

African Journal of Agricultural Research

Review

Fish, Tilapia, and Shigellosis: A review

Alejandro De Jesús Cortés-Sánchez^{1*}, Luis Daniel Espinosa-Chaurand¹, Raquel Garcia-Barrientos² and Alejandra Lorena San Martin-Azocar³

¹Consejo Nacional de Ciencia y Tecnología (CONACYT). Unidad Nayarit del Centro de Investigaciones Biológicas del Noroeste (UNCIBNOR+). Calle Dos No. 23. Cd. del Conocimiento. Av. Emilio M. González. Cd. Industrial C.P. 63173, Tepic, Nayarit, México.

²Lab. Procesos Biotecnológicos, Universidad Politécnica de Tlaxcala. Av. Universidad Politécnica No. 1 San Pedro Xalcaltzinco, Tepeyanco Tlaxcala, C.P. 90180, México.

³Instituto Tecnológico y de Estudios Superiores de Monterrey, Campus Querétaro, Escuela de Ingenieria y Ciencias. Epigmenio González 500, Fracc. San Pablo C.P.76130 Querétaro, Mexico.

Received 7 January, 2021; Accepted 8 March, 2021

Foodborne diseases are considered a relevant issue in health around the world due to their incidence, mortality and negative effects on the economic and productive sector. Fish is considered a food of high nutritional quality, being of global production, distribution and commercialization mainly for human consumption. Among the fish worldwide obtained from capture fisheries and mainly aquaculture for human consumption is Tilapia, due to the adaptability of this fish under cultivation conditions in addition to the fact that its meat is of quality and accessible economic value. Fish due to its composition, is highly susceptible to deterioration and contamination by different hazards throughout the food chain, putting the safety of products and public health at risk. Shigellosis is among the diseases that may be contracted from the consumption of food contaminated by bacteria of the genus *Shigella* spp.; food contamination is mainly related to inadequate or non-hygienic conditions and practices in the production, processing and handling of food. Therefore, the purpose of this review is to provide a general perspective of foodborne diseases, especially shigellosis, causal agents, conditioning factors, related foods such as fish, as well as control and preventive actions in order to protect the food safety and public health.

Key words: Food pathogens, food safety, food microbiology, enterobacteria, fisheries, aquaculture.

INTRODUCTION

The human being, through food, must consume adequate amounts of essential nutrients for the proper growth and development of his physical and intellectual capacities (Izquierdo et al., 2004). In addition, it has been considered that access to safe and nutritious food in sufficient quantity is essential to maintain life and promote good health (Tafur, 2009; Cortés-Sánchez et al.,

*Corresponding author. E-mail: alecortes_1@hotmail.com.

2019; WHO, 2020b). Food safety is defined as the guarantee that food will not cause harm or illness to the consumer; this characteristic is considered a fundamental aspect of public health, in addition to the fact that, along with the nutritional, sensory and commercial characteristics, they constitute the total quality and wholesomeness of food (Fuertes et al., 2014; Jorquera et

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u> al., 2015; Cortés-Sánchez et al., 2019; Cortés-Sánchez et al., 2020).

Foodborne diseases are the product of the consumption of water and food contaminated with different physical, chemical or microbiological agents, which can be found naturally in food. These diseases constitute a relevant cause of mortality in developed and developing countries mainly, appearing at any time and place, prevailing in those where bad hygienic habits are practiced in the production, preparation, conservation and distribution of food, as well as in crowded conditions (Vásquez, 2003).

Throughout the food chain (from farm to table) it has been pointed out that there are various contaminating dangers of food, among them are those of biological origin, mainly bacteria, coming from commonly species of genera Salmonella spp., Shigella spp., Escherichia spp., Listeria spp., Vibrio spp., Campylobacter spp., Clostridium spp., among others) (Tafur, 2009; Palomino-Camargo, 2018; WHO, 2020b). Food is considered a good means of transport and growth environment for microorganisms due to its water and nutrient content, favouring this growth in hot climates, along with unhygienic product handling practices, pathogens can be transferred to food from many sources such as the soil, water, insects, animals, human beings, among others (Barbosa-Cánovas and Bermúdez-Aguirre, 2010). The presence of these different microorganisms in food can put the health of consumers at risk and trigger diseases considered a serious problem in the health sector worldwide due to their incidence, mortality rate, as well as their negative effects on the social and economic development (Tafur, 2009; Palomino-Camargo, 2018; WHO, 2020b).

Nowadays, the globalization of markets has led to processes of supply and exchange of food products, either fresh or processed, between different countries; therefore, the health and food safety issues are considered essential, and cooperation between different entities such as governments, producers and consumers contribute to guaranteeing food safety and public health (Tafur, 2009; WHO, 2020b). The purpose of this review is to provide a general perspective of foodborne diseases, especially Shigellosis, its causal agents, conditioning factors, foods related to its transmission such as fish and tilapia, control and prevention actions in order to protect food safety and public health.

FOODBORNE DISEASES

Foodborne diseases are considered around the world an important public health problem due to their incidence, mortality and the negative effects on socio-economic aspects due to the decrease in productivity, trade and expenses in health systems for hospitalizations and medications, in addition to economic losses to the food industry because of contamination and deterioration of products (Vázquez, 2003; Barrantes and Achi, 2011; Alerte et al., 2012; López et al., 2013; Jorquera et al., 2015; Hernández et al., 2016; Soto et al., 2016; WHO, 2020).

Foodborne diseases are caused by intentional or accidental consumption of water and food contaminated by microorganisms or chemical substances (Vásquez, 2003; WHO, 2020). The most common symptoms of these diseases are gastrointestinal such as nausea, vomiting, diarrhea, abdominal pain and fever, and presenting, in some severe cases, complications such as sepsis, meningitis, abortions, Reiter's Syndrome, Guillan-Barré Syndrome, cancer, or death (López et al., 2013; Soto et al., 2016; WHO, 2020). Globally, it is estimated that these diseases affect one in 10 people, and that 420,000 people die a year, especially children under 5 years of age (WHO, 2020a). For these diseases, population groups with greater risk and affectation have been considered where, in addition to children under 5 vears, are people in poverty, pregnant women, the elderly, and people with weakened immune systems (Alerte et al., 2012; Soto et al., 2016). About 250 agents that cause foodborne diseases have been described and classified, including: chemical agents (inorganic compounds, antimicrobials, toxic food additives, toxins, lubricants, inks, disinfectants, heavy metals, pesticides), physical agents (glass shards, metal, wood, or others), and biological (bacteria, parasites, viruses and prions), the latter being particularly, bacteria such as Clostridium spp., Bacillus spp., Vibrio spp., Aeromonas spp., Salmonella spp., Campylobacter spp., Shigella spp., Listeria monocytogenes, Staphylococcus aureus and Escherichia coli, main responsible for cases and outbreaks (Alerte et al., 2012; Jorquera et al., 2015; Rodríguez et al., 2015; Cortés-Sánchez, 2016; Soto et al., 2016; Cortés-Sánchez et al., 2019).

Bacteria that cause foodborne diseases are classified: 1) Foodborne infections, which occur when a pathogen is present in the food that establishes and multiplies in the consumer, and which present two variants: a) invasive infections, where the microorganism invades, colonizes tissues and organs of the infected person. Within this group: Salmonella spp., Aeromonas spp., Campylobacter spp., Shigella spp., Yersinia spp., and entero-invasive E. coli (EIEC), and b) Toxi-infections, caused by noninvasive bacteria capable of colonizing and multiplying themselves in the intestinal tract of the host, synthesizing and excreting toxins. Some bacterial toxin producers: Vibrio parahaemolitycus, Vibrio cholerae, Bacillus cereus (enterotoxin-producing strains), Clostridium botulinum, Clostridium perfringens, and enteropathogenic variants of E. coli (Kopper et al., 2009; Cortés-Sánchez et al., 2015; Rodríguez et al., 2015). 2) Foodborne intoxication colloquially referred to as food poisoning, derived from the consumption of food containing toxins produced by bacteria that have contaminated and grown in the food at a certain concentration. Some examples are: Clostridium

botulinum, B. cereus (strains that produce the emetic toxin), and *Staphylococcus aureus*. It should be noted that, in the diagnosis and differentiation with infections or general intoxications, they show a more rapid clinical manifestation (Kooper et al., 2009; Rodríguez et al., 2015).

The presence of these diseases can be a direct indicator of the hygienic quality of the food available for consumption, where the contamination of these can occur throughout their production, processing or by the use of contaminated raw material, because multiple pathogenic bacteria can form part of the normal microbiota of animals destined for human consumption such as poultry, pigs, cattle, among others (Cortés-Sánchez et al., 2015). The global incidence of food-borne diseases has been associated with the sum of several factors such as market globalization, climate change, introduction of new products and manufacturing processes, migration, changes in eating habits with the growing demand for ready-to-eat foods, as well as the consumption of food and water outside the home contaminated mainly due to the lack of hygiene in the preparation of food, and the absence in many occasions of water treatment in food preparation (Jorquera et al., 2015; da Cunha et al., 2017; López et al., 2020).

Shigella

Bacilli of the genus Shigella spp., belong to the Enterobacteriaceae family, which are short bacilli of 1-3 µm, Gram negative, do not form spores, immobile, facultative anaerobes, do not ferment lactose but do glucose to acid where they do not typically produce gas, do not hydrolyze urea, do not produce hydrogen sulphide, do not decarboxylate lysine, and do not use citrate and sodium acetate as the only carbon source (Acha and Szyfres, 2001; Torres, 2004; Álvarez et al., 2008; Lampel, 2009; Puerta-García and Mateos-Rodríguez, 2010; da Cunha et al., 2017), and like other Enterobacteriaceae, they can be universally located in soil, water and vegetation, as well as in the gastrointestinal system of animals and human beings (Puerta-García and Mateos-Rodríguez, 2010). These bacteria grow at temperatures between 10 and 48°C but do not grow at temperatures \leq 7°C, they have an optimal growth pH between 6 and 8, although they can survive in acidic pH (5.5) and low relative humidity, NaCl concentrations of 4 to 5%, have a resistance to heat of 60°C / 5 min, and a survival in water up to six months (Huss, 1997; Elika, 2013; da Cunha et al., 2017).

