Full Length Research Paper

Influence of seed priming on seed germination and vigor traits of *Vicia villosa* ssp. *dasycarpa* (Ten.)

Karta K. Kalsa¹* and Bekele Abebie²

¹Kulumsa Research Center, P.O. Box 489, Asella, Ethiopia. ²Adama University, P.O. Box 1888, Adama, Ethiopia.

Accepted 3 April, 2012

Laboratory and greenhouse experiments were conducted to investigate the effects of hydro- and osmotic priming on seed germination and vigor traits of two seed lots of wooly pod vetch (*Vicia villosa ssp. dasycarpa* Ten.). Factorial combinations of two seed lots (two periods of storage) and six levels of priming treatment (unprimed control, distilled water (hydro-primed), 0.155, 0.310, 0.465 and 0.620 M KNO₃) were laid out in a completely randomized design with four replications. Seeds were primed for 24 h at temperature of $20 \pm 1^{\circ}$ C. The effects of seed lot, priming treatment, and their interactions were significant at (P<0.05) for seed germination and vigour traits. Seed priming significantly improved all traits investigated in both seed lots with the exception of root length, seedling dry weight, and vigor index-II (germination percentage x seedling dry weight) of stored seeds. Both seed lots achieved significantly higher germination percentage at 0.155 M KNO₃, compared to unprimed controls and hydro-primed seeds. However, hydro-priming showed an advantage over unprimed seeds and osmo-primed seeds in most of the traits studied. Generally, the present study has shown that seed priming (hydro- and osmotic priming) has the potential to improve seed germination and vigor traits in wooly pod vetch.

Key words: Wooly pod vetch, ambient storage, osmo-priming, hydro-priming.

INTRODUCTION

Wooly pod vetch (*Vicia villosa* ssp. *dasycarpa* Ten.) is a widely utilized forage legume in the highlands of Ethiopia in monoculture, in mixture with oat (*Avena sativa*), or for over-sowing of natural pastures. Wooly pod vetch varieties, however, are hard seeded (10 to 70% dormancy) and can occur as voluntary weeds with seeds breaking dormancy following crops for many years (Matic et al., 2008). Moreover, its initial growth is poor and in early growth stages, it is a poor competitor with weeds. Minimizing of hard seed (dormancy) proportion and enhancing of germination and seedling growth in wooly pod vetch could, therefore, result in improved establishment rate and competitiveness to weeds, and reduced risk of voluntary weeds in subsequent crops.

Reports indicated that seed priming (pre-sowing

treatment that involves the controlled hydration of seeds is sufficient to allow pre-germination metabolic events to take place but insufficient to allow radicle protrusion through the seed coat) can improve seed and seedling performance in several species. Hydro-priming (soaking seeds in water) and osmo-priming (soaking seed in chemicals that lower osmotic potential in the seed environment) are commonly used to improve seed performance in various cultivated crops. Arin et al. (2011) reported that osmotic priming with potassium salts improved seedling emergence in onion. Mohammadi and Amiri (2010) reported that seed priming has improved seed performance of canola (Brassica napus L.). Umair et al. (2010) reported that osmotic priming of mung bean (Vigna radiate) seeds in potassium salts has improved seed vigor. Mohammadi (2009) reported that seeds primed with KNO₃ showed highest values of germination percentage, seedling dry weight, and germination rate in late-spring seeded soybean (Glycine max L.).

^{*}Corresponding author. E-mail: kartakaske@gmail.com.

Nascimento et al. (2004) reported that germination rate and germination percentage at different temperature regimes were increased by priming muskmelon (Cucumis melo L.) seeds with KNO₃. Hydro-priming also resulted in lower time taken to 50% germination, higher germination rate, vigor index and final germination percentage in maize (Dezfuli et al., 2008). Hydro-priming hairy vetch (Vicia villosa Roth.) seeds resulted in improved germination rate and seedling emergence from compacted soil (Snapp et al., 2008). On the other hand, Mohammadi and Amiri (2010) indicated that canola seeds primed with KNO₃ showed better vigor performance than those primed with distilled water.

