ISSN 1991-637X ©2011 Academic Journals

### Full Length Research Paper

# Effect of *Astragalus membranaceus* polysaccharides on oxidative damage in skeletal muscle of exhaustive exercise rats

## Zhi-hong Deng<sup>1\*</sup> and Qing-lan Hu<sup>2</sup>

<sup>1</sup>Kunming University, Kunming, Yunnan Province, 650214, People's Republic of China. <sup>2</sup>Kunming University of Science and Technology, Kunming, Yunnan Province, 650224, People's Republic of China.

Accepted 11 July, 2011

The aim of this study was to determine the effects of *Astragalus membranaceus* polysaccharides (AMP) on oxidative damage in skeletal muscle of exhaustive exercise rats. The animals were divided into four groups at random, with eight rats in each group. Group 1, assigned as a control group, received normal saline (vehicle); groups 2 to 4 received AMP at different dosages (50, 100 and 200 mg/kg bw) respectively for 30 successive days. Rats were made to perform an exhaustive running test on a treadmill at a final speed of 30 m/min, 10% grade, at approximately 70 to 75% VO<sub>2</sub>max. The mean endurance times of running on a treadmill to exhaustion of each rat were examined. And skeletal muscle samples were collected for determining the activity of superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), catalase (CAT) and levels of malondialdehyde (MDA). The results showed groups 2 to 4 significantly increased in exhaustive exercise time compared with groups 1. MDA concentration of skeletal muscle of the groups 2 to 4 were significantly decreased compared with that of the groups 1. The antioxidant enzyme activities of SOD, GSH-Px and CAT of skeletal muscle of the groups 2 to 4 significantly increased compared with that of the groups 1. It is concluded that AMP supplementation increased performance of exhaustive exercise of rats. And it is beneficial for increasing antioxidant status and inhibits oxidative stress induced by exhaustive exercise.

Key words: Astragalus membranaceus polysaccharides, oxidative damage, exhaustive exercise, rats.

#### INTRODUCTION

During physical exercise, oxygen flux to active skeletal muscles increases, which leads to enhanced production of reactive oxygen species (ROS) and free radicals (Chiaradia et al., 1998). Production of ROS in quantities that overwhelm the endogenous antioxidant defense system has been referred to as oxidative stress (Bloomer et al., 2006; Goldfarb et al., 2007). Regular physical exercise has beneficial effects on health, and acute exhaustive exercise may attenuate these benefits via the induction of oxidative stress, which if increased can be harmful to all cellular macromolecules, such as lipids, proteins, and DNA (Niu et al., 2008). The supplementation

of exogenous antioxidants may provide support for the endogenous antioxidant defense system to assist in the handling of the increased production of ROS (Aguiló et al., 2005).

The roots of Astragalus membranaceus (AM) are one of the most popular health-promoting plants in East Asia, and have been used as herbs for more than 2000 years (Lee et al., 2007). The main constituents of AM roots are polysaccharides, saponins, flavonoids, amino acids, and trace elements (Zheng et al., 1998; Ma et al., 2002). Recently AMP is reported to have antioxidant, antidiabetic, anti-hypertensive, and immunomodulatory activities (Wu et al., 2005; Bian and Li, 2009; Zhang et al., 2009; Yang et al., 2009), but few researches have been reported on the effects of AMP on exercise function. In this study, we investigated the effects of AMP on oxidative damage in skeletal muscle of exhaustive

<sup>\*</sup>Corresponding author. E-mail: zh\_deng95@yahoo.cn. Tel: +086-0871-5820360. Fax: +086-0871-5350648.

exercise rats.

#### **MATERIALS AND METHODS**

#### Reagents

SOD, GSH-Px, CAT and MDA assay kits were purchased from the Nanjing Jiancheng Bioengineering Institute (Nanjing, China); Other chemicals were of analytical-reagent grade and were purchased from Kunming Chemical Reagent Co. (Yunnan, China), unless otherwise stated.