The genus *Shigella* spp., presents four species, and each one of them have several serotypes according to somatic antigen "O", being: *Shigella dysenteriae* (serogroup A, 15 serotypes), *Shigella flexneri* (serogroup B, 8 serotypes), *Shigella boydii* (serogroup C, 20 serotypes), and *Shigella sonnei* (serogroup D, one

serotype), all of them having epidemiological importance (Romero, 1999; Acha and Szyfres, 2001; Álvarez et al., 2008; Lampel, 2009). These bacteria are of worldwide distribution and importance in human health: Shigella dysenteriae (Africa, Asia and Central America), S. flexneri (Developing countries), S. sonnei (developed countries), and S. boydii (India and developed countries) all responsible for Shigellosis, a disease transmitted by contaminated water and food whose severity and lethality depend on the host (age and nutritional status) and causal species (Acha and Szyfres, 2001; León-Ramírez, 2001; Torres, 2004; Lampel, 2009; Puerta-García and Mateos-Rodríguez, 2010; Barrantes and Achi, 2011; Elika, 2013; Ranjbar et al., 2014; da Cunha et al., 2017; CDC, 2020). The infection has an incubation period of 1 to 2 days after consuming contaminated food with a duration of 5 to 7 days of illness (FDA, 2018; CDC, 2020). The disease involves inflammation of the gastrointestinal tract, fever, diarrhea with blood and mucus, sometimes vomiting and headache, while in severe cases complications such as intestinal perforation, reactive arthritis, and Hemolytic-Uremic Syndrome (HUS) can occur as well (León-Ramírez, 2001; Acha and Szyfres, 2001; Torres, 2004; Lampel, 2009; Puerta-García and Mateos-Rodríguez, 2010; Barrantes and Achi, 2011; Elika, 2013; Ranjbar et al., 2014; da Cunha et al., 2017; CDC, 2020).

Shigellosis presents as its main vulnerable groups the elderly, people with weakened immune systems, and children under 5 years of age (Huss, 1998; da Cunha et al., 2017; CDC, 2020). In the latter group, Shigellosis makes an important contribution to the infant mortality rate estimated worldwide at approximately 28,000 children each year (Lluque et al., 2015). Where, *S. dysenteriae* is considered the cause of large and prolonged epidemics of Shigellosis or dysentery and its infection the more serious and lethal than other groups of *Shigella* (Acha and Szyfres, 2001; León-Ramírez, 2002; Mindy et al., 2004).

The treatment of Shigella infections involves reducing water and electrolyte disorders and the use of antibiotics such ampicillin, tetracyclines. erythromycin, as sulfamethoxazole-trimethoprim, azithromycin, and quinolones in severe cases (Romero, 1999; León-Ramírez, 2002; Álvarez et al., 2008; Lluque et al., 2015; da Cunha et al., 2017; CDC, 2020). However, an increase in the incidence of antimicrobial resistance has been reported by isolates of Shigella spp., of clinical origin and food, including fish, considering the phenomenon of resistance in food pathogens a global health problem and the development of a Shigella vaccine is taken as a high priority (Guchi and Ashenafi, 2010; Llugue et al., 2015; Hosseini et al., 2016; Ogbonna and Inana, 2018; Noor et al., 2020). The species of the genus Shigella have a phylogenetic relationship with the genera Salmonella and Escherichia, but unlike these, it does not have animal reservoirs, being detected only in

humans and in some cases primates that can be considered accidental hosts, meaning that there is a risk of zoonotic transmission, in addition to its presence in the environment being related to faecal contamination (Huss, 1997; Acha and Szyfres, 2001; Lampel, 2009; Elika, 2013; da Cunha et al., 2017). Among the pathogenicity mechanisms of these bacteria are the fimbriae that intervene in adhesion phenomena, antigens of the invasive plasmid (Ipa) that is directly related to the infectious process (entry) to the host cell to colonize intestinal mucosa epithelium. endotoxins (lipopolysaccharide wall), enterotoxins (ShET-1 and ShET-2), cytotoxins or shiga toxins (VT-1 and VT2) (Romero, 1999; Leon-Ramirez, 2002; Madigan et al., 2004; Ausina and Moreno, 2005; Alvarez et al., 2008; Polifroni et al., 2009).

Shigella species are highly transmissible, having a low infectious dose of around 10 to 200 cells that can be transmitted through the faecal-oral route by direct contact due to lack of hygiene from person to person, and by consumption of food or water contaminated with human faeces, derived from inadequate hygiene practices in the preparation and handling (infected or asymptomatic food handlers, contaminated equipment and surfaces), or by contamination by vectors such as insects specifically flies (Huss, 1998; Leon-Ramirez, 2002; Álvarez et al., 2008; Lampel, 2009; Elika, 2013; Hernández, 2016; da Cunha et al., 2017; FDA, 2018; CDC, 2020). The most commonly implicated foods in the transmission and cases of Shigellosis are raw or insufficiently heat-treated foods such as raw fruits and vegetables consumed in salads, raw milk and dairy products, raw fish and ovsters, beef. chicken, and drinking contaminated water (Huss, 1998; FAO, 2009; Elika, 2013; FDA, 2018).

FISH

Fish is defined as any food that can be extracted from oceanic or continental waters (fresh or brackish) destined for human or animal consumption (Soares and Gonçalves, 2012). Fish is a food commercialized around the world, highly nutritious, and an important constituent of the human diet since it is a source of polyunsaturated lipids, vitamins, minerals, and proteins with a high biological value and digestibility (Soares and Gonçalves, 2012; Fuertes et al., 2014; FAO, 2020). Fish has been estimated to be one of the main suppliers of animal protein for human consumption around the world, supplying 25% of animal protein in developed countries, and over 75% in developing countries (Baltazar, 2007; Romero et al., 2015). Fish destined for human consumption comes from capture fishing and aquaculture activities where, according to a FAO total world production estimate in 2018 from both activities reached 178.5 million tons, with a per capita consumption worldwide of 20.5 kg (FAO, 2020).

On the other hand, in addition to the high nutritional

contribution, production. distribution and alobal consumption of fish, it should be noted that it is a food with high susceptibility to contamination, deterioration and short commercial life, due to intrinsic properties such as pH close to neutrality, high water activity, and nutrient content easily usable by microorganisms, which leads to economic losses for the food industry (Esteveao and Pucci, 2000; Soares and Gonçalves, 2012; Romero et al., 2015). Foods with these characteristics constitute a favourable substrate for the development of microorganisms, where a minimal microbial contamination, coupled with an adequate incubation temperature and time, makes it a food of high risk of diseases to consumers (Kopper et al., 2009).

Fish contamination can occur at any stage of the food chain by various microorganisms or chemical agents through water, soil, air, fauna, ingredients, utensils, humans, processing or storage conditions, representing a high risk to public health due to the transmission of diseases (Hernández, 2010; Fuertes et al., 2014; WHO, 2020). Fish after fishing and harvesting can be altered and lose its quality through different processes such as autolysis, oxidation and microbial activity (Huss, 1998; Fuertes et al., 2014). In addition, that factors such as species, age, the medium they live in, feeding, conditions of capture, handling, processing, conservation, conditions of transport, distribution and commercialization influence the nutritional value, quality and safety of the fish (Huss, 1998; Soares and Gonçalves, 2012; Fuertes et al., 2014). The microbial activity in fish is relevant in aspects of deterioration and safety, where various members of the fish microbiota are related (Huss, 1998; Fuentes et al., 2011; Romero et al., 2015). Microorganisms are found on all external surfaces - skin and gills - and intestines of live fish. The total number of microorganisms varies enormously, establishing a normal range of 10²-10⁷ / cm² on the surface of the skin, and for gills and intestines between 10³ and 10⁹ CFU / g (Huss, 1999). Bacteria found on the skin and gastrointestinal content of living fish do not invade the muscle or meat, it is sterile due to the protection of its natural defenses, but when the fish dies the bacteria penetrate into the fish (Romero-Jarero and Negrete-Redondo, 2011; Romero et al., 2015). Within the microbiota of fish there are predominantly Gram-negative bacteria belonging to the genus Pseudomonas spp., Moraxella spp., Acinetobacter spp., Shewanella spp., Flavobacterium spp., Vibrio spp., Photobacterium spp., and Aeromonas spp., followed by Gram positives ones such as Bacillus spp., Micrococcus Clostridium spp., Lactobacillus spp., and spp., Corynebacterium spp. (Huss, 1999; Romero et al., 2015). These microorganisms have been associated with the deterioration of fish, and their presence and activity are a function of the nature and conditions of the aquatic environment, where temperature is considered a selective factor (Huss, 1999; Romero-Jarero and Negrete-Redondo 2011; Soares and Gonçalves, 2012; Romero et al., 2015).

The loss of quality and deterioration of fish are especially a consequence of bacterial activity including pathogens (Esteveao and Pucci, 2000). The deterioration of the fish can be perceptible sensorily, through presence of odours derived from the bacterial, and use of small and watersoluble molecules in the fish tissue such as non-protein nitrogen (amino acids, small peptides and trimethylamine oxide), alteration of chain fatty acids short (lactic acid, butyric acid), tissue lipid aldehydes and ketones, volatile sulphides (hydrogen sulphide, dimethyl sulphide, methyl mercaptan), amines (indole, skatole), biogenic amines (histamine, putrescine, and cadaverine), and ammonia from amino acids or metabolized proteins (Romero et al., 2015).

Within aspects of food safety, the pathogenic bacteria present as part of the fish microbiota, and the risk of transmitting diseases to the consumer, are classified into two groups: 1) Autochthonous, widely distributed in aquatic environments around the world, where the temperature of the water presents a selective effect. Some of these microorganisms are: Vibrio spp., Clostridium botulinum. Aeromonas hvdrophila. Plesiomonas shigelloides, and Listeria monocytogenes; and 2) Non-autochthonous, consisting of different Enterobacteriaceae such as Serratia spp., Citrobacter spp., Proteus spp., Salmonella spp., Shigella spp., Enterobacter cloacae, Escherichia. coli, among others, as well as Gram positive ones such as S. aureus, S. epidermidis, S. agalactiae, S. pyogenes, found in fishery products due to faecal contamination from the animal and / or human reservoir of natural waters or aquatic environments and direct contamination of the products during their processing, conservation and handling (Huss, 1998, 1999; Romero-Jarero and Negrete-Redondo 2011; Cortés-Sánchez et al., 2019).

