

Full Length Research Paper

Effect of feeding regime on fatty acid composition and conjugated linoleic acid content of perirenal, omental and tail fat in Akkaraman lambs

Gokalp Ozmen Guler¹ and Abdurrahman Aktumsek^{2*}

¹Department of Biological Education, Ahmet Kelesoglu Education Faculty, Selcuk University, Konya, Turkey.

²Department of Biology, Science Faculty, Selcuk University, Konya, Turkey.

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In this study, the effect of feeding regime on fatty acid composition including conjugated linoleic acid (CLA) of omental, perirenal and tail fat from Akkaraman lambs, the most widespread sheep breed in central Anatolia, was investigated. Forty-five suckling lambs, born in the same farm, were fed mainly maternal milk from birth to weaning and then the lambs were divided into three groups. One group (maternal milk-fed group) of the lambs was directly slaughtered after weaning. A second group (pasture group) was allowed to graze a natural pasture and slaughtered at three months after weaning. Third group (concentrate group) was fed concentrate *ad-libitum* together with 150 g/day alfalfa and slaughtered at three months after weaning. In all feeding regime, the predominant fatty acids were C 16:0 palmitic and C 18:0 stearic acid as saturated fatty acid (SFA), C 18:1 ω 9 oleic acid as monounsaturated fatty acid (MUFA) and C 18:2 ω 6 linoleic acid as polyunsaturated fatty acid (PUFA). Omental, perirenal and tail fat of the pasture-fed lambs contained more total CLA, total ω 3, ω 3/ ω 6 ratio compared with that of the concentrate-fed lambs. Moreover, omental, perirenal and tail fat of concentrate-fed lambs had higher ω 6/ ω 3 ratio and this ratio was decreased by pasture feeding.

Key words: Akkaraman lambs, pasture, suckling, concentrate, fatty acid composition, conjugated linoleic acid.

INTRODUCTION

Conjugated linoleic acid (CLA) is a collective term for different positional and geometric isomers of octadecadienoic acid and naturally occurring fatty acid found in ruminant fats. Two of the isomers (c9,t11 and t10,c12) are known to possess biological activity (Pariza et al., 2001). The major CLA isomer, C 18:2 c9,t11, is produced in the rumen during the microbial

biohydrogenation of dietary C 18:2 ω 6 linoleic acid and in the tissues through Δ 9 desaturation of C 18:1t11 (Griinari and Bauman, 1999). The c9,t11 CLA isomer appears to be the most biologically active isomer and accounts for more than 80% of CLA in ruminant products (Ha et al., 1990). There has been much interest in CLA because of its potential health benefits such as anti-carcinogenic, antiobesity, antidiabetogenic, antiatherogenic and antioxidative properties (Ha et al., 1990; Ip et al., 1994; Lee et al., 1994; Pariza et al., 1996; Parodi, 1997; Whigham et al., 2000; Kritchevsky, 2003; Park and Pariza, 2007).

Fatty acid composition of muscle and adipose tissues can be affected by many factors such as diet, breed, age of slaughter, fatness, body weight and sex (Kemp et al., 1981; Enser, 1991; Aharoni et al., 1995; Rule et al., 1995; Wood and Enser, 1997; Nürnberg et al., 1998; Mahgoub

*Corresponding author. E-mail: aktumsek@selcuk.edu.tr. Tel: +90 332 223 18 66. Fax: +90 332 241 01 06.

Abbreviations: CLA, Conjugated linoleic acid; SFA, saturated fatty acid; MUFA, monounsaturated fatty acid; PUFA, polyunsaturated fatty acid; FID, flame ionization detector; FAME, fatty acid methyl ester; GC, gas chromatography; TFA, *trans* fatty acid.

Table 1. Ingredients and chemical composition of the concentrate feed.

Ingredient	Percentage (%)
Corn	50
Bran	18.2
Soybean meal	4
Sodium chloride	1
Sunflower seed meal	21.6
Vegetable oil	2.15
Marble powder	2.8
Vitamin and mineral premix	0.25
Chemical composition	
Moisture	8.3
Ash	6.96
Crude protein	14.14
Starch	41.41
Crude fat	4.5
Crude fiber	9.81
Calculated metabolizable energy (kcal/kg)	2505

