11 mm in diameter and 1 m in length. The pipe was lowered by hand to the surface of the sediment, while one end was closed with the thumb. The pipe was then moved in a circular direction on the surface and the thumb was slightly loosened to suck the sediment into the pipe. The collected sediment samples were transferred into plastic bottles and taken to the laboratory for further examination. The sediment samples were put into petri dishes and allowed to settle for 4 to 6 h. The supernatant was removed from the petri dishes by micropipetting and cover glasses were placed over the sediments. After 24 h, the cover glasses were carefully taken and washed into beakers. All benthic algae were identified and counted before fixation. Thereafter, diatoms were prepared following standard techniques: carbonate dissolution by HCl followed by oxidation of organic matter using H₂O₂ and repeated washing of the resultant diatom frustule in demineralized water (Battarbee, 1986). At least 400 diatom valves were counted on each slide along randomly chosen transects. Identification and enumeration was made using an Olympus Vano microscope with brightfield optics at a magnification of 1600×. The number of total diatoms (per cm²) was calculated by using Round's methodology (Round, 1953) ($p \leq 0.05$) (Table 1). Then, all benthic algae were converted to relative abundances (that is, proportions of species to the total for each sample). Taxonomic identifications of species were made following Patrick and Reimer (1966, 1975), Komárek and Anagnostidis (2005) and Krammer and Lange-Bertalot (1986, 1988, 1991a, 1999a,b).

Environmental variables

Dissolved oxygen (DO) concentration, water temperature (YSI 55B Model), conductivity and pH (Cyber Scan 510) were measured in the field. Water for chemical analyses was stored under cold dark conditions in acid-washed 1-L Pyrex bottles, following filtration through glass fiber cartridges (GF/C) filters for ammonium, nitrate

and soluble reactive phosphorus (SRP) determinations. Unfiltered water was used for other variables. All analyses were completed within 18 h of sampling. Alkalinity was determined by titration with HCl using BDH 4.5 indicator. SRP, total soluble phosphorus (TSP), total phosphorus (TP), silicate (SiO_3), chlorine (Cl^-), calcium (Ca^{2+}), sulphate (SO_4^{2-}) and ammonium (NH_4^+), and were determined according to Mackereth et al. (1978) to a precision of $\pm 4\%$. Nitrate was determined by reduction to nitrite on spongy cadmium and subsequent diazotization to a pink dye, determined spectrophotometrically, to a precision of $\pm 3\%$. For determination of chlorophyll-a (Chl-a) concentration, water volumes of 500 mL^{-1} were filtered immediately through GF/C filters after the addition of 0.2 mL^{-1} saturated MgCO_3 . Filters were extracted in cold 90% acetone for 18 to 24 h. Following absorption measurements, the equations of Tailing and Driver (1961) were used to determine Chl-a concentrations corrected for phaeopigments.

Diversity indices

A comparative analysis of these diatom communities was performed by means of diversity measures. We calculated diatom diversity using the Shannon-Weaver (H') and evenness (J) index:

$$H' = -\sum_{i=1}^S P_i \log_2 P_i \quad (P_i = N_i / N) ; J = H' / \log_2 S$$

Where $P_i = N_i / N$ relative abundance for each species; S = total number of species, N_i = number of individuals of a species in sample; N = total number of individuals of all species in the sample (Shannon Weaver, 1949; Pielou, 1966).

Cluster analysis

Investigation of similarities in benthic algae compositions among the samples was performed by means of cluster analysis based on the Euclidean distance calculated from square root transformed relative abundance (an alternative method of calculating the Hellinger distance as proposed by Legendre and Gallagher (2001)). The complete linkage method was preferred to determine the groups of samples that are most similar. Computations were performed using Minitab v 13.1.

RESULTS AND DISCUSSION

During the sampling period, we determined total of 55 taxa. Predominantly *Gomphosphaeria*, *Dictyosphaerium*, *Nitzschia*, *Navicula* and *Gomphonema* often have been observed but *Bacillariophyta* were dominant in terms of species number during the study period. Cyanobacteria, especially *Gomphosphaeria* sp. and *Chlorophyta*, *Dictyosphaerium granulatum* became dominant in Karagöl Marsh with a rapid increase in late spring. Komárek and Anagnostidis (2005) reported that *Gomphosphaeria* and *Dictyosphaerium* are the majority of species free living in the methaphyton of the littoral of lakes and in swamps and pools, among other algae and water plants (one in salinic swamps), usually with limited areas of distribution. As mentioned in the report that such a limited amount of space has been determined at very high rates (Figure 3). The reason for this in our opinion is

a very high phosphate and nitrate was caused by the rapid change of water regime (Table 2). Because of the hydro-period and ionic composition of the Marsh, system affects benthic algae flora both directly via their influence on physiological processes and indirectly via their influence on biogeochemical cycles. A combination of high water, N and P availability over the sediment surface increased the density of meiofauna, cyanobacteria, diatoms and also probably bacteria in autumn during the decay after leaf fall, by enhancing macrophyte decomposition.