Prolonged seed storage period under adverse conditions (high temperature and relative humidity) is reported to have a positive influence on seed germination and emergence of several species with impermeable seed coat. Storing Vicia dasycarpa, Trifolium tembense, and Vicia sativa seeds for two to five years under ambient temperature and relative humidity has rather improved germination percentage, germination rate, and seedling emergence (Tsegahun et al., 2009; Kalsa et al., 2011). Moreover, Čupić (2005) found in alfalfa (Medicago sativa L.) that the share of hard seed was decreased by storage time, temperature and air moisture changes; and therefore, directly kept the level of seed germination in a longer period of storage. Kalsa et al. (2011) found that there is a significant interaction between hydro-priming treatment and seed age of common vetch. However, published data on effects of seed priming, seed age and their interactions on seed germination and vigor of wooly pod vetch are limited. Hence, the present study was conducted to evaluate the effect of seed priming and seed age on germination and vigor of wooly pod vetch seeds.

MATERIALS AND METHODS

Laboratory and greenhouse experiments were conducted at Kulumsa Agricultural Research Centre, Ethiopia, to determine the effects of seed priming on seed germination and vigor traits in wooly pod vetch. A 2 \times 6 factorial combination, comprising of two seed lots (two periods of storage) and five priming treatments was laid out in a completely randomized design with four replications. Seed lots of wooly pod vetch (*V. villosa ssp. dasycarpa* Ten.) variety Kauhak (which was obtained from the International Center for Agricultural Research Centre, Ethiopia, in the years 2007, and 2008. Seeds from 2007 harvest were stored for one year under ambient conditions (Figure 1) whereas; the 2008 harvests were used as fresh seed without storage. Seed moisture content at harvest of the two seed lots ranged from 10 to 11%.

In the laboratory, seeds were primed with distilled water, 0.155, 0.310, 0.465, 0.620 M KNO₃ by complete immersion soaking, and incubated for 24 h at 20 \pm 1°C under dark condition. The seeds were then washed with distilled water and surface-dried on blotting paper for Ca. 6 h at room temperature. Part of a seed lot was kept unprimed. For every treatment combination, the germination percentage was evaluated by taking 400 seeds (100 seeds in each replication), according to the ISTA (2005) rules. After six hours of

surface drying, seeds were placed in plastic boxes filled (up to 3 cm) with moistened fine sand (0.05 to 0.85 mm), and the boxes were placed in a cement concrete germination room until the final counts were taken after 14 days. The temperature of the room was adjusted to $20 \pm 1^{\circ}$ C.

After the final count in the standard germination test, seedling growth was assessed by measuring root length and seedling dry weight on 10 normal seedlings randomly taken per replicate from the standard germination test. For root length and seedling dry weight, means were computed based on ten seedlings. Root length was measured from tip of the tap root to the joining point of the cotyledon.

After measuring of root length, the seedlings were cut free from their cotyledons, placed in envelopes, then oven-dried at $80 \pm 1^{\circ}$ C for 24 h for determination of seedling dry weight, as suggested by Fiala (1987). Since the seedlings were obtained from the standard germination test, age of the seedlings was 14 days after planting.

The germination rate was measured in four replicates of 25 seeds from each treatment. Seeds were placed on double-layered Whatman #101 filter paper in 90 mm diameter Petri dishes, and kept in an incubator at $20 \pm 1^{\circ}$ C for 20 days, until no further germination took place. Each day, normally germinated seeds (with radicles emerged more than 5 mm out of the seed coat) were removed, until seed germination had ceased. An index was calculated as follows:

Germination Rate = $\sum (G_t/D_t)$, where G_t is the number of germinated seeds on day t and D_t is the number of days after planting when the germinated seeds were counted.

Seedling vigor indices were calculated as follows:

Vigor Index-I = Standard Germination (%) × Seedling Root length (cm);

Vigor Index-II = Standard Germination (%) × Seedling dry weight (mg).

The emergence index and days required for 50% emergence were recorded in a greenhouse on seedlings grown in pots filled with untreated agricultural (clay loam) soil to simulate field conditions. Temperature and humidity control facilities in the greenhouse were not operational during the study period. In each replication, 25 seeds were placed at equal level and covered with a 3 cm thick layer of soil. The soil was then compacted gently and pots were placed in a watered bath to supply water from below. Daily seedling emergence was counted, and continued until constant readings were obtained. The emergence index was calculated as:

Emergence Index = $\sum (E_t/D_t)$,

Where E_t is the number of seedlings emerged, D_t is the number of days after planting when the seedlings were counted (Yang et al., 2005).

Percentage values were arc-sin transformed and analysis of variance was carried out using the Statistical Analysis Software for Windows, Version 9 (SAS Institute Inc, 2002). Multiple comparisons on main effects as well as, their interactions were performed using least square means with Turkey's adjustment.