Fish consumption has been associated with cases and outbreaks of diseases around the world, having mainly as causative agents' various bacterial pathogens such as Campylobacter jejuni, E.coli, Listeria monocytogenes, Clostridium spp., Salmonella spp., Vibrio spp., Streptococcus spp., Shigella spp., and Staphylococcus spp. (Soares and Goncalves, 2012; Alerte et al., 2012; Espinosa et al., 2014; Soto et al., 2016). Therefore, fish can be considered a food of sanitary risk as it is a carrier of pathogenic bacteria, either as part of its microbiota or as a consequence of the contamination that can occur in phases of capture, cultivation, processing, handling, conservation, transport, elaboration, and preparation (Esteveao and Pucci, 2000; Romero-Jarero and Negrete-Redondo 2011; Fuentes et al., 2011; Espinosa et al., 2014).

Tilapia

Various species of fish of the *Cichlidae* family are known by the name of Tilapia, which belong to the genus

Sarotherodon, Oreochromis and Tilapia, thus classified according to their parental care behaviour (Baltazar, 2007; Amal and Zamri-Saad, 2011; Jácome et al., 2019). These freshwater fish are of African origin and are highly relevant in the production of animal protein in tropical and subtropical waters around the world, especially in developing countries (Baltazar, 2007; INP, 2018; Jácome et al., 2019).

The genus of greatest interest in aquaculture activities is the *Oreochromis* (*O. niloticus, O. mossambicus,* and *O. aureus*), and hybrids, because of their high growth rates, rapid growth, resistance to diseases, adaptation to captivity, acceptance of feeding diets, and for having good quality meat at an affordable price (Baltazar, 2007; Jácome et al., 2019). Tilapia is one of the finfish with the highest aquaculture production in the world with 10.2% of total production, being the *Oreochromis niloticus* species the most produced in Asian countries, having China among the largest producers (FAO, 2020).

In Latin America, countries like Mexico produce Tilapia mainly through the aquaculture of several species such a) Tilapia rendalli (herbivorous Tilapia). as: b) Oreochromis niloticus (Nile Tilapia), c) O. niloticus Variety Stirling (Stirling Tilapia), d) O. niloticus Rocky Mountain Variety (White Tilapia, Blue Tilapia), e) O. aureus, O. mossambicus, (Mozambique tilapia), f) O. mossambicus Orange Variety (Orange Tilapia), g) O. urolepis hornorum (Tilapia mojarra), and h) Oreochromis sp. (0. mossambicus x O. urolepis hornorum) Florida red Tilapia (hybrid) (INP, 2018; SAGARPA, 2018). The cultivation systems can be extensive, intensive, and semi-intensive according to the cultivation density, food supply and type of cultivation system (INP, 2018). Mexico is considered the fifth largest producer worldwide (SAGARPA, 2018). These fish are cultivated in almost the entire national territory, being the states with the highest production: Chiapas, Jalisco, Nayarit, Sinaloa, Tabasco, Hidalgo, Guerrero, Estado de Mexico, and Veracruz (SAGARPA, 2018; INP, 2018). Finally, this food is marketed with an average weight of 250 - 300 g as fresh whole gutted, whole frozen gutted and processed into fresh or frozen fillet, reaching an annual per capita consumption of 2 kg (INP, 2018).

In aspects of quality, deterioration and food safety, the microbiota of fish and tilapia is relevant and is related to that existing in the waters and habitat from which it comes, temperature, depth and degree of contamination of the waters. Highlighting the presence of Gramnegatives such as: Pseudomonas spp., Shewanella spp., Moraxella spp., Acinetobacter spp., Flavobacterium spp., Vibrionaceae and Aeromonadaceae. Gram positive there is a variable proportion of Bacillus spp., Micrococcus Clostridium Lactobacillus spp., SDD.. spp., and Corynebacterium spp. Bacteria of sanitary interest and contamination indicators members mainly of the Enterobacteriaceae family, including Shigella spp., are found in fish from waters highly contaminated with faecal

matter or subjected to inadequate hygiene conditions during handling or processing (Morales et al., 2004; Onyango et al., 2009; Awuor et al., 2011; Fuentes et al., 2011).

CONTROL, PREVENTION, AND SANITARY REGULATION OF FOOD IN THE CONTEXT OF SHIGELLOSIS

The quality and safety of food products are aspects of relevance in public health, which is evidenced by the development of quality regulations around the world in the different phases of the production chain (Soares and Goncalves, 2012). Generally for fish and fishery products, a large part of the associated foodborne diseases are derived from the lack of good hygiene and handling practices of the fish after capture, as well as the contamination of the fish with pathogenic microorganisms in the primary production stage (aquaculture) due to exposure to harmful microorganisms present in human or animal excreta (WHO/FAO 2016).

International organizations as the Food and Agriculture Organization of the United Nations (FAO) and the World Health Organization (WHO), have developed different codes in order to achieve the objective of making available food safe and suitable for human consumption. Among the codes developed are the essential general principles of food hygiene applied throughout the food chain "CAC / RCP 1-1969" (FAO / WHO, 2020).

Also, the code of practice for fish and fishery and aquaculture products "CXC 52-2003". Its purpose is to provide basic information for the development of fish and shellfish process management systems that incorporate Good Manufacturing Practices (GMP), as well as the application of Hazard Analysis and Critical Control Points systems (HACCP) (FAO / WHO, 2020). It has also been generally established that the actions and key points for the safety of aquaculture products are the hygiene of personnel, equipment and materials for capture or handling, cleaning of ponds, fish health and water quality (WHO / FAO 2016).

Likewise, FAO has developed guidelines for risk-based fish inspection that provide technical information to support fish inspection systems on food safety aspects in order to guarantee consumers access to safe and nutritious fish, in addition to helping to develop, harmonize and apply preventive inspection systems, as well as to guide the development of regulatory frameworks for fish inspection in developing member countries (FAO, 2009).Within the food safety regulations developed for fishery and aquaculture products in European countries such as Spain, it has been through the Order of August 2, 1991, where the standards and microbiological limits for Shigella spp., were established, which must be absence of Salmonella-Shigella in 25 g of sample of fresh, salty, refrigerated, frozen, dry, salty, salty and dry, semi-pickled and smoked products (BOE,

1991). However, this provision was repealed by Royal Decree 135/2010, of February 12, 2010 by transposition of community regulations, such as Regulation (EC) No. 2073/2005, relating to microbiological criteria applicable to food products (BOE, 2010).

The legislation on food safety in the European community for the production, acceptability and commercialization of food, can be found in the Regulation (CE) No. 178/2002 laying down the general principles and requirements of food law, establishing the European Food Safety Authority, and laying down procedures in matters of food safety. Regulation (EC) No 852/2004 on the hygiene of food products that includes the general application of procedures based on the principles of Hazard Analysis and Critical Control Points (HACCP) that, along with the application of correct hygienic practices, reinforce the responsibility of food business operators. Regulation (CE) No 853/2004 establishes specific hygiene rules for food of animal origin, including fish and products. Regulation (EC) No 854/2004 establishes specific rules for the organization of official controls of products of animal origin intended for human consumption. Regulation (EC) No 882/2004, carried out on official controls, to guarantee the verification of compliance with the legislation on feed and food and the regulations on animal health and animal welfare, and Regulation (EC) No 2073/2005 which establishes the microbiological criteria applicable to food products including fish and its various biological hazards, providing guidance on the acceptability of food products and manufacturing processes (EC, 2020). However, the latter does not establish specific microbiological limits for food and its contamination by Shigella (Elika, 2013).

In Latin American countries such as Mexico, sanitary regulation regarding fish safety through the food chain begins in primary production, with the implementation of good aquaculture practices during primary fish production and the implementation of good aquaculture practices specifically in Tilapia production to ensure food safety (García and Calvario, 2008; SENASICA, 2019). Likewise, the regulatory framework is made up of the Official Mexican Standard "NOM-242-SSA1-2009" for fresh, refrigerated, frozen and processed fish products, which include the sanitary specifications and test methods for these products, although for Shigella spp., no sanitary specification is established in these foods, and it has been suggested as a quality criterion for raw foods of <1 CFU / 25g of sample (Hernández, 2016). The "NOM-251-SSA1-2009" standard, which establishes the minimum requirements of good hygiene practices that must be observed in the process of food, beverages or food supplements, and their raw materials in order to avoid their contamination throughout their process; it, also establishes the Hazard Analysis and Critical Control Point (HACCP) system and guidelines for its application. For the specific case of the fish product processing industry, the "NOM-128-SSA1-1994" standard establishes the application of a system HACCP in the industrial

processing plant. The "NOM-210-SSA1-2014" standard establishes the general and alternative test methods for the determination of microbial and pathogenic indicators in food for human use and consumption, although it does not establish specific methods for *Shigella* spp., the "NOM-230-SSA1-2002" standard, which specifies the sanitary requirements that public and private supply systems must meet for the management of water intended for human use and consumption. And the "NMX-F-605-NORMEX-2016" standard that establishes the provisions of good hygiene and sanitation practices that food and beverage service providers must comply with to obtain the distinctive "H".

In general, actions for the control and prevention of foodborne diseases such as Shigellosis should be all along the food chain, from primary production, handling, processing or transformation of food, to consumption (Figure 1), which is achieved through the implementation of conditions hygienic and sanitary. Sanitation Standard Operation Procedure (SSOP), good hygiene practices with an emphasis on hygiene training and information for food handlers (detection and control of symptomatic and asymptomatic carriers), microbiological criteria of raw materials and final products for their acceptability, operational temperature control where the food must be subjected to heat treatments above 60°C, as well as to preserve the cold chain during transport, storage and distribution of raw foods susceptible to contamination, and finally the strengthening of all the previous measures with control systems based on the Hazard Analysis and Critical Control Points (HACCP), and traceability systems for the inspection, certification and sanitary control of products throughout the food chain (Huss, 1998; Leon-Ramirez, 2002; CAC/GL 60-2006; Lampel, 2009; Bracho et al., 2012; Elika, 2013; Hernandez, 2016; da Cunha et al., 2017; Palomino-Camargo, 2018; PAHO, 2020).