#### **Experimental animals**

Male inbred Sprague-Dawley strain rats, 6 to 8 weeks old, were raised under controlled environment in the institute's animal house at 25  $\pm$  1°C and 12 h light-dark cycle. The animals were fed standard animal food, that is, pellet and water ad libitum. All the experiments were performed based on the regulations specified by the Institutional Animal Ethical Committee (IAEC) and conformed to national guidelines on the care and use of laboratory animals, China.

# Plant materials and preparation of *A. membranaceus* polysaccharides

The roots of A. membranaceus (AM) were purchased from Kunming herb market (Yunnan, China), and identified by Professor Yang of Kunming University of Science and Technology. Voucher specimens (KMY-DK2073) were preserved in Yunnan Natural Product Research Institute. AMP was prepared by the method of Sun et al. (2005). The dried roots of AM were ground to fine powder, and then extracted with hot water (65°C) for three times (1:35, w/v). The extract was concentrated in a rotary evaporator under reduced pressure, precipitated by 95% (v/v) ethanol at 4℃ for 24 h, and then centrifuged (10 min, 3000 rpm). The sediment was washed with ethanol, acetone, and ether alternately for three times to exclude lipid groups. The precipitate was vacuum freezedried, and crude AMP were obtained, which was dissolved in water and discolored by H<sub>2</sub>O<sub>2</sub>, then proteins were removed by the sevage method (Zhang et al., 2006). Furthermore, this crude polysaccharide was suspended in water, dialyzed to eliminate small molecules to obtain the pure polysaccharide, which was collected and precipitated with 95% (v/v) ethanol and then lyophilized. The yield of AMP was 2.07%.

#### **Exercise protocol**

The animals were divided into four groups at random, with eight rats in each group. Group 1, assigned as a control group, received 1.5 ml of sterile normal saline (vehicle) for 30 successive days. While groups 2 to 4 received AMP at different dosages (50, 100 and 200 mg/kg bw) respectively for 30 successive days, The AMP were redissolved in 1.5 ml of sterile normal saline and administered orally by a canule.

Before exercise test, the rats were introduced to running on a treadmill with 15 to 20 min exercise bouts at 15 to 30 m/min for 7 days to accustom them to running (Ji et al., 1991). The treadmill was equipped with an electric shocking grid on the rear barrier to motivate the animal to run (PBC47700 treadmill device; Coulbourn Instruments Co., USA). On the day of the exercise test, they were required to run to exhaustion on a five-lane inclined (10°) treadmill at a final speed of 30 m/min, which was approximately 70 to 75% of

 $VO_2$ max (Brooks and White, 1978). The measurement of maximal  $O_2$  consumption (100%  $VO_2$  max) was considered valid only when the animal could no longer maintain pace with the treadmill (Somani et al., 1995). Therefore, exhaustion was determined as the rat being unable to upright itself when placed on its back. To eliminate diurnal effects, the experiments were performed at the same time (08.30 to 11.30 h).

#### Tissue preparation

After exhaustive exercise, rats were anaesthetized with thiopenton sodium (50 mg/kg) skeletal muscle was excised immediately and immersed in physiological saline. For the preparation of skeletal muscle homogenates (1 g of tissue plus 10 ml homogenization buffer), the frozen pieces were thawed on ice and then homogenized. The suspension was centrifuged at 671  $\times$  g at 4°C for 10 min and clear supernatant was used for the following estimations of activity of SOD, GSH-Px, CAT and levels of MDA.

#### Assay of oxidation products and enzymatic antioxidants

Lipid peroxidation was assessed by determining the level of MDA in skeletal muscle homogenates by using commercial diagnostic kit. Enzymatic antioxidants were assessed by determining the activity of SOD, CAT and GSH-Px by using commercial diagnostic kits.