Our results indicated that some major environmental factors such as hydro-period, phosphate, nitrate and temperature control the benthic algae assemblages. Thomas et al. (2006) indicated that changes in the hydrologic regime (that is, duration and timing of flooding, water depth) can greatly influence periphyton community structure and function. During the dry season in short-hydroperiod Marshes, periphyton communities are dormant and unproductive. Following re-flooding, periphyton recovers within days, but not before large quantities of nutrients has been released back into the water column. In stagnant situations, these nutrients may be re-sequestered by the community upon recovery but if water is flowing, the released nutrients can affect downstream communities (Gottlieb et al., 2006). One of the most important factors affecting the development of benthic algae was nutrients and hydro-period. We showed that periphyton communities can quickly adapt in composition and structure if exposed to alternating hydrologic regimes. These fast responses to environmental changes strongly advocates for the use of periphyton as an indicator for environmental monitoring in Aksaz-Karagöl Marsh.

The pattern of Shannon-Weaver (H') has shown that the diversity is greater in the period of elevated water than in period of decreased water. This diversity may be related to greater environmental variability (tidal fluctuations, nutrient and temperature). Shannon-Weaver (H') indices varied from 1.03 to 1.48, and evenness from 0.42 to 0.67, giving averages as 1.21 and 0.52, respectively ($p \leq 0.05$). The highest diversity index was recorded in March (swamp-wet) and the lowest in July and August (swamp-dry). The diversity index values tend to increase during swamp-wet (March, April and May) and certain species, especially *Gomphosphaeria* spp. and *D. granulatum* are dominant.

The dendrogram in Figure 2 shows sample affinities based on the root transformed relative abundance of a total of 36 benthic algae species using the Euclidean measure of similarity. The dashed line drawn at the arbitrary similarity level of 40% clearly defines threemajor clusters. Three major clusters, diatom groups A-B swamp-wet and C swamp-dry, occur at the highest level of similarity. A useful clustering can also be recognized at the 0.4 dissimilarity level and this has been used to distinguish each of the benthic algae groups shown in

Table 1. Benthic algae flora of Karagöl-Aksaz Marsh.

Division	Class	Species
Chlorophyta	Trebouxiophyceae	<i>Chlorella vulgaris</i> Beijerinck
		<i>Acutodesmus acuminatus</i> (Lagerheim) Tsarenko
		<i>Desmodesmus opoliensis</i> (P. G. Richter) E.H.Hegewald
		<i>D. granulatum</i> Hindák
		<i>Kirchneriella incurvata</i> Belcher and Swale
		<i>Micromelum minutum</i> (Nägeli) Komárková-legnerová
		<i>Micromelum pusillum</i> (Printz) Komárková-L.
		<i>Pediastrum</i> spp.
		<i>Scenedesmus Bijuga</i> (Turpin) Lagerheim
		<i>Scenedesmus obliquus</i> (Turpin) Kützing
		<i>Scenedesmus</i> spp.
		<i>Tetraedron</i> spp.
		<i>Chroococcus</i> spp.
		<i>Gomphosphaeria</i> spp.
		Cyanobacteria
<i>Phormidium limosum</i> (Dillwyn) Silva		
<i>Pseudanabaena limnetica</i> (Lemmermann) Komárek		
<i>Euglena acus</i> Ehrenberg		
Euglanzoa	Euglenophyceae	<i>Lepocinclis acus</i> (O. F. Müller) Marin and Melkonian
		<i>Aulacoseira</i> spp.
Bacillariophyta	Coccinodiscophyceae	<i>Melosira</i> spp.
		<i>Amphipleura pellucida</i> Kützing
		<i>Amphora ovalis</i> (Kützing) Kützing
		<i>Bacillaria paxillifer</i> (O.F. Müller) Hendey
		<i>Cocconeis placentula</i> var. <i>euglypta</i> (Ehr.) Grunow
		<i>Caloneis silicula</i> (Ehrenberg) Cleve
		<i>Cymatopleura solea</i> (Brébisson) W. Smith
		<i>Epithemia sorex</i> Kützing
		<i>Encyonema silesiacum</i> (Bleisch) D.G. Mann
		<i>Eucocconeis flexella</i> (Kützing) Meister
		<i>Frustulia rhomboides</i> (Ehrenberg) De Toni
		<i>Gomphonema acuminatum</i> Ehrenberg
		<i>G. truncatum</i> Ehr.
		<i>Gyrosigma acuminatum</i> (Kützing) Rabenhorst
		<i>Navicula capitatoradiata</i> Germain
	Bacillariophyceae	<i>Navicula cryptocephala</i> Kutz.
		<i>Navicula radiosa</i> Kutz.
		<i>Navicula rhyncocephala</i> Kützing
		<i>Navicula tripunctata</i> (O.F.Müll.) Bory
		<i>Nitzschia capitellata</i> Hustedt
		<i>Nitzschia dissipata</i> (Kützing) Grunow
		<i>Nitzschia linearis</i> (Agardh) W. Smith
		<i>Nitzschia recta</i> Hantzsch
		<i>Nitzschia sigmoidea</i> (Ehrenberg) W. Smith
		<i>Nitzschia tryblionella</i> Hantzsch
		<i>Nitzschia veneta</i> Kutz.
		<i>Pinnularia divergens</i> W. Smith
		<i>Rhoicosphenia abbreviata</i> (Agardh)Lange-Berthalot