et al., 2002; Oriani et al., 2005; Demirel et al., 2006; Arousseau et al., 2007a; Serra et al., 2009). The nutritional value of ω 3 polyunsaturated fatty acids (PUFAs) in the human diet is well recognized and an increased consumption of these fatty acids has been recommended (Department of Health, 1994). Increasing the intake of ω 3 PUFA appears to lower the risk of platelet aggregation and blood clotting, therefore decreasing the risk of thrombosis (Vanschoonbeek et al., 2003). Diet has been shown to be one of the main factors influencing fatty acid composition of fat in lambs (Wood et al., 2004). Grass-fed lambs display higher ω 3 PUFA levels, while the proportion of ω 6 PUFA increases in those fed with concentrate (Mitchell et al., 1991). Pasture feeding enhances the CLA in the tissue lipids of beef cattle (Dannenberger et al., 2005). Nutritional guidelines recommend a higher consumption of n-3 PUFA, suggesting a n-6/n-3 ratio at 4/1 or lower for the total diet (Department of Health, 1994). So, it is necessary to increase the ω 3 unsaturated fatty acids and CLA with different feeding regimes for human health.

There is no information about the effect of feeding regime (maternal milk, pasture and concentrate) on CLA content, which is important for human health and fatty acid composition of omental, perirenal and tail fat from Akkaraman lambs. Akkaraman is the most widespread sheep breed in central Anatolia and accounts for 40 to 50% of sheep population in Turkey (Akman et al., 2001).

Akkaraman sheep is one of the fat-tailed breeds and approximately 87% of the sheep population in Turkey is fat-tailed breeds (Anonymous, 2000).

The objective of the study was to characterize the effects of different feeding regime on fatty acid composition of omental, perirenal and tail fat, especially ω 3 fatty acids and CLA, of Akkaraman lambs.

MATERIALS AND METHODS

Animals and feeding regime

Forty-five male Akkaraman suckling lambs, born in the same farm, were fed mainly maternal milk and a small amount of lamb starter during first three months from birth to weaning and then the suckling lambs were divided into three equal groups, each of 15 heads, with an average live weight of 25 kg. Determination of differences of fatty acid composition in lambs fed on three different feed was aimed. So, one group of the suckling lambs (only maternal milk-fed group) was directly slaughtered after weaning. After one week of adaptation period, another group of the suckling lambs was allowed to graze everyday on natural pasture (pasture group) from weaning to slaughter. These lambs were slaughtered at three months after weaning. A third group (concentrate group) was fed concentrate *ad libitum* together with 150 g/day alfalfa per lamb from weaning to slaughter. These lambs were slaughtered at three months after weaning. Concentrate-fed lambs were reared in "Selcuk University Agriculture Faculty, Department of Animal Science Prof. Dr. Orhan Düzgüneş Research and Application Farm". Ingredients and fatty acid composition of the concentrate feed are presented in Tables 1 and 2, respectively. Chemical composition of the feed sample was analyzed using the Weende-method.

Sampling

After slaughtering, carcasses were immediately transferred to cooler at 4°C. After 24 h conservation period, 10 g omental, perirenal and tail fat samples were collected from each carcass. Samples were vacuum packaged, frozen and stored at -27°C until analysis.

Fatty acid analyses

Total lipids of lambs were extracted with chloroform/methanol (2:1 v/v) according to Folch et al. (1957) method. Methyl esters were prepared by transmethylation, using KOH 2 mol/l in methanol and n-heptane, according to method 5509 of the ISO (1978).

The fatty acid methyl esters were analyzed on a HP Agilent 6890N model gas chromatograph (GC), equipped with a flame ionization detector (FID) and fitted with a HP-88 capillary column (100 m, 0.25 mm i.d. and 0.2 μ m). Chromatographic conditions were performed according to Ledoux et al. (2005) method modified as follows: injector and detector temperatures were 250 and 280°C, respectively. The oven was programmed at 60°C initial temperature and 1 min initial time. Thereafter, the temperature increased at 20°C /min to 190°C held for 60 min then increased at 1°C/min to 220°C and held for 10 min at 220°C. Total run time was 107.5 min. The carrier gas was helium (1 ml/min).

Identification of fatty acids and *trans* isomers were carried out by comparing sample fatty acid methyl ester (FAME) peak relative

Table 2. Fatty acid composition of concentrate feed ^a.

Fatty acid	Percentage (%)
C 14:0	0.08 ± 0.01 ^b
C 15:0	0.03 ± 0.00
C 16:0	12.33 ± 0.03
C 17:0	0.09 ± 0.02
C 18:0	3.18 ± 0.03
C 20:0	0.47 ± 0.08
Σ SFA ^c	16.18 ± 0.09
C 16:1ω-7	0.11 ± 0.02
C 17:1ω-8	0.07 ± 0.01
C 18:1ω-9	26.80 ± 0.07
C 20:1ω-9	0.35 ± 0.03
Σ MUFA ^c	27.32 ± 0.03
C 18:2ω-6	51.81 ± 0.15
C 18:3ω-6	0.27 ± 0.05
C 18:3ω-3	4.02 ± 0.01
C 20:5ω-3	0.24 ± 0.04
C 22:5ω-6	0.04 ± 0.01
C 22:5ω-3	0.14 ± 0.01
Σ PUFA ^c	56.51 ± 20.15
Σ ω-3	4.40 ± 0.05
Σ ω-6	52.12 ± 0.11
ω-3/ω-6	0.08 ± 0.00
ω-6/ω-3	11.85 ± 0.14