RESULTS

Results showed that effects of priming treatment, seed age, and their interactions were significant at (P<0.05) for all traits investigated (Tables 1 and 2). Because of the significant interactions, the two seed ages were analyzed



Figure 1. Temperature and relative humidity of open air at Kulumsa during the one year period of seed storage.

Table 1. Analysis of val	riance for germinatio	n percentage, har	rd seed percentage,	germination rate,	root length, a	and seedling dry
weight of wooly pod veto	ch seeds.					

Sources of variation	DF	Germination percentage	Hard seed percentage	Germination rate (per day)	Root length (cm)	Seedling dry weight (mg)
Seed lot (L)	1	7146.8**	1576.2**	377.1**	24.3**	12.1**
Priming (P)	5	173.9**	39.8**	28.2**	6.7**	15.1**
L*P	5	124.5**	10.3**	7.6**	6.8**	4.9*
Error	36	4.3	0.5	0.13	0.44	1.43
R ²		0.98	0.98	0.99	0.85	0.68
CV		3.8	9.4	6.1	6.7	7.2
Mean		54.4	7.9	5.9	9.9	16.5

*, ** Effect is significant at α = 0.05 and α = 0.01 levels of significance, respectively.

Table 2. Analysis of variance for vigor indices, seedling emergence and electrolyte leakage of wooly pod vetch seeds.

Sources of variation	DF	Vigor index-l (cm. %)	Vigor index-II (mg. %)	Emergence index	Days to 50% emergence	Electrical conductivity (µScm ⁻¹ g ⁻¹)
Seed lot (L)	1	308078.7**	1155071.1**	579.4**	150.5**	52381.1**
Priming (P)	5	45119.4**	91427.2**	75.7**	3.2**	73898.0**
L*P	5	39182.4**	57742.7**	89.7**	19.2**	19427.0**
Error	36	1697.3	9057.3	1.6	0.31	97.1
R ²		0.92	0.85	0.96	0.96	0.99
CV (%)		7.8	7.3	8.4	6.5	5.9
Mean		531.1	1298.1	14.7	8.6	167.6

*, ** Effect is significant at α =0.05 and α = 0.01 levels of significance, respectively.

Driming treatments	Germination	percentage	Hard seed percentage				
Priming treatments	Lot-1 (Stored)	Lot-2 (fresh)	Lot-1 (stored)	Lot-2 (Fresh)			
Unprimed	88.0 ± 0.4 (61.7) ^{†c}	58.3 ± 1.5 (35.6) ^f	$9.3 \pm 0.3 (5.3)^{c}$	31.0 ± 1.1 (18.2) ^a			
Distilled water	94.3 ± 1.2 (70.8) ^b	63.8 ± 0.6 (39.6) ^e	3.8 ± 0.3 (2.1) ^d	19.5 ± 0.6 (11.2) ^b			
0.155 M KNO3	95.3 ± 0.5 (72.3) ^a	73.0 ± 1.1 (46.9) ^d	2.3 ± 0.3 (1.3) ^d	21.8 ± 0.6 (12.6) ^b			
0.310 M KNO3	97.0 ± 0.4 (76.0) ^a	67.5 ± 1.5 (42.5) ^e	3.0 ± 0.0 (1.7) ^d	29.0 ± 0.8 (16.9) ^a			
0.465 M KNO3	89.0 ± 0.9 (62.9) ^c	71.8 ± 1.3 (55.9) ^d	3.0 ± 0.0 (1.7) ^d	19.3 ± 0.3 (11.1) ^b			
0.620 M KNO3	83.0 ± 0.4 (56.1) ^c	68.0 ± 1.2 (42.9) ^e	1.0 ± 0.0 (0.6) ^d	20.0 ± 1.1 (11.5) ^b			
Mean	91.1 ± 1.0 (66.6)	67.0 ± 1.1 (42.2)	3.7 ± 0.5 (2.1	23.5 ± 1.1 (13.6)			

Table 3. Means± SE for period of seed storage x priming treatment effects on germination percentage and hard seed percentage of wooly pod vetch seeds.

Means under same parameter sharing letters are not significantly different at α =0.05. [†]Numbers in bracket are arcsine transformed values.

separately.