While the actions at the level of manipulation and end consumer in homes are suggested to being carried out with good hygiene and manipulation practices in the preparation and cooking of food to prevent contamination, the World Health Organization has developed manuals to promote safety of food in this context through the cleaning and hygiene of food preparation areas, handlers, equipment and utensils to be used, to avoid cross contamination (raw and cooked food), to avoid the presence of vectors such as flies, favour conditions of time and temperature in the preparation (> 60°C), and preservation of food (<5°C), as well as to have a good sanitary quality of the water used (OMS, 2007; Elika, 2013; Hernandez, 2016).

Food analysis in the laboratory

As part of the control and prevention of diseases through food, government sanitary agencies and the food industry, in addition to hygiene education, inspection of facilities and activities, consider microbiological analysis (Huss, 1997). The objective of microbiological analysis in foods such as fish is the detection of pathogens, indicators of faecal contamination, inadequate or poor processing conditions, and practices in hygiene aspects, to ensure the quality is at an adequate microbiological level (Huss, 1998).

The detection of pathogens in food such as *Shigella* spp., is generally carried out by conventional cultivation because its execution and costs are accessible. These methods are based on phenotypic characteristics of microorganisms such as their morphology, development, biochemical, metabolic, and antigenic properties (Table 1) (Prats and Mirelis, 1998; Barrantes and Achi, 2011; Bou et al., 2011; da Cunha et al., 2017).

In traditional food microbiological analysis methods, it must be considered that *Shigella* is sensitive to environmental conditions and can die quickly, particularly when exposed to direct sunlight or food processing conditions (dehydration), intrinsic characteristics of the food (composition, pH and saline concentration), in addition to the natural microbiota that competes for nutrients, affect and complicate the isolation of the pathogen from food, so that the analysis may require enrichment methods for better detection when it is in low numbers or weakened physiological state (Lampel, 2009; Anselmo et al., 2020).

In traditional culture, various differential and selective culture media are used for the isolation and detection of Shigella spp.; the most used are Eosin Methylene Blue Agar (EMB Agar), Mac Conkey agar, and the Salmonella-Shigella agar (SS), where typical colonies are colourless or transparent (because lactose is not fermented), the Xylose-Lysine-Deoxycholate (XLD) agar, where colonies are transparent (no xylose fermentation or lysine modification), the Hektoen agar, where colonies are transparent green (no lactose fermentation and no hydrogen sulphide production), and the Tergitol 7 agar, where Shigella colonies appear blue (lactose negative). Suspicious colonies are inoculated onto Kligler's media (KIA) or Triple Sugar Iron agar (TSI) and Lysine Iron Agar (LIA). Any colony that ferments glucose in these media without producing gas, does not use lactose, does not produce H₂S, is immobile, does not decarboxylate and deaminates lysine, does not deaminates tryptophan, in addition to not having urease or oxidase production is suspected Shigella spp. (Prats and Mirelis, 1998; León-Ramirez, 2002; Mindy et al., 2004; Álvarez et al., 2008; Lampel, 2009; Wallace and Andrew, 2018; Anselmo et al., 2020). The biochemical identification and differentiation tests of bacteria can be manual or automatic, having commercially available manual multiple test systems to API (bioMérieux), Enterotube (BBL), Oxi / Ferm Tube (BD), RapID and MicroID systems (Remel), biochemical ID systems (Microgen). While the automated multiple test systems available are MicroScan, ATB, Pasco, Wider, Phoenix, Vitek among others (Bou et al.,



Figure 1. General scheme of contaminating factors caused by *Shigella* and prevention actions in the food chain of fish and products. SSOP: Sanitation Standard Operation Procedure.

2011). In addition to serological identification by agglutination with specific antisera (Prats and Mirelis, 1998; León-Ramirez, 2002; Mindy et al., 2004; Álvarez et al., 2008; Lampel, 2009; Wallace and Andrew, 2018; Anselmo et al., 2020).

Various standardized methods have been developed for the isolation and identification of Shigella spp., in food matrices. Some of these methods are that of Wallace and Andrew (2018), present in the Bacteriological Analytical Manual (BAM) of the U.S. Food and Drugs Administration (FDA), where through a traditional culture method that uses a selective enrichment phase where 25 g of sample are placed and homogenized in 225 ml of Shigella broth added with novobiocin (0.5 to 3 µg / ml), the pH is adjusted to 7 and incubated under anaerobic conditions at 44°C (for S. sonnei) and 42°C (for the rest of Shigella species), then the enrichment culture suspension is shaken and sub-cultured on Mac Conkey agar incubating for 20 h at 35°C, and subsequently analyzing the growth of typical colonies (slightly pink and translucent, with or without rough edges). The colonies are selected for confirmation through different biochemical tests (glucose broth, TSI agar, lysine broth decarboxylase, motility agar and tryptone) and serological identification tests.

Another standardized traditional microbiological analysis method is the one developed by the International Organization for Standardization (ISO) ISO 21567:2004, where the enrichment is done in *Shigella* broth with novobiocin, incubated at 41.5± 1°C from 16 to 20 h in anaerobic conditions. Afterwards, varied differential and selective culture media, such as Mac Conkey agar, XLD agar, and Hektoen agar, are inoculated and incubated at 37°C for 20h to 24 h. Colonies with typical morphology are subjected to biochemical confirmation tests using Triple Sugar Iron agar (TSI), motility agar, urea, lysine decarboxylase, ornithine decarboxylase, indole formation, detection of β-galactosidase, and the use of different carbohydrates and as additional acetate, mucate and citrate tests (Figure 2) (ANMAT-RENAPRA, 2013).

However, despite being the traditional microbiological methods of frequent and generalized use in the isolation and identification of pathogens in food, these can present drawbacks, such as the absence of correlation between the morphological or phenotypic characteristics of the isolate analyzed, and that corresponding to the strain or type species used, the analysis time and the results obtained may take days or weeks, they may present low sensitivity, in addition to the fact that some bacterial cells

Test	Shigella spp.	Escherichia spp.	Salmonella spp.	Klebsiella spp.
Oxidase	-	-	-	-
Nitrates reduction	+	+	+	+
Arginine	-	V	+	-
Gelatin Hydrolysis	-	-	V	V
Alanine deaminase	-	-	-	-
Acid production of: Adonitol	-	-	-	V
Inositol	-	-	V	+
Carbon source: Citrate	-	-	+	V
Malonate	-	-	V	V
Tartrate	-	+	V	V
Voges Proskauer	-	-	-	V
Methyl Red	+	+	+	V

Table 1. Biochemical identification characteristics of different Enterobacteriaceae (Pelczar, 1982; Madigan et al., 2004).

V = variable reaction, - negative reaction, + positive reaction.

may enter a Viable But Non-Culturable state (VBNC) that consists of a state of survival to adverse conditions (food processing) and that most *Shigella* species such as *S. dysenteriae* can present and the method used may not be efficient (Ishrat et al., 1996; Wen-Bin et al., 2007; Barrantes and Achi, 2011; Bou et al., 2011; Palomino-Camargo and González-Muñoz, 2014; Huertas-Caro et al., 2019).

To try to avoid the drawbacks of traditional microbiological analysis methods, alternative methods have been developed for the detection of food pathogens based on the Polymerase Chain Reaction (PCR) and its multiple variants; which includes the detection of different targets of genetic material that include housekeeping genes, genes that encode virulence factors, as well as the Whole Genome Sequencing (WGS) by Single Nucleotide Polymorphism (SNP) of the whole genome, markers and k-mers. In this way, for food safety, epidemiological or phylogenetic studies, a greater number of samples can be analyzed, and the results are obtained with greater speed and sensitivity (Barrantes and Achi, 2011; Shao et al., 2011; Palomino-Camargo and González-Muñoz, 2014; Ragupathi et al., 2018; Huertas-Caro et al., 2019).

For *Shigella* spp., various methods have been developed using virulence regions or factors for its detection, such as: the *ial* region (invasion associated locus) present in the invasion plasmid, whose product is 320 bp and is encoded in the SSTT spa's maximum operon (essential in the invasive phenotype) (Barrantes and Achi, 2011), Shiga toxin gene (*Stx*) (Ishrat et al., 1996; Villacrés et al., 2015), enterotoxin 1 (ShET-1), encoded protein in the bacterial chromosome by genes *set1A* (subunit A of the protein) of 309 bp and *set1B* (subunit B of the protein) of 147 bp (Villacres et al., 2015), insertion sequence 1 (IS1) 768-bp DNA fragment (Wen-Bin et al., 2007), 215 bp *virA* gene (Villalobo and

Torres, 1998; Alipour et al., 2012), *icsA* gene encodes the IcsA protein, for intercellular mobility and dispersion (Alipour et al., 2012), and plasmid invasion antigen "H" or *ipaH* gene of 610 bp specific for *Shigella* spp., being determined as a stable gene to be detected because it is found in the chromosome and in plasmid DNA in multiple copies (Pichel and Caffer, 2016; Wang et al., 1997; Thiem et al., 2004; Lampel, 2009; Barrantes and Achi, 2011; Mokhtari et al., 2013; Radhika et al., 2014; Lin et al., 2010; Ranjbar et al. 2014; Villacrés et al., 2015; Nadella et al., 2017).