Table 1. Contd.

	<i>Rhopalodia gibba</i> (Ehrenberg) O. F. Müller
	<i>Sellaphora pupula</i> (Kützing) Mereschkovsky
	<i>Surirella minuta</i> Brébisson
	<i>Surirella ovalis</i> Brébisson
	<i>Tryblionella hungarica</i> (Grunow) Frenguelli
	<i>T. calida</i> (Grun.in Cleve and Grun.) D.G. Mann
Fragilariophyceae	<i>Fragilaria ulna</i> (Nitzsch) Lange-Bertalot

Table 2. Abbreviations and units of environmental variables with basic statistical summaries ($p \leq 0.05$).

Environmental variable	Abbreviation	Unit	Min	Max	Mean	SE
Soluble reactive phosphorus	SRP	$\mu\text{g.L}^{-1}$	14.10	58.98	27.42	8.31
Total phosphorus	TP	$\mu\text{g.L}^{-1}$	48.39	230.72	119.58	33.34
Ammonium	NH ₄	$\mu\text{g.L}^{-1}$	14.76	76.50	36.69	11.59
Nitrate	NO ₃	$\mu\text{g.L}^{-1}$	0.03	0.07	0.06	0.01
Chlorine	Cl	mg.L^{-1}	10.23	96.20	50.64	23.07
Chlorophyll-a	Chl-a	$\mu\text{g.L}^{-1}$	0.88	12.54	5.5	2.50
Caroten	Chr	$\mu\text{g.L}^{-1}$	1.40	12.40	6.4	2.09
Temperature	Temp.	°C	11.6	23.50	15.4	2.15
pH	pH		8.08	8.66	8.48	0.10
Dissolve oxygen concentration	DO	mg.L^{-1}	8.9	11.1	9.9	0.46

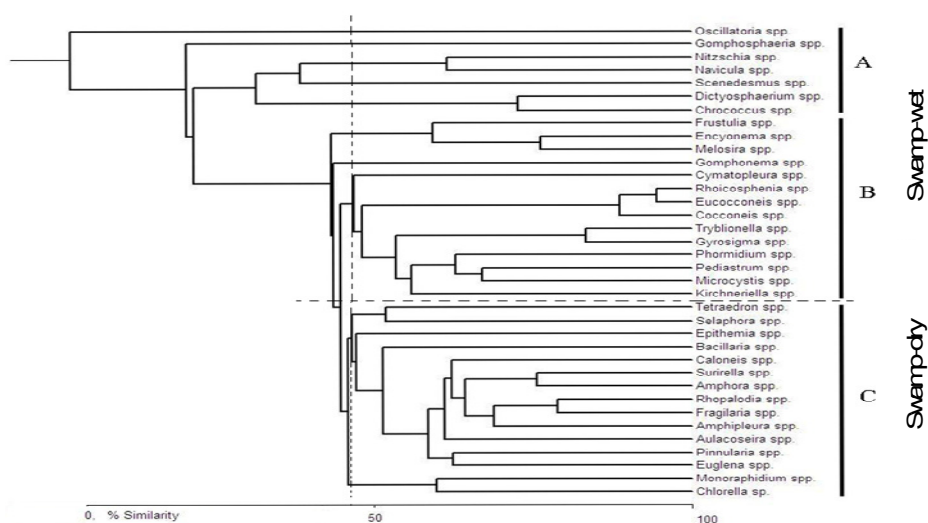


Figure 2. Dendrogram resulting from cluster analysis A, B (Swamp-wet) and C (Swamp-dry) indicate three groups defined by the dotted line arbitrarily drawn at 40% similarity.