Seed germination percentage and hard seed proportion

There was a significant effect of seed lot (as explained by period of seed storage) and priming treatment on germination percentage and hard seed proportion. The mean germination percentage was higher for seeds stored for one year under ambient conditions (Table 3). Seed priming improved performance of germination percentage of both seed lots as compared to their untreated controls (Table 3). In the stored seed lot, maximum gains in germination percentage over the control were about 6, 7 and 9% due to priming seeds with water, 0.155 M and 0.310 M KNO₃, respectively. Further increase in concentration of KNO₃ tended to decrease in germination percentage. There was about 5 to 14% increase in germination percentage by priming freshly harvested seeds with water or KNO₃. Highest percentage of 97% for stored seeds was obtained by priming with 0.310 M KNO₃ while the highest 73% germination of freshly harvested seeds was obtained by priming with 0.155 M KNO₃. Our results also showed that the proportion of hard seed of the lot stored for one year was lower than that of freshly harvested seeds (Table 3). As compared to controls, seed priming was significantly decreased hard seed proportion in both seed lots except for freshly harvested seeds at 0.310 M KNO₃.

Germination rate, root length and seedling dry weight

The effects of seed age, seed priming and their interaction were significant on germination rate root length, and seedling dry weight (Table 1). Germination rate of unprimed seeds was faster in stored seeds than in freshly harvested seeds while there was no significant effect on root length and seedling dry weight (Table 4). Priming in water or 0.155 M KNO_3 increased germination

rate by about 100% in stored seeds and by about 180 to 250% in freshly harvested seeds. The effect of priming treatment was non-significant on both root length and seedling dry weight of stored seeds. However, both root length and seedling dry weight of fresh seeds were significantly improved by priming with KNO₃ solutions (Table 4). Priming fresh seeds with 0.155M KNO₃ has increased root length by about 43% while seedling dry weight of the same lot was improved by about 3 to 4 mg by priming with 0.310 to 0.620 M KNO₃ solutions.

Seedling vigor index

The effects of period of seed storage, seed priming and their interaction were significant on seedling Vigor indices I and II (Table 1). Observations from unprimed seeds indicated that stored seeds have significantly better seedling vigor indices as compared to that of fresh seeds (Table 5). Storing wooly pod vetch seeds for one year improved vigor index I and II by 42 and 57%, respectively. Seed priming with water or lower concentrations of KNO₃ significantly improved vigor index I of stored seeds. Vigor index II of fresh seeds was significantly improved by priming with higher concentrations of KNO₃.

Seedling emergence

Both emergence index and days to 50% emergence were significantly influenced by seed priming depending on the period of seed storage. As observed on unprimed controls, stored seeds exhibited higher emergence index and required shorter days to achieve 50% emergence (Table 6). The highest emergence index of stored seeds was recorded by seeds primed with water, followed by those primed with 0.310 M KNO₃. Priming stored seeds with water significantly decreased number of days required for 50% emergence as compared to unprimed seeds. Maximum emergence indices of the younger seed

Table 4.	Means	± SE fo	r period of	f seed	storage	x priming	treatment	effects	on	germination	rate,	root	length,	and	seedling	dry '	weight	of
wooly po	od vetch	seeds.																

Priming treatments	Germination r	ate (per day)	Root len	gth (cm)	Seedling dry weight (mg)		
	Lot-1 (stored)	Lot-2 (fresh)	Lot-1 (stored)	Lot-2 (fresh)	Lot-1 (stored)	Lot-2 (fresh)	
Unprimed	5.7 ± 0.3^{d}	1.2 ± 0.0^{g}	$8.6 \pm 0.3^{\circ}$	8.7 ± 0.2 ^c	15.6 ± 0.8^{bc}	15.1 ± 0.6 ^c	
Distilled Water	11.8 ± 0.2^{a}	3.4 ± 0.1^{e}	9.7 ± 0.3^{bc}	$8.2 \pm 0.3^{\circ}$	15.7 ± 0.6^{bc}	$15.3 \pm 0.6^{\circ}$	
0.155 M KNO ₃	11.4 ± 0.3^{a}	4.2 ± 0.2^{e}	9.0 ± 0.3^{bc}	12.5 ± 0.4^{a}	$15.4 \pm 0.8^{\circ}$	$15.3 \pm 0.6^{\circ}$	
0.310 M KNO ₃	10.5 ± 0.2^{b}	4.2 ± 0.1^{e}	9.8 ± 0.4^{bc}	11.5 ± 0.4^{a}	$15.3 \pm 0.6^{\circ}$	18.4 ± 0.4^{a}	
0.465 M KNO3	$6.9 \pm 0.1^{\circ}$	3.3 ± 0.1^{e}	$8.7 \pm 0.5^{\circ}$	10.6 ± 0.4^{bc}	16.0 ± 0.3^{bc}	18.4 ± 0.5^{a}	
0.620 M KNO3	$6.2 \pm 0.2^{\circ}$	2.4 ± 0.1^{f}	9.1 ± 0.2^{bc}	12.0 ± 0.2^{ab}	17.9 ± 0.7^{abc}	19.6 ± 0.2^{a}	
Mean	8.7 ± 0.5	3.3 ± 0.2	9.1 ± 0.2	10.6 ± 0.4	16.0 ± 0.3	17.0 ± 0.4	