Likewise, molecular subtyping methods such as Pulsed-Field Gel Electrophoresis (PFGE) and Multilocus Variable-Number Tandem Repeat Analysis (MLVA) have been incorporated to determine the genetic relationships between isolates of Shigella spp., which contribute to epidemiological surveillance aspects, identification, and study of outbreaks caused in different geographical regions by the pathogen (Garitano et al., 2011; Pichel et al., 2012; Guzman-Herrador et al., 2011; Pichel and Caffer, 2016; Bakhshi et al., 2017). Another alternative method in the identification of isolates of food pathogens such as Shigella are those based on the analysis of proteins (mainly 2-20 KDa ribosomes) expressed by the genome through mass spectrometry, where a spectrum of specific masses, the "Footprint", for each genus and species by using Matrix Assisted Laser Desorption Ionization - Time Of Flight - Mass Spectrometry (MALDI-TOF MS) (Bou et al., 2011; Schaumann et al., 2013; Paauw et al., 2015; Ragupathi et al., 2018; Huertas-Caro et al., 2019); those are among the characteristics of these systems in their use that they do not present extraction procedures. A bacterial colony can be used directly, demonstrating that it is possible to compare the spectra generated with previous databases, and also shows a speed of execution of the technique, obtaining and precision of results (Bou et al., 2011; Paauw et al., 2015).



Figure 2. Flow chart for the isolation of Shigella spp., in food (ISO 21567:2004; ANMAT-RENAPRA, 2013).

CONCLUSIONS

Fish is one of the constituent foods of the human diet due to its nutritional value. Tilapia is among the world's most widely produced fish through fishing and aquaculture, mainly intended for human consumption. Fish, despite being a food of high nutritional value and part of the human diet, is very susceptible to deterioration and contamination through the different phases of the food chain by various bacterial agents that cause diseases, so it can be considered a sanitary risk food.

Shigellosis is a Foodborne disease caused by different species of bacteria of the Shigella genus. Foods related to disease cases and outbreaks are those eaten raw or subjected to inadequate cooking and storage conditions. The causative agents of shigellosis are cosmopolitan in distribution with humans and animals as a reservoir and contaminate food. such as fish. through can contamination with fecal material from culture or capture waters, as well as by inadequate hygiene conditions and practices during handling in production and processing. The general measures indicated to guarantee safety throughout the fish food chain and protect public health from shigellosis causative agents are the implementation of good hygiene practices and process controls, implementation of Hazard Analysis and Critical Control Points systems (HACCP), application of microbiological tests and their correlation with microbiological criteria for the monitoring of product quality and production conditions, as well as constant training in hygiene for food handlers.

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

REFERENCES

- Acha PN, Szyfres B (2001). Zoonosis y enfermedades transmisibles comunes al hombre y a los animales. Publicación Científica y Técnica No. 580. Tercera edición. Volumen I. Bacteriosis y Micosis. Organización Panamericana de la Salud (OPS). Organización Mundial de la Salud (OMS). Available at: https://www.paho.org/hq/dmdocuments/2017/acha-zoonosis-spa.pdf. Alerte V, Cortés AS, Díaz TJ, Vollaire ZJ, Espinoza MME, Solari GV,
- Cerda LJ, Torres HM (2012). Foodborne disease outbreaks around

the urban Chilean areas from 2005 to 2010. Revista chilena de infectología, 29(1):26-31. Available at: https://dx.doi.org/10.4067/S0716-10182012000100004

- Alipour M, Talebjannat M, Nabiuni M (2012). Polymerase chain reaction method for the rapid detection of virulent *Shigella* spp. International Journal of Molecular and Clinical Microbiology 2:134-137. Available at: https://www.sid.ir/en/Journal/ViewPaper.aspx?ID=285309.
- Álvarez Martínez M, Buesa Gómez J, Castillo García J, Vila Estape J (2008). Editores: Cercenado Emilia y Cantón Rafael. 30. Diagnóstico microbiológico de las infecciones gastrointestinales. Procedimientos en Microbiología Clínica. Recomendaciones de la Sociedad Española de Enfermedades. Infecciosas y Microbiología Clínica. Available at:

https://www.seimc.org/contenidos/documentoscientificos/procedimien tosmicrobiologia/seimc-procedimientomicrobiologia30.pdf

Amal MNA, Zamri-Saad M (2011). Streptococcosis in tilapia (Oreochromis niloticus): a review. Pertanika Journal of Tropical Agricultural Science 34(2):195-206. Available at: https://www.researchgate.net/profile/Mohammad_Noor_Amal_Azmai/ publication/220038538_1_Amal_MNA_and_Zamri-

Saad_M_2011_Streptococcosis_in_Tilapia_Oreochromis_niloticus_A _Review_PERTANIKA_Journal_of_Tropical_Agricultural_Science_34 2_195-206/links/5fb106c445851518fda6e3ae/1-Amal-MNA-and-Zamri-Saad-M-2011-Streptococcosis-in-Tilapia-Oreochromisniloticus-A-Review-PERTANIKA-Journal-of-Tropical-Agricultural-Science-342-195-206.pdf.

- ANMAT/RENAPRA (2013). Shigelosis. Enfermedades transmitidas por alimentos. Ficha técnica N° 6. Administración Nacional de Medicamentos, Alimentos y Tecnología Médica (ANMAT). Red Nacional de Protección de Alimentos (RENAPRA). República de Argentina. Available at: http://www.anmat.gov.ar/webanmat/Publicaciones/Shigelosis.pdf.
- Anselmo RJ, Ojeda PA, Barrios HA (2020). Detection and antimicrobial susceptibility of *Shigella spp.* in prepared, ready to eat salads. Información Tecnológica 31(1):13-20. Available at: https://dx.doi.org/10.4067/S0718-07642020000100013
- Ausina Ruiz V, Moreno GS (2005). Tratado SEIMC de enfermedades infecciosas y microbiología clínica. 1ra edición. Editorial Médica Panamericana. Buenos aires; Madrid.1595 pp.
- Awuor WS, Miruka OD, Eliud WN (2011). Characterisation of Salmonella Isolated from Nile Tilapia (Oreochromis niloticus) along Lake Victoria Beaches in Western Kenya. International Journal of Biological, Life and Agricultural Sciences 4.0(8). Available at: http://doi.org/10.5281/zenodo.1329651.
- Bakhshi B, Bayat B, Lari AR (2017). Multiple-locus variable-number tandem-repeat analysis (MLVA) of *Shigella sonnei* isolates of 2012 outbreak IR Iran. Microbial Pathogenesis 102:69-73. Available at: https://doi.org/10.1016/j.micpath.2016.10.021.
- Baltazar PM (2007). Tilapia in Peru: aquiculture, market, and perspectives. Revista Peruana de Biología 13(3):267-273. Available at: http://www.scielo.org.pe/scielo.php?script=sci_arttext&pid=S1727-99332007000100022&Ing=es&tIng=es.
- Barbosa-Cánovas G, Bermúdez-Aguirre D (2010). Nonthermal Processing of Food. Scientia Agropecuaria 1(1):81-93. Available at: https://revistas.unitru.edu.pe/index.php/scientiaagrop/article/view/20
- Barrantes K, Achí R (2011). Microbiological quality and pathogen analysis (Shigella and Salmonella) of lettuce. Revista de la Sociedad Venezolana de Microbiología 31(1):31-36. Available at: https://www.redalyc.org/pdf/1994/199421829007.pdf
- BOE (1991). Orden de 2 de agosto de 1991 por la que se aprueban las normas microbiológicas, los límites de contenido en metales pesados y los métodos analíticos para la determinación de metales pesados para los productos de la pesca y la acuicultura. Documento BOE-A-1991-20734. Boletín Oficial del Estado (BOE). Gobierno de España. Available at: https://www.boe.es/buscar/doc.php?id=BOE-A-1991-20734
- BOE (2010). Real Decreto 135/2010, de 12 de febrero, por el que se derogan disposiciones relativas a los criterios microbiológicos de los productos alimenticios. Documento BOE-A-2010-3032. Boletín oficial del estado (BOE). Gobierno de España. Available at: https://www.boe.es/buscar/doc.php?id=BOE-A-2010-3032.

Bou G, Fernández-Olmos A, García C, Sáez-Nieto JA, Valdezate S

(2011). Bacterial identification methods in the microbiology laboratory. Enfermedades Infecciosas y Microbiología Clínica 29(8):601-608. Available at: DOI: 10.1016/j.eimc.2011.03.012.

- Bracho Rubio MG, Muñoz Montiel MA, María del Carmen Gómez, Márquez A, Ávila Larreal AG, Castillo Machado EM (2012). Prevalence of Salmonella and Shigella among Food Handlers. Multiciencias 12:295-299. Available at: https://www.redalyc.org/pdf/904/90431109048.pdf.
- CAC/GL 60-2006. Principles for traceability/product tracing as a tool within a food inspection and certification system. OMS. FAO. Codex Alimentarius. Available at: http://www.fao.org/fao-who-codexalimentarius/sh-proxy/es/?lnk=1&url=https%253A%252F%252Fworkspace.fao.org%2 52Fsites%252Fcodex%252FStandards%252FCXG%2B60-

52Fsites%252Fcodex%252 2006%252FCXG_060e.pdf.