Figure 2. The Cluster A corresponds to spring months involving high percentage of *Gomphosphaeria* spp., largely *D. granulatum*, along with *Navicula* spp. and very closely associated with increasing the amount of water, which are areas of high nutrient concentration. Cluster Group B, composed mainly of last spring months and decreasing amount of water, is characterized by the notable decrease in relative abundance of *Gomphonema* spp. and also by the increase in number of species

derived primarily from *Rhoicosphenia* spp., *Cocconeis* spp. and *Pediastrum* spp. Summer months formed the Cluster Group C (third group) dominated by *Selaphora* and *Fragilaria* species. The main benthic algae are cyanobacteria rather than diatoms. Cyanobacteria can reproduce quickly in high temperatures and nutrient-rich conditions (Meng et al., 2010; Min et al., 2012). Due to a massive outburst, it makes all samples have the highest percentage of *Gomphosphaeria* (37%) throughout the

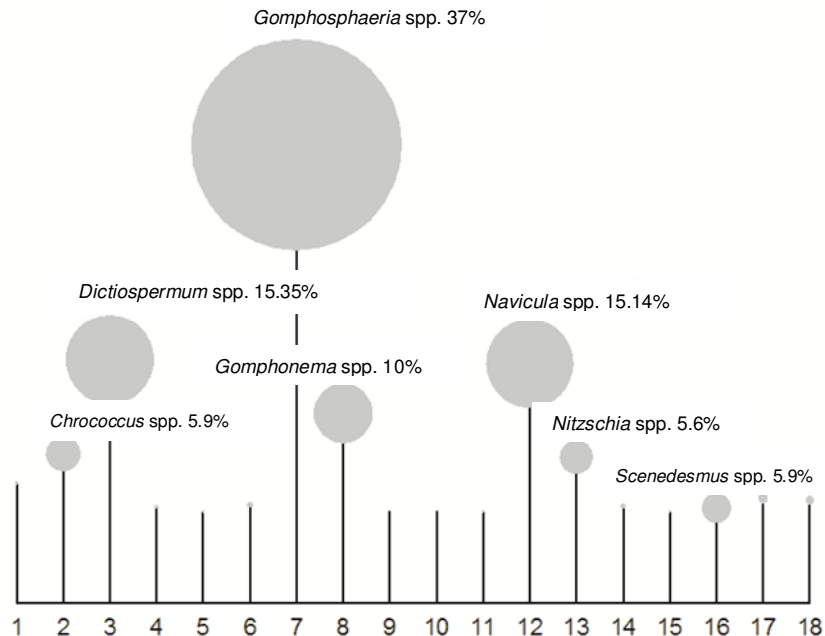


Figure 3. Percentage rates of some dominant genus in total benthic algae.

year (Figure 3). Nutrient availability also greatly influences benthic algae. Nutrient parameters were important determinants of benthic algae distribution patterns in both swamp-wet and dry. However, due to high concentrations of dissolved organic material in these systems (Arts et al., 2000), nutrients may be complexed and unavailable for biological uptake and diversity by benthic algae. Information on seasonal changes in benthic algae community structure and distribution patterns in Karagöl-Aksaz Marsh lake will allow us to understand mechanisms behind changes in important ecosystem processes such as water regime, changing nutrient, diversity and how these valuable Marsh systems may respond to future climate changes.