^aAverage of three lots analyzed; ^bvalues reported are mean ± SD. ^cSFA: Saturated fatty acid; MUFA: monounsaturated fatty acid; PUFA: polyunsaturated fatty acid.

retention times with those obtained from Alltech, Nu-Check Prep. Inc. USA and Accu standards. Linoleic acid conjugated methyl ester (mixture of *cis*- and *trans*-9,11- and -10,12-octadecadienoic acid methyl esters, catalog number O5632) was purchased from Sigma-Aldrich (St Louis, MO, USA). Results were expressed as FID response area relative percentages. Each reported result is the average value of three GC analyses. The results were presented as mean ± SD.

Statistical analysis

The results were submitted to analysis of variance (ANOVA), at 0.05 significance level, using SPSS 10.0 for Windows. The mean values were compared with Duncan test.

RESULTS AND DISCUSSION

Slaughter traits and total lipid levels in omental, perirenal and tail fat are presented in Tables 3 and 4, respectively. Fatty acid composition of omental, perirenal and tail fat from lambs of the three groups are presented in Tables 5, 6 and 7, respectively.

In all feeding regime, the predominant fatty acids in perirenal, omental and tail fat were C 16:0 palmitic acid and C 18:0 stearic acid as saturated fatty acid (SFA), C 18:1ω9 oleic acid as monounsaturated fatty acid (MUFA) and C 18:2ω6 linoleic acid as PUFA. These results are similar to those reported by Ünsal and Aktaş (2003), Ünsal and Yanlıc (2005) and Yılmaz and Karakaya (2010) on the fatty acid composition of sheep tail fat and intestinal fat. Moharrey (2007) have also reported that, three major fatty acids in omental fat of fat-tailed Badghisian sheep were palmitic acid, stearic acid and oleic acid. Osorio et al. (2009) have also reported similar results for fatty acid composition of omental and perirenal fat of suckling lambs reared on ewe's milk.

Total SFA was 54.67, 48.95 and 63.75% of total fatty acids in omental fat, 55.48, 49.29 and 61.06% of total fatty acids in perirenal fat and 47.42, 38.82 and 47.49% of total fatty acids in tail fat from maternal milk, concentrate and pasture-fed lambs, respectively. Arana et al. (2006) have also reported similar values of total SFA (49.68%) for perirenal depots of lambs fed a concentrate diet. In the present study, total SFA in maternal milk, concentrate and pasture-fed lamb's omental and perirenal fat was significantly affected by feeding regime ($P < 0.05$). Additionally, total SFA in tail fat from concentrate-fed lambs was significantly lower than those of maternal milk and pasture-fed lambs ($P < 0.05$). Palmitic acid, stearic acid and myristic acid (C 14:0) were the major SFA from maternal milk, pasture and concentrate-fed lambs in omental, perirenal and tail fat. These results agree with those reported by Ünsal and Aktaş (2003), Ünsal and Yanlıc (2005) and Yılmaz and Karakaya (2010) for tail fat, Moharrey (2007) and Osorio et al. (2009) for omental fat and Arana et al. (2006) and Osorio et al. (2009) for perirenal fat who reported that palmitic acid, stearic acid and myristic acid were major SFA in sheep and lambs. In the present study, total SFA, palmitic acid, stearic acid, myristic acid and lauric acid (C 12:0) in omental, perirenal and tail fat of pasture-fed lambs were higher than those of concentrate-fed lambs. Nuernberg et al. (2008) have also reported that total SFA, palmitic acid, stearic acid, myristic acid and lauric acid in tail fat of male Skudde lambs fed pasture were higher than those of concentrate-fed lambs.

Total MUFA was 35.51, 34.16 and 30.40% of total fatty acids in omental fat, 35.27, 34.01 and 27.47% of total fatty acids in perirenal fat and 42.86, 45.34 and 42.43% of total fatty acids in tail fat from maternal milk, concentrate and pasture-fed lambs, respectively. In previous studies, total MUFA have been determined as 30.33% in perirenal fat of alpaca reared under a traditional un-specialized production system at the Andean region of Peru (Salva et al., 2009), 36.74% in omental fat deposit of suckling lambs reared on ewe's milk (Osorio et al., 2009), 42.16% in tail fat of Akkaraman sheep (Yılmaz and Karakaya, 2010) and 47.93% in tail fat of Akkaraman

Table 3. Slaughter traits in the three groups.