#### Statistical analysis

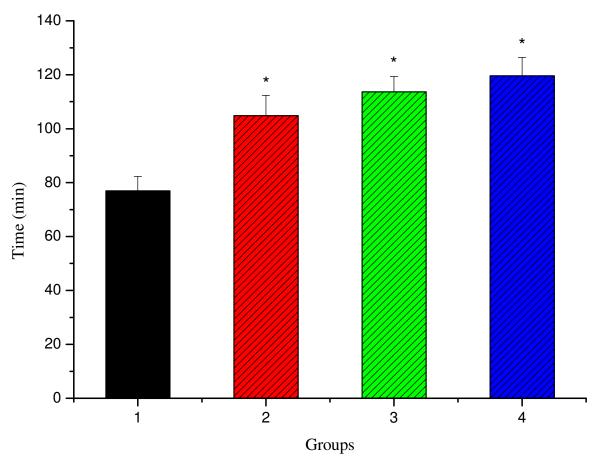
All values are presented as means  $\pm$  SE. Comparisons of data were made by one-way ANOVA. When data were significant, each group was compared with the others by Fisher's protected least significant difference test (StatView; SAS Institute, Cary, NC). Results with values of P<0.05 were considered statistically significant.

#### **RESULTS**

At the end of 30 days, body weights in the groups 1 to 4 were  $413.5 \pm 8.4$ ,  $407.8 \pm 10.5$ ,  $401.6 \pm 8.7$  and  $414.8 \pm 9.7$  g, respectively, and no significant differences were detected among the four groups. As shown in Figure 1,the mean endurance times of running on a treadmill to exhaustion in the groups 2 to 4 were significantly prolonged compared with that in the group 1 (P < 0.05). As shown in Figure 2, MDA concentration of skeletal muscle of the groups 2 to 4 were significantly decreased compared with that of the groups 1 (P < 0.05). Table 1 shows the antioxidant enzyme activities. Compared with group 1, the antioxidant enzyme activities of SOD, GSH-Px and CAT of skeletal muscle of the groups 2 to 4 were all significantly higher.

#### **DISCUSSION**

This study aimed to examine the effects of AMP on oxidative damage in skeletal muscle of exhaustive exercise rats. The laboratory rat is a commonly used model for the evaluation of exhaustive exercise effects on biochemical changes in man. In this research, the mean

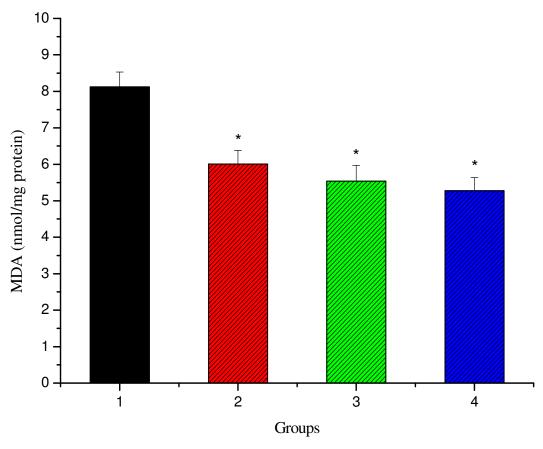


**Figure 1.** Effects of AMP on the endurance times of treadmill running to exhaustion P < 0.05, compared with the group 1. Eight rats were observed and tested for each group during the experimental period.

exhaustive exercise time of running on a treadmill in the groups 2 to 4 were significantly prolonged compared with that of the group 1 (Figure 1). It indicated that AMP supplementation could increase exhaustive time.