Means under same column sharing letters are not significantly different at α = 0.05.

Table 5. Means \pm SE for period of seed storage x priming treatment effects on seedling vigor indices of wooly pod vetch seeds.

Driming treatmente	Vigor index	κ-I (cm. %)	Vigor index-II (mg. %)			
Priming treatments	Lot-1 (stored)	Lot-2 (fresh)	Lot-1 (stored)	Lot-2 (fresh)		
Unprimed	532.1 ± 19.6 ^c	309.5 ± 13.5 ^d	1374.4 ± 70.0^{ab}	876.3 ± 30.9 ^c		
Distilled water	684.5 ± 28.9 ^{ab}	324.8 ± 13.4 ^d	1480.9 ± 52.4 ^a	977.2 ± 41.5c		
0.155 M KNO ₃	650.6 ± 16.7 ^{ab}	584.3 ± 11.9 ^{bc}	1463.9 ± 71.9 ^a	1111.7 ± 28.1 ^{bc}		
0.310 M KNO ₃	743.8 ± 29.8^{a}	488.9 ± 26.8 ^c	1486.2 ± 60.2 ^a	1240.4 ± 16.3 ^b		
0.465 M KNO ₃	546.3 ± 29.4 ^c	485.6 ± 11.9 ^c	1426.2 ± 32.0 ^a	1321.0 ± 37.5 ^{ab}		
0.620 M KNO ₃	510.2 ± 15.7 ^c	513.1 ± 15.3 ^c	1488.0 ± 52.4 ^a	1331.7 ± 42.6 ^{ab}		
Mean	611.3 ± 20.1	451.0 ± 21.7	1453.3 ± 22.6	1143.0 ± 37.8		

Means under same column sharing letters are not significantly different at $\alpha = 0.05$ level of significance.

Table 6. Means ± SE for period of seed storage x priming treatment effects on seedling emergence index, days to 50% emergence, and seed electrolyte leakage of wooly pod vetch seeds.

Priming treatments	Emergen	ce index	Days to 50%	emergence	Electrical conductivity (µScm ⁻¹ g ⁻¹)			
	Lot-1 (stored)	Lot-2 (fresh)	Lot-1 (stored)	Lot-2 (fresh)	Lot-1 (stored)	Lot-2 (fresh)		
Unprimed	$14.3 \pm 0.6^{\circ}$	9.0 ± 0.4^{e}	7 ± 0^{cd}	11 ± 0^{ab}	59.4 ± 3.5^{9}	146.3 ± 8.0^{e}		
Distilled Water	25.7 ± 0.8^{a}	12.6 ± 0.5^{cd}	5 ± 0^d	10 ± 0^{b}	52.3 ± 0.7^{g}	36.5 ± 1.5^{9}		
0.155 M KNO ₃	21.2 ± 1.2 ^b	9.1±0.4 ^e	6 ± 0^{d}	12 ± 10^{a}	131.0 ± 2.3 ^e	85.9 ± 2.6 ^f		
0.310 M KNO ₃	23.9 ± 0.7a ^b	11.0 ± 0.1 ^{de}	6 ± 0^{d}	10 ± 0^{b}	248.8 ± 5.8 ^b	163.4 ± 3.3 ^{de}		
0.465 M KNO ₃	12.5 ± 0.8^{cd}	13.3 ± 0.4^{cd}	10 ± 0^{b}	8 ± 0^{c}	360.0 ± 7.3^{a}	171.4 ± 1.3 ^d		
0.620 M KNO ₃	11.6 ± 0.5 ^{cde}	12.5 ± 0.3^{cd}	7 ± 0^{cd}	11 ± 1 ^b	352.3 ± 8.7^{a}	$203.9 \pm 5.0^{\circ}$		
Mean	18.2 ± 1.2	11.3 ± 0.4	6.8 ± 0.4	10 ± 0	200.6 ± 26.7	134.6 ± 11.9		