REFERENCES

- Arts MT, Roberts RD, Kasai F, Waiser MJ, Tumber VP, Plante AJ, Rai H, Lange HJ (2000). The attenuation of ultraviolet radiation in high dissolved organic carbon waters of wetlands and lakes on the northern Great Plains. *Limnol Oceanogr.* 45:292-299.
- Battarbee RW (1986). Diatom analysis. Chichester Wiley U.K: B.E. Berglund. Handbook of Holocene Palaeoecology and Palaeohydrology. pp. 527-570.
- Danilov RA (2002). Seasonal succession of phytoplankton in two brackish bays off the eastern coast of Sweden. *Int. J. Alg.* 4(1):18-24.
- Fleeger JW, Johnson DS, Galván KA, Deegan LA (2008). Top-down and bottom-up control of in fauna varies across the saltMarsh landscape. *J. Exp. Mar. Biol. Ecol.* 357(3):20-34.
- Gottlieb AD, Richards JH, Gaiser EE (2006). Comparative study of periphyton community structure in long and short hydroperiod Everglades Marshes. *Hydrobiologia* 569:195-207.
- Horton BP, Corbett R, Culver SJ, Edwards RJ, Hillier C (2006). Modern saltMarsh diatom distributions of the Outer Banks, North Carolina, and the development of a transfer function for high resolution reconstructions of sea level. *Est. Coas. Sh. Sci.* 69:381-394.
- Komárek J, Anagnostidis K (2005). Süßwasserflora von Mitteleuropa Cyanoprokaryota 2. Teil/ 2nd Part: Oscillatoriales: Büdel B, Krienitz L, Gärtner G, Schagerl M. (eds): 19/2, Elsevier/Spektrum, Heidelberg. pp. 759.
- Krammer K, Lange-Bertalot H (1999a) Süßwasserflora von Mitteleuropa Bacillariophyceae, Band 2/1,1 Teil: Naviculaceae. Spectrum Akademischer Verlag, Heidelberg, Berlin.
- Krammer K, Lange-Bertalot H (1991b). Süßwasserflora von Mitteleuropa, Band 2/3, 3 Teil: Centrales, Frailariaceae, Eunotiaceae. Gustav Fischer-Verlag, Stuttgart.
- Krammer K, Lange-Bertalot H (1991b). Süßwasserflora von Mitteleuropa, Band 2/4, 4 Teil: Achanataceae, Kritische Ergänzungen zu *Navicula* (Lineolatae) und *Gomphonema* Gesamtliteraturverzeichnis. Gustav Fischer Verlag, Stuttgart.
- Krammer K, Lange-Bertalot H (1999b). Süßwasserflora von Mitteleuropa Bacillariophyceae, Band 2/2, Teil 2: Bacillariaceae, Epithemiaceae, Surirellaceae. Spectrum Akademischer-Verlag, Heidelberg, Berlin.
- Legendre P, Gallagher E (2001). Ecologically meaningful transformations for ordination of species data. *Oeco* 129:271-280.
- Mackereth HJH, Heron J, Talling JF (1978). Water Analysis and Some Revised Methods for Limnologists: Biol. Ambleside, UK. Sci. Pub. Freshwater.
- Meng SL, Chen JZ, Hu GD, Qu JH (2010). Phytoplankton community characteristics and its eco-assessment on water quality in Lihu Lake, Taihu Lake. *Res. Environ.Yan. Ba.* 19:30-37
- Min Y, Yan-Ling L, Xiang DY, Qian Liu (2011). Three-year changes in planktonic diatom communities in a eutrophic lake in Nanjing, Jiangsu Province. *Chin. J. Fresh Ecol.* 26 (1):133-141
- Newell SY, Porter D (2000). Microbial secondary production from salt Marsh grass shoots, and its known and potential fates. In: M.P Weinstein, D.A. Kreeger (eds) Concepts and Controversies in Tidal Marsh Ecology, Kluwer Academic Publishers, Dordrecht. pp. 159-185.
- Özener FS (1998). Ecosystems and Geomorphology of interesting patterns in the west of Sinop investigation in terms of ecotourism. 1nd ed. Tübitak Ankara.
- Patrick R, Reimer CW (1966). The Diatoms of the United States, exclusive of Alaska and Hawaii, Philadelphia Monogr. Acad. Nat. Sci. 13(1):1-688.
- Patrick R, Reimer CW (1975). The Diatoms of the United States, exclusive of Alaska and Hawaii, Philadelphia Monogr. Acad. Nat. Sci.

13(2/1):1-213

- Pielou EC (1966). Species-diversity and pattern-diversity in the study of ecological succession. *J. Theor. Biol.* 10:370-383.
- Round FE (1953). An investigation of two benthic algal communities in Malham Tarn, Yorkshire. *J. Ecol.* 41:97-174.
- Shannon CE, Weaver W (1949). *The mathematical theory of communication*: Urbana, USA University of Illinois Press.
- Smol JP, Stoermer EF (2010). *The Diatoms: Applications for the Environmental and Earth Sciences*. Second Edition. UK. Cambridge: University Press.
- Menéndez M, Sanmarti N (2007). Geratology and decomposition of *Spartina versicolor* in a brackish Mediterranean Marsh Estuarine. *Coas. Sh. Sci.* 74:320-330.
- Tailing JF, Driver D (1961). Some problems in the extraction of chlorophyll *a* in phytoplankton. USA Honolulu Hawaii: M. Doty. Atomic Energy Engineering Commission. Proceedings on primary production measurement, marine and freshwater. pp. 142-146.
- Thomas S, Gaiser EE, Gantar M, Scinto L, Jones RD (2006). Quantifying the response of calcareous periphyton crusts to rehydration: a microcosm study (Florida Everglades). *Aquat. Bot.* 84:317-323.