CAC/RCP 1-196. General principles of food hygiene. World Health Organization (WHO). Food and Agriculture Organization of the United Nations (FAO). Codex Alimentarius. Available at: http://www.fao.org/fao-who-codexalimentarius/shproxy/es/?lnk=1&url=https%253A%252F%252Fworkspace.fao.org%2 52Fsites%252Fcodex%252FStandards%252FCXC%2B1-

1969%252FCXP 001e.pdf

- Centers for Disease Control (CDC) (2020). Shigella Shigellosis. Centers for Disease Control and Prevention, National Center for Emerging and Zoonotic Infectious Diseases (NCEZID), Division of Foodborne, Waterborne, and Environmental Diseases (DFWED). Available at: https://www.cdc.gov/shigella/general-information.html.
- Cortés-Sánchez ADJ, Díaz-Ramírez M, Salgado-Cruz MDP, Hernandez-Nava RG (2020). Food Safety and Fish Production the Case of Staphylococcus aureus: A Review. OnLine Journal of Biological Sciences 20(4):296-311. Available at: DOI: 10.3844/ojbsci.2020.296.311.
- Cortés-Sánchez ADJ, Díaz-Ramírez M, Sánchez-Mundo MLDL, Hernández-Álvarez AJ, and Barrón-Sosa LR (2015). Foodborne diseases, probiotics and health. Asian Journal of Microbiology, Biotechnology and Environmental Sciences 17(3):763-774.
- Cortés-Sánchez ADJ, Diaz-Ramirez M, Villanueva-Carvajal A, San Martin-Azocar AL (2016). Vibrio Cholerae as it Relates to Food and Health. Pakistan Journal of Nutrition 15: 607-617. Available at: DOI: 10.3923/pjn.2016.607.617
- Cortés-Sánchez ADJ, Espinosa-Chaurand LD, Garza-Torres R, Diaz-Ramirez M, Salgado-Cruz MDLP, Sanchez-Minutii L, Garcia-Barrientos R. (2019). Foodborne diseases, fish and the case of Aeromonas spp. African Journal of Agricultural Research 14(11):617-628. Available at: https://doi.org/10.5897/AJAR2019.13872.
- Cortés-Sánchez ADJ, Guzmán-Robles ML, Garza-Torres R, Espinosa-Chaurand LD, Díaz-Ramírez M (2019). Food Safety, Fish and Listeriosis. Turkish Journal of Agriculture-Food Science and Technology 7(11):1908-1916. Available at: DOI: https://doi.org/10.24925/turjaf.v7i11.1908-1916.2888.
- CXC 52-2003. Code of practice for fish and fishery products. World Health Organization (WHO). Food and Agriculture Organization of the United Nations (FAO). Codex Alimentarius. Available at: http://www.fao.org/fao-who-codexalimentarius/sh-

proxy/es/?lnk=1&url=https%253A%252F%252Fworkspace.fao.org%2 52Fsites%252Fcodex%252FStandards%252FCXC%2B52-2003%252FCXC 052e.pdf

- da Cunha FDPL, Vilela MLAS, Maximiano T, Barbosa TMM, de Lima Guimarães DA, Toledo RCC (2017). Shigella sp: Um problema de saúde pública. Higiene Alimentar 31:264-265. Available at: https://docs.bvsalud.org/biblioref/2017/04/833025/264-265sitecompressed-52-57.pdf.
- European Comission (EC) (2020). Food. Microbiological criteria. European Comission (EC). Available at: https://ec.europa.eu/food/safety/biosafety/food_hygiene/microbiologic al_criteria_en.
- Elika (2013). Shigella. Fundación vasca para la seguridad agroalimentaria. Available at: https://seguridadalimentaria.elika.eus/wpcontent/uploads/2018/01/12. Shigella.pdf.
- Espinosa L, Varela C, Martínez EV, Cano R (2015). Brotes de enfermedades transmitidas por alimentos. España, 2008-2011

(excluye brotes hídricos). Boletín Epidemiológico Semanal 22(11):130-136. Available at: http://revista.isciii.es/index.php/bes/article/view/889/1070.

Estevao BS, Pucci OH (2000). Microbiological controls and critical control points during control "for export" hake fillets manufacturing process. Archivos Latinoamericanos de Nutrición 50(2):171-176. Available

http://ve.scielo.org/scielo.php?script=sci_arttext&pid=S0004-0622200000200010&Ing=es&tIng=pt.

- Food and Agriculture Organization (FAO) (2009). Directrices para la inspección del pescado basada en los riesgos. Estudio FAO alimentación y nutrición 90. Organización de las Naciones Unidas para la Agricultura y la Alimentación (FAO). Available at: http://www.fao.org/3/a-i0468s.pdf.
- Food and Agriculture Organization (FAO) (2020). El estado mundial de la pesca y la acuicultura 2020. La sostenibilidad en acción. Roma. Organización de las Naciones Unidas para la Agricultura y la Alimentación (FAO). Available at: https://doi.org/10.4060/ca9229es. http://www.fao.org/3/ca9229es/ca9229es.pdf.
- Food and Drugs Administration (FDA) (2018). Foodborne Pathogens Resources for Medical Professionals from Food Safety for Moms to Be. U.S. Food and Drugs Administration. Available at: https://www.fda.gov/food/people-risk-foodborne-illness/foodbornepathogens-resources-medical-professionals-food-safety-moms-be.
- Fuente Salcido NM, Barboza Corona JE (2010). Inocuidad y bioconservación de alimentos. Acta Universitaria 20(1):43-52. Available at: https://www.redalyc.org/pdf/416/41613084005.pdf.
- Fuentes M, Valladares J, Grass G, Pico Y (2011). Microbial of the public health interest of Oreochromis spp. (Red tilapia) cultured in fresh water floating cages. Revista Cubana de Investigaciones Pesqueras 28(2):74-80. Available at: https://www.oceandocs.org/bitstream/handle/1834/4662/Mayel%edn. pdf?sequence=1.
- Fuertes Vicente HG, Paredes López F, Saavedra Gálvez DI (2014).Good practice manufacturing and preservation on board: fish safe. Big bang faustiniano Revista Indizada de Investigación Científica Huacho, Perú 3(4):41-45. Available at: http://200.48.129.168/index.php/BIGBANG/article/view/234.
- García Ortega A, Calvario Martínez O (2008). Manual de Buenas Prácticas de Producción Acuícola de Tilapia para la Inocuidad Alimentaria. 1ra edición. Centro de Investigación en Alimentación y Desarrollo, A.C. Unidad Mazatlán en Acuicultura y Manejo Ambiental. Servicio Nacional de Sanidad, Inocuidad y Calidad Agroalimentaria (SENASICA). Gobierno de México. Available at: https://www.gob.mx/cms/uploads/attachment/file/167794/7_Manual_ Tilapia.pdf.
- Garitano IG, Naranjo M, Forier A, Hendriks R, De Schrijver K, Bertrand S, Quoilin S (2011). Shigellosis outbreak linked to canteen-food consumption in a public institution: a matched case-control study. Epidemiology and Infection 139(12):1956-1964. Available at: DOI: https://doi.org/10.1017/S0950268810003110
- Guchi B, Ashenafi M (2010). Microbial load, prevalence and antibiograms of Salmonella and Shigella in lettuce and green peppers. Ethiopian Journal of Health sciences 20(1):41-8. Available at: DOI: 10.4314/ejhs.v20i1.69431
- Guzman-Herrador B, Vold L, Comelli H, MacDonald E, Heier BT, Wester AL, Stavnes TL, Jensvoll L, Lindegård Aanstad A, Severinsen G, Aasgaard Grini J, Werner Johansen Ø, and Cudjoe K, Nygård K. (2011). Outbreak of *Shigella sonnei* infection in Norway linked to consumption of fresh basil. Eurosurveillance 16(44):pii=20007. Available at: https://doi.org/10.2807/ese.16.44.20007-en

Hernández Urzúa MA (2016). Microbiología de alimentos. Fundamentos y aplicaciones en ciencias de la salud. Editorial Médica Panamericana. México D.F. P 232.

- Hosseini Nave H, Mansouri S, Sadeghi A, Moradi M (2016). Molecular diagnosis and anti-microbial resistance patterns among *Shigella* spp. isolated from patients with diarrhea. Gastroenterology and hepatology from Bed to Bench 9(3):205–210. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4947135/
- Huertas-Caro C, Urbano-Cáceres E, Torres-Caycedo M (2019). Molecular diagnosis: an alternative for the detection of pathogens in food. Revista habanera de Ciencias Médicas 18(3):513-528.

Available

http://www.revhabanera.sld.cu/index.php/rhab/article/view/2352

- Huss HH (1997). Aseguramiento de la calidad de los productos pesqueros. FAO Documento Técnico de Pesca. No. 334. Roma, FAO. 1997. 174p. Available at: http://www.fao.org/3/t1768s/T1768S00.htm#TOC
- Huss HH (1999). El Pescado Fresco: Su Calidad y Cambios de su Calidad. FAO documento técnico de
- pesca 348. Organización de las Naciones Unidas para la Agricultura y la Alimentación. Laboratorio Tecnológico. Ministerio de Pesca. Dinamarca. Available at: http://www.fao.org/3/v7180s/v7180s00.htm#Contents
- Instituto Nacional de Pesca (INP) (2018). Acuacultura| Tilapia. Acuacultura comercial. Instituto Nacional de Pesca (INP). Gobierno de México. Available at: https://www.gob.mx/inapesca/acciones-yprogramas/acuacultura-tilapia.
- Ishrat Rahman, Shahamat M, Chowdhury MAR, Colwell RR (1996). Potential Virulence of Viable but Nonculturable *Shigella dysenteriae* Type 1. Applied and Environmental Microbiology 62(1):115–120. Available at: https://aem.asm.org/content/aem/62/1/115.full.pdf.
- ISO 21567:2004 Microbiology of food and animal feeding stuffs Horizontal method for the detection of *Shigella* spp. ISO -International Organization for Standardization.
- Izquierdo Hernández A, Armenteros Borrell M, Lancés Cotilla L, Martín González I (2004). Alimentación saludable. Revista Cubana de Enfermería 20(1):1. Available. at: http://scielo.sld.cu/scielo.php?script=sci_arttext&pid=S0864-03192004000100012&Ing=es&tIng=es.
- Jácome J, Quezada Abad Č, Sánchez Romero O, Pérez JE, Nirchio M (2019). Tilapia in Ecuador: paradox between aquaculture production and the protection of Ecuadorian biodiversity. Revista Peruana de Biología 26(4):543-550. Available at: https://dx.doi.org/10.15381/rpb.v26i4.16343.
- Jorquera D, Galarce N, Borie C (2015). The challenge of controlling foodborne diseases: bacteriophages as a new biotechnological tool. Revista Chilena de Infectología 32(6):678-688. Available at: https://dx.doi.org/10.4067/S0716-10182015000700010.
- Kopper G, Calderón G, Schneider S, Domínguez W, Gutiérrez G, Rosell C, Mejía D (2009). Enfermedades transmitidas por alimentos y su impacto socioeconómico. Informe de la Organización de las Naciones Unidas para la agricultura y la alimentación. Roma, Italia: FAO 6:1-194.Available at: http://www.proyectosame.com/brotes/Apoyo/Bibliograf%C3%ADa/26.
- %20Enfermedades%20transmitidas%20por%20alimentos..pdf. Lampel KA (2009). Shigella. Pathogens and Toxins in Foods: Challenges and Interventions pp. 131-145. ASM Press. Available at: DOI: 10.1128/9781555815936.
- León-Ramírez S (2001). Shigelosis (disentería bacilar). Salud en Tabasco 7(3):20-23. Available at: https://www.redalyc.org/articulo.oa?id=487/48770306
- Lin WS, Cheng CM, Van KT (2010). A quantitative PCR assay for rapid detection of *Shigella* species in fresh produce. Journal of Food Protection 73(2):221-233. Available at: https://doi.org/10.4315/0362-028X-73.2.221.
- Lluque A, Mosquito S, Gomes C, Riveros M, Durand D, Tilley DH, Bernal M, Prada A, Ochoa TJ, Ruiz J (2015). Virulence factors and mechanisms of antimicrobial resistance in Shigella strains from periurban areas of Lima (Peru). International Journal of Medical Microbiology 305(4-5):480-490. Available at: https://doi.org/10.1016/j.ijmm.2015.04.005.
- López Aday D, Rivero Álvarez E, Martínez Torres A, Alegret Rodríguez M (2013). Foodborne diseases in Villa Clara. Revista Cubana de Higiene y Epidemiología 51(2):203-213. Available at: http://scielo.sld.cu/scielo.php?script=sci_arttext&pid=S1561-30032013000200009.
- López Cabo M, L Romalde J, Simal-Gandara J, Gago Martínez A, Giráldez Fernández J, Bernárdez Costas M, Rodríguez Herrera J (2020). Identification of Emerging Hazards in Mussels by the Galician Emerging Food Safety Risks Network (RISEGAL). A First Approach. Foods 9(11):1641. Available at: doi:10.3390/foods9111641.
- Madigan MT, Martinko JM, Parker J (2004). Brock. Biología de los microorganismos. 10 décima edición. Pearson Educación, S.A.