Means under same column sharing letters are not significantly different at $\alpha = 0.05$ level of significance.

lot were recorded by priming seeds with distilled water or 0.465 and 0.620 M KNO₃ (Table 6). Moreover, priming freshly harvested seeds with distilled water and 0.465 M KNO₃ has decreased days to 50% emergence by about 1 and 3 days, respectively, as compared to the unprimed seeds.

Relationship of seedling emergence with germination rate and vigor indices

Linear regression analysis indicated that both emergence index and days to 50% emergence have strong and statistically significant relationships with germination rate,



Figure 2. Relationship of seedling emergence with germination rate (seeds per day) of wooly pod vetch seeds.



Figure 1. Relationship between seedling emergence with vigor index I and vigor index-II of wooly pod vetch seeds.

vigor index I and II. Germination rate exhibited positive relationship with emergence index and opposite association with days to 50% seedling emergence (Figure 2). Germination rate explained about 77% of variations in emergence index and 61% of variations in days to 50% emergence, with standard errors of 2.69 and 1.52, respectively.

Both vigor index I and II had exhibited a positive linear relationship with seedling emergence index while their associations were negative with days to 50% emergence (Figure 3). Vigor index I explained 27% of variation in emergence index while 31% of variation in the same trait was explained by vigor index II. Vigor index II had also explained bout 44 of variation in days to 50% emergence with

a standard error of 1.82 (Figure 3).

DISCUSSION

There was a positive relationship of seed age and seed germination and vigor of wooly pod vetch up to one year of storage. Increase in germination percentage and seedling vigor in older seeds is due to gradual decrease in the proportion of hard seeds through storage time. Such a gradual decrease in the proportion of hard seeds by storage time and an increase in germination in older seeds of alfalfa were also reported by other workers (Čupić et al., 2005).

Improvement in germination traits of wooly pod vetch seeds due to priming is attributed to removal of inhibitory substances in seed coats through hydro-priming or enhanced release of ethylene within embryonic tissues in the presence of imbibed NO₃⁻¹ ion (Benech-Arnold et al., 2000; Varier et al., 2010). Better seedling vigor traits of primed seeds is a combined effect of better status of nutrient reserves in freshly harvested seeds (Murthy et al., 2003), increased seed nitrate level (Benech-Arnold et al., 2000), enhanced permeability of loosened seed coat to water and oxygen (Baskin et al., 2000), and earlier nutrient mobilization (Bewley, 1997).

Mohammadi (2009) reported a significant improvement in seedling dry weight of soybean seeds primed with potassium nitrate. In this study, priming freshly harvested seeds with 0.620 M KNO₃ improved seedling dry weight by about 50 mg and vigor index-II by about 1136% mg over the unprimed control (Tables 4 and 5). Continuous increase in seedling dry weight with KNO₃ concentration is attributed to the positive influence of increased seed nitrate level (Benech-Arnold et al., 2000).

The reason for increase in emergence index and decreased in time for 50% emergence is due to faster seed germination as a result of loosening of seed coats through storage (Čupić et al., 2005), and washing out of inhibitory substances in the seed coat during priming (Leadem, 1997; Baskin et al., 2000; Moïse et al., 2005).

In conclusion, the present study has indicated that seed priming improves seed germination and seedling vigor traits of wooly pod vetch. Hydro-priming is more economical for older seeds while osmo-priming at 0.155 M KNO₃ is recommendable for freshly harvested wooly pod vetch seeds. Moreover, storing wooly pod vetch seeds before sowing is helpful to reduce hard seed (dormancy) proportion. The greater germination due to priming may be statistically significant, but this can be solved by small increases in seeding densities. In contrast, reductions in dormancy appear to result in large decreases in the remnant seed that can be weedy in subsequent years.