During exercise the need for oxygen increases. The process of delivering the oxygen to the working muscles may actually result in damage to polyunsaturated fatty acids in membrane structures (Hsieh et al., 2009), which has been documented by numerous investigations demonstrating increases in the byproducts of lipid peroxidation following exercise (Alessio and Goldfarb, 1988). When a hydroxyl radical reacts with an unsaturated fatty acid, a lipid peroxyl radical is formed. In the presence of oxygen this new free radical incites a chain of events referred to as lipid peroxidation. Lipid peroxidation of cell membranes results in decreased membrane fluidity, inability to maintain ionic gradients. cellular swelling, and tissue inflammation (Margaritis et al., 2003). MDA, a major end-product of oxidation of polyunsaturated fatty acids, has been frequently measured as an indicator of lipid peroxidation and oxidative stress in-vivo. Many studies have found acute exercise induced increases in MDA concentration in tissues. In the present research, MDA concentration of skeletal muscle tissues of the groups 2 to 4 were significantly decreased compared with that of the group 1 (Figure 2). It indicated that AMP supplementation could effectively reduce lipid peroxidation to prevent oxidative damage in rat skeletal muscle.

Cells are equipped with a host of enzymes directly or indirectly involved in the antioxidant defense against ROS, providing primary defenses such as SOD, GSH-Px, and CAT. SOD, the most important enzyme present virtually in all aerobic organisms, catalyzes the dismutation of superoxide into oxygen and hydrogen peroxide. GSH-Px is a selenoenzyme which catalyzes the reduction of hydroperoxides at the expense of reduced glutathione. CAT is a primary antioxidant defense component that works to catalyze the decomposition of hydrogen peroxide to water, sharing this function with GSH-Px (Cotgreave et al., 1988). In this research, the antioxidant enzyme activities of SOD, GSH-Px and CAT of skeletal muscle of the groups 2 to 4 were significantly increased compared with that of the group 1 (Table 1). It indicated that AMP supplementation had beneficial effects on attenuating the oxidative stress



**Figure 2.** Effects of AMP on MDA concentration \*P < 0.05, compared with the groups 1. Eight rats were observed and tested for each group during the experimental period.

Table 1. Effect of AMP on the activities of anti-oxidant enzymes in skeletal muscle of rats.

Groups	SOD (U/mg protein)	GPx (U/mg protein)	CAT (U/mg protein)
1	106.81±11.43	7.87±0.49	3.41±0.27
2	149.55±13.74*	9.01±0.93*	4.73±0.31*
3	161.62±9.91*	13.16±0.84*	5.76±0.37*
4	175.73±10.86*	15.87 ±1.19*	5.14±0.29*

<sup>\*</sup>P<0.05, compared with the groups 1. Eight rats were observed and tested for each group during the experimental period.

induced by exhaustive exercise.

In conclusion, AMP supplementation increased performance of exhaustive exercise. And it is beneficial for increasing antioxidant status and inhibits oxidative stress induced by exhaustive exercise. It still needs further investigation and pathological observation in the future study.

#### **ACKNOWLEDGEMENT**

The authors gratefully acknowledge Ms Jian Zhang for helping in editing the manuscript. The authors are also

grateful to Professor Yang MN Sebastian for her timely technical support.

#### **REFERENCES**

Aguiló A, Tauler P, Fuentespina E, Tur JA, Córdova A, Pons A (2005). Antioxidant response to oxidative stress induced by exhaustive exercise. Physiol. Behav., 84(1):1-7.

Alessio HM, Goldfarb AH (1988). Lipid peroxidation and scavenger enzymes during exercise: adaptive response to training. J. Appl. Physiol., 64(4): 1333-1336.

Bian Y, Li P (2009). Antioxidant activity of different extracts from Astragalus mongholicus. Zhongguo Zhong Yao Za Zhi, 34(22): 2924-2927 (Chinese).