509

Madrid P 1096.

Mindy JP, Gloria A, Cheryl B, John E, Richard F, Joan SK, Tanja P, Joy W, Scott FD (2004). Manual de Laboratorio para la Identificación y Prueba de Susceptibilidad a los Antimicrobianos de Patógenos Bacterianos de Importancia para la Salud Pública en el Mundo en Desarrollo. Haemophilus influenzae, Neisseria meningitidis, Streptococcus pneumoniae, Neisseria gonorrhoeae, Salmonella serotipo Typhi, Shigella y Vibrio cholerae. Centros para el Control y la Prevención de Enfermedades: Centro Nacional para las Enfermedades Infecciosas y Organización Mundial de la Salud, Enfermedades Transmisibles: Vigilancia y Respuesta. Organización (ÓMS). Mundial Salud de la Available at: https://www.who.int/drugresistance/infosharing/WHO-

CDS_CSR_RMD_2003_6_Manual_Laboratorio.pdf.

- Mokhtari W, Nsaibia S, Gharbi A, Aouni M (2013). Real-time PCR using SYBR Green for the detection of *Shigella* spp. in food and stool samples. Molecular and Cellular Probes 27(1):53-59. Available at: https://doi.org/10.1016/j.mcp.2012.09.002.
- Morales G, Blanco L, Arias ML, Chaves C (2004). Evaluación de la calidad bacteriológica de tilapia fresca (Oreochromis niloticus) proveniente de la Zona Norte de Costa Rica. Archivos Latinoamericanos de Nutrición 54(4):433-437. Available at: http://ve.scielo.org/scielo.php?script=sci_arttext&pid=S0004-06222004000400010&Ing=es&tIng=pt.
- Nadella RK, Murugadas V, Toms Č. Joseph, Lalitha KV, Prasad MM (2017). Detection of *Shigella* spp. from seafood: Need for protocol harmonization. Fishtech Report 3(2):20-23. Available at: http://14.139.191.134/bitstream/handle/123456789/3950/Detection% 20of%20shigella%20spp%20from%20seafood%20%281%29.pdf?se quence=5&isAllowed=y
- NMX-F-605-NORMEX-2016. Alimentos manejo higiénico en el servicio de alimentos preparados para la obtención del distintivo "H". Norma Mexicana. Available at: https://www.gob.mx/cms/uploads/attachment/file/197511/NMX-F-605-NORMEX-2016_7_de_diciembre_de_2015_firmada_002_.pdf
- NOM-128-SSA1-1994. Bienes y servicios. Que establece la aplicación de un sistema de análisis de riesgos y control de puntos críticos en la planta industrial procesadora de productos de la pesca. Norma oficial mexicana. Estados unidos mexicanos. Secretaría de salud. Available at: http://www.salud.gob.mx/unidades/cdi/nom/128ssa14.html
- NOM-210-SSA1-2014. Productos y servicios. Métodos de prueba microbiológicos. Determinación de microorganismos indicadores. Determinación de microorganismos patógenos. Norma Oficial Mexicana. Available at: http://dof.gob.mx/nota_detalle.php?codigo=5398468&fecha=26/06/20

ntp://doi.gob.mx/nota_detaile.pnp?codigo=5398468&iecna=26/06/20 15

NOM-230-SSA1-2002, Salud ambiental. Agua para uso y consumo humano, requisitos sanitarios que se deben cumplir en los sistemas de abastecimiento públicos y privados durante el manejo del agua. Procedimientos sanitarios para el muestreo. Norma Oficial Mexicana. Available at:

http://www.salud.gob.mx/unidades/cdi/nom/230ssa102.html

NOM-242-SSA1-2009. Productos y servicios. Productos de la pesca frescos, refrigerados, congelados y procesados. Especificaciones sanitarias y métodos de prueba. Norma Oficial Mexicana. Available at:

http://dof.gob.mx/nota_detalle.php?codigo=5177531&fecha=10/02/20 11

- NOM-251-SSA1-2009. Prácticas de higiene para el proceso de alimentos, bebidas o suplementos alimenticios. Norma Oficial Mexicana. Available at: https://www.dof.gob.mx/normasOficiales/3980/salud/salud.htm
- Noor R, Acharjee M, Ahmed T, Das KK, Paul L, Munshi SK, Alam M (2020). Microbiological study of major sea fish available in local markets of Dhaka city, Bangladesh. Journal of Microbiology, Biotechnology and Food Sciences 9(4):2420-2430. Available at: https://www.jmbfs.org/wp-

content/uploads/2013/01/jmbfs_0271_noor.pdf

Ogbonna DN, Inana ME (2018). Characterization and multiple antibiotic resistance of bacterial isolates associated with fish aquaculture in ponds and rivers in Port Harcourt, Nigeria. Journal of Advances in Microbiology 10(4):1-14. Available at: DOI:

10.9734/JAMB/2018/41073

- OMS (2007). Manual sobre las cinco claves para la inocuidad de los alimentos. Departamento de Inocuidad de los Alimentos, Zoonosis y Enfermedades de Transmisión Alimentaria. Organización Mundial de la Salud (OMS). Available at: https://www.who.int/foodsafety/publications/consumer/manual_keys_ es.pdf?ua=1.
- OMS/FAO (2016). Cinco claves para una mayor inocuidad de los productos de acuicultura con objeto de proteger la salud pública. Organización Mundial de la Salud (OMS). Organización de las Naciones Unidas para la Alimentación y la Agricultura (FAO). Available at:

https://apps.who.int/iris/bitstream/handle/10665/251672/9789243510 316-

spa.pdf;jsessionid=65319AE2383229CDD1355E02542F3E53?seque nce=1.

Onyango DM, Wandili S, Kakai R, Waindi EN (2009). Isolation of Salmonella and Shigella from fish harvested from the Winam Gulf of Lake Victoria, Kenya. Journal Infect Developing Countries 3(2):99-104. Available at:

http://edocs.maseno.ac.ke/handle/123456789/2478.

- Paauw A, Jonker D, Roeselers G, Heng JM, Mars-Groenendijk RH, Trip H, Majchrzykiewicz-Koehorst JA (2015). Rapid and reliable discrimination between *Shigella* species and *Escherichia coli* using MALDI-TOF mass spectrometry. International Journal of Medical Microbiology 305(4-5):446-452. Available at: http://dx.doi.org/10.1016/j.ijmm.2015.04.001.
- PAHO (2020). Buenas prácticas: Control de Operaciones. Pan American Health Organization (PAHO). World Health Organization (WHO). Available at: https://www.paho.org/hq/index.php?option=com_content&view=articl

e&id=10564:2015-buenas-practicas-control-

operaciones&Itemid=42210&lang=en.

- Palomino-Camargo C, González-Muñoz Y (2014). Molecular techniques for detection and identification of pathogens in food: advantages and limitations. Revista Peruana de Medicina Experimental y Salud Pública 31:535-546. Available at: https://www.scielosp.org/article/rpmesp/2014.v31n3/535-546/.
- Palomino-Camargo C, González-Muñoz Y, Pérez-Sira E, Aguilar VH (2018). Delphi methodology in food safety management and foodborne disease prevention. Revista Peruana de Medicina Experimental y Salud Pública 35:483-490. Available at: https://doi.org/10.17843/rpmesp.2018.353.3086.
- Pelczar Jr. MJ, Reid DR, Chan ECS (1982). Microbiología. 4ta edición. Mc Graw-Hill.
- Pichel M, Brengi SP, Cooper KL, Ribot EM, Al-Busaidy S, Araya P, Nielsen EM (2012). Standardization and international multicenter validation of a PulseNet pulsed-field gel electrophoresis protocol for subtyping Shigella flexneri isolates. Foodborne Pathogens and Disease 9(5):418-424. Available at: https://doi.org/10.1089/fpd.2011.1067
- Pichel M, Caffer MI (2016). Parte II.c.1 Enterobacterias. Capítulo II.c.1.2 Shigella spp. Manual de microbiología clínica de la asociación argentina de microbiología volumen I Bacterias de ImportanciaClínica. Editores
- Horacio A. Lopardo, Silvia C. Predari, Carlos Vay. Ciudad Autónoma de Buenos Aires. Asociación Argentina de Microbiología. Available at: https://aam.org.ar/manual%20bacteriologia%20clinica.pdf.
- Polifroni R, Etcheverría AI, Padola NL, Parma AE (2009). Verocytotoxigenic Escherichia coli (VTEC). Characteristics of virulence and persistence in the environment. InVet 11(1):65-70. Available

https://digital.cic.gba.gob.ar/bitstream/handle/11746/4441/11746_444 1.pdfPDFA.pdf?sequence=1&isAllowed=y.