REFERENCES

- Arin L, Polat S, Deveci M, Salk A (2011). Effects of different osmotic solutions on onion seed emergence. Afr. J. Agric. Res., 6:986-991.
- Baskin MJ, Baskin CC and Li X (2000). Taxonomy, anatomy and evolution of physical dormancy in seeds. Plant Sp. Biol., 15: 139-152.
- Benech-Arnold RL, SaÂnchez RA, Forcella F, Kruk BC, Ghersa CM (2000). Environmental control of dormancy in weed seed banks in soil. Field Crops Res., 67: 105-122.
- Bewley JD (1997). Seed germination and dormancy. The Plant Cell, 9: 1055-1066.
- Čupić T, Popović S, Grljušić S, Tucak M, Andrić L & Šimic B (2005). Effect of storage time on alfalfa seed quality. J. C. Eur. Agric., 6: 65-68.
- Dezfuli PM, Sharif-zadeh F, Janmohammadi M (2008). Influence of priming techniques on seed germination behavior of maize inbred lines (*Zea mays* L.). J. Agric. Biol. Sci., 3: 22-25.
- Fiala F (1987). Handbook of Vigor Test Methods. International Seed Testing Association, Zurich, Switzerland.
- Kalsa KK, Tomer RPS, Abebie B (2011). Effects of storage duration and hydro-priming on seed germination and vigor of Common vetch. J. Sci. Dev., 1: 65-73.
- Leadem CL (1997). Dormancy unlocking seed secrets. In: Landis TD; Thompson, JR., tech. coords. National Proceedings, Forest and Conservation Nursery Associations. Gen. Tech. Rep. PNWG TR-419. Portland, OR: U.S. Department of Agriculture, Forest Service, Pacific Northwest Research Station: 43-52. Available at: http://www.fcnanet.org/proceedings/1997/leadem1.pdf.
- Matic R, Nagel S, Kirby G. (2008). Factsheet-Wooly pod vetch. http://www.pasturepicker.com.au/Html/Factsheet%20-%20Woolly%20pod%20vetch.pdf, as of 18 June 2011.
- Mohammadi GR (2009). The effect of seed priming on plant traits of late-spring seeded soybean (*Glycine max* L.). Am. Eur. J. Agric. Environ. Sci., 5: 322-326.
- Mohammadi GR, Amiri F (2010). The effect of priming on seed performance of canola (*Brassica napus* L.) under drought stress. Am. Eur. J. Agric. Environ. Sci., 9: 202-207.
- Moïse JA, Han S, Gudynaite,-Savitch L, Johnson DA, Miki BLA (2005). Seed coats: structure, development, composition, and biotechnology. *In Vitro* Cell. Dev. Biol.—Plant, 41 :620–644.
- Murthy UMN, Kumar PP, Sun WQ (2003). Mechanisms of seed ageing under different storage conditions for *Vigna radiata* (L.) Wilczek: lipid per-oxidation, sugar hydrolysis, Millard reactions and their relationship to glass state transition. J. Exp. Bot., 54: 1057-1067.
- Nascimento WM, de Aragao FAS (2004). Muskmelon seed priming in relation to seed vigor. Sci. Agric., 61: 114-117.
- SAS (Statistical Analysis Software) Institute (2002). The SAS System for Windows, Version 9.0, SAS Institute Inc., Cary, NC, USA.
- Snapp S, Price R, Morton M (2008). Seed priming of winter annual cover crops improves germination and emergence. Agron. J., 100: 1506-1510.
- Tsegahun A, Yeheyis L, Amane A, Ziku N, Mekonnen T, Sebsibe A and Muche Y (2009). The impact of storage age on germination and vigor of four recommended highland forage species in Ethiopia. In: Climate change, livestock and people: Challenges, opportunities, and the way forward. Zelalem Yilma and Aynalem Haile (Eds). Proceedings of the 17th Annual conference of the Ethiopian Society of Animal Production (ESAP) held in Addis Ababa, Ethiopia, September 24 to 26, 2009. ESAP, Addis Ababa. pp. 289-294.
- Umair A, Ali S, Bashir K, Hussain S. (2010). Evaluation of different seed priming techniques in mung bean (*Vigna radiate*). Soil Environ., 29: 181-186.
- Varier A, Vari AK, Dadlani M (2010). The sub-cellular basis of seed priming. Curr. Sci., 99: 450-456.
- Yang Q, Ye W, Deng X, Cao H, Zhang Y and Xu K (2005). Seed germination ecophysiology of *Mikania micrantha* H.B.K. Bot. Bul. Acad. Sci., 46: 293-299.ISTA (International Seed Testing Association) (2005). International Rules for Seed Testing. Edition 2005. International Seed Testing Association, Bassersdorf, Switzerland.