- Bloomer RJ, Goldfarb AH, McKenzie MJ (2006). Oxidative stress response to aerobic exercise: comparison of antioxidant supplements. Med. Sci. Sports Exerc., 38(6): 1098-105.
- Brooks GA, White TP (1978). Determination of metabolic and heart rate responses of rats to treadmill exercise. J. Appl. Physiol., 45(6): 1009-1015.
- Chiaradia E, Avellini L, Rueca F, Spaterna A, Porciello F, Antonioni MT, Gaiti A (1998). Physical exercise, oxidative stress and muscle damage in racehorses. Comp. Biochem. Physiol. B Biochem. Mol. Biol., 119(4): 833-836.
- Cotgreave IA, Moldéus P, Orrenius S (1988). Host biochemical defense mechanisms against prooxidants. Annu. Rev. Pharmacol. Toxicol., 28: 189-212.
- Goldfarb AH, McKenzie MJ, Bloomer RJ (2007). Gender comparisons of exercise-induced oxidative stress: Influence of antioxidant supplementation. Appl. Physiol. Nutr. Metab., 32(6): 1124-1131.
- Hsieh CC, Wang VH, Liu CC, Lin TH (2009). Protective Effects of Oolong Tea on Oxidative Stress Induced by Exhaustive Exercise in Rat Muscle. J. Phys. Educ. Higher Educ., 11(3): 105-116.
- Ji LL, Dillon D, Wu E. (1991). Myocardial aging: Antioxidant enzyme systems and related biochemical properties. Am. J. Physiol., 261(2 Pt 2): R386-392.
- Lee SJ, Oh SG, Seo SW, Ahn HJ, Geum D, Cho JJ, Park CS (2007). Oral administration of Astragalus membranaceus inhibits the development of DNFB-induced dermatitis in NC/Nga mice. Biol. Pharm. Bull., 30(8): 1468-1471.
- Ma XQ, Shi Q, Duan JA, Dong TT, Tsim KW (2002). Chemical analysis of Radix Astragali (Huangqi) in China: A comparison with its adulterants and seasonal variations. J. Agric. Food Chem., 50(17): 4861-4866.
- Margaritis I, Palazzetti S, Rousseau AS, Richard MJ, Favier A (2003). Antioxidant supplementation and tapering exercise improve exercise-induced antioxidant response. J. Am. Coll. Nutr., 22(2): 147-156.

- Niu AJ, Wu JM, Yu DH, Wang R (2008). Protective effect of Lycium barbarum polysaccharides on oxidative damage in skeletal muscle of exhaustive exercise rats. Int. J. Biol. Macromol., 42(5): 447-449.
- Somani SM, Frank S, Rybak LP (1995). Responses of antioxidant system to acute and trained exercise in rat heart subcellular fractions. Pharmacol. Biochem. Behav., 51(4): 627-634.
- Sun RM, Liu DP, Chen J, Liu ZF, Liu XP, Li FX, Pu XH (2005). Extraction of Astragalus Polysaccharides in Astragalus. Acta scientiarum naturalium universitatis nankaiensis (natural science edition), 38(1): 146-148 (Chinese).
- Wu Y, Ou-Yang JP, Wu K, Wang Y, Zhou YF, Wen CY (2005). Hypoglycemic effect of Astragalus polysaccharide and its effect on PTP1B. Acta Pharmacol. Sin., 26(3): 345-352.
- Yang WJ, Li DP, Li JK, Li MH, Chen YL, Zhang PZ (2009). Synergistic antioxidant activities of eight traditional Chinese herb pairs. Biol. Pharm. Bull., 32(6): 1021-1026.
- Zhang J, Huang Y, Hou T, Wang Y (2006). Hypoglycaemic effect of Artemisia sphaerocephala Krasch seed polysaccharide in alloxan-induced diabetic rats. Swiss Med Wkly, 136(33-34): 529-532.
- Zhang J, Xie X, Li C, Fu P (2009). Systematic review of the renal protective effect of *Astragalus membranaceus* (root) on diabetic nephropathy in animal models. J. Ethnopharmacol., 126(2): 189-196.
- Zheng Z, Liu D, Song C, Cheng C, Hu Z (1998). Studies on chemical constituents and immunological function activity of hairy root of Astragalus membranaceus. Chin. J. Biotechnol., 14(2): 93-97.