- Prats G, Mirelis B (1998). Género Shigella: aspectos prácticos para el laboratorio de microbiología. Servei de Microbiología, Hospital de Sant Pau, Barcelona. Available at: https://www.seimc.org/contenidos/ccs/revisionestematicas/bacteriolo gia/rshigella.pdf.
- Puerta-García A, Mateos-Rodríguez F (2010). Enterobacterias. Medicine.10(51):3426-31.

Radhika M, Saugata M, Murali HS, Batra HV (2014). A novel multiplex

PCR for the simultaneous detection of Salmonella enterica and *Shigella* species. Brazilian Journal of Microbiology 45(2):667-676. Available at: https://doi.org/10.1590/S1517-83822014005000041

- Ragupathi ND, Sethuvel DM, Inbanathan FY, Veeraraghavan B (2018). Accurate differentiation of *Escherichia coli* and *Shigella serogroups*: challenges and strategies. New Microbes and New Infections 21:58-62. Available at: https://doi.org/10.1016/j.nmni.2017.09.003
- Ranjbar R, Afshar D, Mehrabi Tavana A, Najafi A, Pourali F, Safiri Z, Sorouri Zanjani R, Jonaidi Jafari N (2014). Development of Multiplex PCR for Simultaneous Detection of Three Pathogenic Shigella Species. Iranian Journal of Public Health 43(12):1657-1663. Available

https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4499087/.

- Regulation (EC) No 178/2002 of the European parliament and of the council of 28 January2002 laying down the general principles and requirements of food law, establishing the European Food Safety Authority and laying down procedures in matters of food safety. Official Journal of the European Communities. Available at: https://eur-lex.europa.eu/legal
 - content/EN/TXT/PDF/?uri=CELEX:32002R0178&from=ES.
- Regulation (EC) No 2073/2005 of 15 November 2005 on microbiological criteria for foodstuffs. Official Journal of the European Union. Available at: https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32005R2073&from=ES.
- Regulation (EC) No 852/2004 of the European parliament and of the council of 29 April 2004 on the hygiene of foodstuffs. Official Journal of the European Union. Available at: https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32004R0852&from=ES
- Regulation (EC) No 853/2004 of the European parliament and of the council of 29 April 2004 laying down specific hygiene rules for on the hygiene of foodstuffs. Official Journal of the European Union. Available at: https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32004R0853&from=ES
- Regulation (EC) No 854/2004 of the European parliament and of the council of 29 April 2004 laying down specific rules for the organization of official controls on products of animal origin intended for human consumption. Official Journal of the European Union. Available at: https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32004R0854&from=ES
- Regulation (EC) No 882/2004 of the European parliament and of the council of 29 April 2004 on official controls performed to ensure the verification of compliance with feed and food law, animal health and animal welfare rules. Official Journal of the European Union. Available at: https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32004R0882&from=es
- Rodríguez Torrens H, Barreto Argilagos G, Sedrés Cabrera M, Bertot Valdés J, Martínez Sáez S, Guevara Viera G (2015). The foodborne diseases, a health problem inherited and increased in the new millennium. REDVET. Revista Electrónica de Veterinaria, 16(8): 1-27. Available at: https://www.redalyc.org/pdf/636/63641401002.pdf
- Romero Cabello R (1999). Microbiología y parasitología humana. Bases etiológicas de las enfermedades infecciosas.2 edición. Editorial Medica Panamericana. p.873.
- Romero García JL, Grande Burgos MJ, Pérez Pulido R, Gálvez A, Lucas R (2015). Resistencias a biocidas de cepas aisladas de diferentes pescados. Anales de la Real Academia de Ciencias Veterinarias de Andalucía Oriental 28(1):236-256. Available at: https://helvia.uco.es/handle/10396/17082
- Romero-Jarero JM, Negrete-Redondo MP (2011). Presence of Gram negative bacteria in fish muscle of commercial importance in the Mexican Caribbean zone. Revista Mexicana de Biodiversidad 82(2):599-606. Available at: http://www.scielo.org.mx/scielo.php?script=sci_arttext&pid=S1870-34532011000200019&Ing=es&tIng=es.
- SAGARPA (2018). Atlas agroalimentario 2012-2018. Secretaria de agricultura, ganadería, desarrollo rural, pesca y alimentación (SAGARPA). Gobierno de México. Available at: https://nube.siap.gob.mx/gobmx_publicaciones_siap/pag/2018/Atlas-Agroalimentario-2018
- Schaumann R, Knoop N, Genzel GH, Losensky K, Rosenkranz C, Stîngu CS, Schellenberger W, Rodloff AC, Eschrich K (2013). Discrimination of Enterobacteriaceae and Non-fermenting Gram

Negative Bacilli by MALDI-TOF Mass Spectrometry. The open Microbiology Journal 7:118-122. Available at: https://doi.org/10.2174/1874285801307010118

SENÁSICA (2019). Manual de Buenas Prácticas Acuícolas durante la Producción Primaria de Peces. Servicio Nacional de Sanidad, Inocuidad y Calidad Agroalimentaria (SENASICA). Gobierno de México. Available at:

https://www.gob.mx/senasica/documentos/manuales-de-buenas-practicas-pecuarias-acuicolas-y-pesqueras.

- Shao Y, Zhu S, Jin C, Chen F (2011). Development of multiplex loopmediated isothermal amplification-RFLP (mLAMP-RFLP) to detect Salmonella spp. and *Shigella* spp. in milk. International Journal of Food Microbiology 148(2):75-79. Available at: https://doi.org/10.1016/j.ijfoodmicro.2011.05.004.
- Soares KMP, Gonçalves AA (2012). Seafood quality and safety. Revista do Instituto Adolfo Lutz 71(1):1-10. Available at: http://periodicos.ses.sp.bvs.br/pdf/rial/v71n1/v71n1a01.pdf
- Soto Varela Z, Pérez Lavalle L, Estrada Alvarado D (2016). Bacteria causing of foodborne diseases: an overview at Colombia. Salud Uninorte. Barranquilla 32(1):105-122. Available at: http://www.scielo.org.co/pdf/sun/v32n1/v32n1a10.pdf
- Tafur Garzón M (2009). La inocuidad de alimentos y el comercio internacional. Revista Colombiana de Ciencias Pecuarias 22(3):330-338. Available at: http://www.scielo.org.co/scielo.php?script=sci_arttext&pid=S0120

06902009000300009&Ing=en&tIng=en.

- Thiem VD, Sethabutr O, Von Seidlein L, Van Tung T, Chien BT, Lee H, Trach DD (2004). Detection of Shigella by a PCR assay targeting the ipaH gene suggests increased prevalence of shigellosis in Nha Trang, Vietnam. Journal of Clinical Microbiology 42(5):2031-2035. Available at: DOI: 10.1128/JCM.42.5.2031–2035.
- Torres AG (2004). Current aspects of Shigella pathogenesis. Revista Latinoamericana de Microbiología 46(3-4):89-97. Available at: https://www.medigraphic.com/pdfs/lamicro/mi-2004/mi04-3 4f.pdf
- Vásquez de Plata G (2003). La contaminación de los alimentos, un problema por resolver. Revista salud UIS,35:48-57. Available at: https://revistas.uis.edu.co/index.php/revistasaluduis/article/view/728
- Villacrés I, Alcocer I, Zurita J, Riglos MR (2015). Sensibilidad antimicrobiana y detección de genes de virulencia en aislados clínicos de Shigella spp. Revista Ecuatoriana de Medicina y Ciencias Biológicas 36(1-2):67-76. Available at: DOI: https://doi.org/10.26807/remcb.v36i1-2.69
- Villalobo E, Torres A (1998). PCR for detection of *Shigella* spp. in mayonnaise. Applied and Environmental Microbiology 64(4):1242-1245. Available at: https://aem.asm.org/content/aem/64/4/1242.full.pdf
- Wallace HA, Andrew J (2018). Shigella. Chapter 6. Bacteriological Analytical Manual (BAM). U.S. Food and Drugs Administration (FDA). Available at: https://www.fda.gov/food/laboratory-methods-food/bamchapter-6-shigella
- Wang RF, Cao WW, Cerniglia CE (1997). A universal protocol for PCR detection of 13 species of foodborne pathogens in foods. Journal of Applied Microbiology 83:727-736. Available at: https://sfamjournals.onlinelibrary.wiley.com/doi/pdf/10.1046/j.1365-2672.1997.00300.x
- Wen-Bin H, Jiann-Haur W, Pei-Chun C, Yu-Shiou L, Jiann-Hwa C (2007). Detecting low concentrations of *Shigella sonnei* in environmental water samples by PCR. FEMS Microbiology Letters 270(2): 291–298. Available at:https://doi.org/10.1111/j.1574-6968.2007.00686.x
- World Health Organization(WHO) (2020a). Foodborne diseases. The World Health Organization(WHO). Available at: https://www.who.int/health-topics/foodborne-diseases#tab=tab_1
- World Health Organization (WHO) (2020). Food safety. Infographics: Estimates of the global burden of foodborne diseases. The World Health Organization(WHO). Available at: https://www.who.int/foodsafety/areas_work/foodbornediseases/ferg_infographics/en/
- World Health Organization (WHO) (2020b). Food safety. The World Health Organization (WHO). Available at: https://www.who.int/news-room/fact-sheets/detail/food-safety.
- WHO/FAO (2020). Codex Alimentarius. International food standards.

World Health Organization (WHO). Food and Agriculture Organization of the United Nations (FAO). Codex Alimentarius. Available at: http://www.fao.org/fao-who-codexalimentarius/codex-texts/codes-of-practice/en/