Slaughter trait	Maternal milk group (n=15) mean \pm SE*	Concentrate group (n=15) mean \pm SE*	Pasture group (n=15) mean \pm SE*
Age at slaughter (days)	90	180	180
Live weight at slaughter (kg)	25.18 \pm 0.75	46.08 \pm 0.75	35.07 \pm 0.55
Hot carcass weight (kg)	12.44 \pm 0.49	24.05 \pm 1.60	17.20 \pm 0.94

*SE, Standard error of the mean.

Table 4. Total lipid levels in omental, perirenal and tail fat of Akkaraman lambs.

Group	Total lipid (%)		
	Omental fat, mean \pm SD*	Perirenal fat, mean \pm SD	Tail fat, mean \pm SD
Maternal milk-fed group	66.1 \pm 1.19	67.1 \pm 1.23	76.8 \pm 0.53
Concentrate-fed group	76.3 \pm 0.5	72.2 \pm 0.87	78.2 \pm 0.7
Pasture-fed group	42.9 \pm 0.46	59.8 \pm 0.93	66.1 \pm 0.86

*SD, Standard deviation of the mean.

lambs fed with fresh alfalfa (Ciftci et al., 2010). In the present study, total MUFA in omental and perirenal fat of maternal milk-fed lambs was significantly higher than those of pasture-fed lambs ($P < 0.05$). Perirenal and omental fat from animals fed maternal milk and concentrate diets had significantly higher oleic acid than pasture-fed ($P < 0.05$). In oleic acid of tail fat, no statistical differences were observed between feeding regime ($P > 0.05$).

Total PUFA was 3.75, 4.74 and 2.20% of total fatty acids in omental fat, 3.81, 4.75 and 3.07% of total fatty acids in perirenal fat and 3.73, 4.98 and 3.14% of total fatty acids in tail fat from maternal milk, concentrate and pasture-fed lambs, respectively. Ciftci et al. (2010) have also reported higher values of PUFA (10.48%) for tail fat of lambs fed with fresh alfalfa compared with results of the present study. Total PUFA have been stated as 2.11% in tail fat of Akkaraman sheep (Yilmaz and Karakaya, 2010). Osorio et al. (2009) have also reported higher values of PUFA in omental (5.45%) and perirenal fat (5.75%) deposit of suckling lambs reared on ewe's milk compared with our values. Salva et al. (2009) have found that, total PUFA was 5.75% in perirenal fat of alpaca. In the present study, total PUFA in omental, perirenal and tail fat was significantly higher in concentrate-fed lambs than maternal milk and pasture-fed lambs ($P < 0.05$). The high value of linoleic acid (51.81% of total fatty acids; Table 2) in concentrate feed increased this fatty acid and total PUFA in omental, perirenal and tail fat of concentrate-fed lambs. On the other hand, Nuernberg et al. (2008) have determined that, total PUFA in tail fat of lambs fed on pasture was higher than those of concentrate-fed lambs. In the present study, linoleic acid was the most represented PUFA and was significantly higher in concentrate-fed

lambs than maternal milk and pasture-fed lambs in omental, perirenal and tail fat ($P < 0.05$). These results agree with those reported by Nuernberg et al. (2008), who related that, linoleic acid was higher in tail fat of lambs fed concentrate compared with grass fed animals. Grazing lambs on pasture led to a significant increase of C 18:3 ω 3 α -linolenic acid in the perirenal and tail fat of Akkaraman lambs as they consumed grass which is rich in linolenic acid ($P < 0.05$).

The predominant CLA isomers of three CLA isomers in omental, perirenal and tail fat was C18:2 c9,t11. Total CLA was significantly higher in omental fat of maternal milk-fed lambs than those of concentrate and pasture-fed lambs ($P < 0.05$; Table 5). However, pasture feeding significantly enhanced total CLA in perirenal fat of Akkaraman lambs ($P < 0.05$; Tables 6). Aourousseau et al. (2007b) stated that, grazing have lowered ω 6 PUFA and increased ω 3 PUFA and C 18:2 c9,t11 compared with concentrate feeding. Feeding grass-based diets increases the 18:2 c9,t11 content of ruminant fat (French et al., 2000). In the present study, C18:2 c9,t11 and total CLA in perirenal and tail fat of pasture-fed lambs was higher about two or three times than those of concentrate-fed lambs. CLA amounts in adipose tissues have been determined as 3 to 4 times higher in Akkaraman lambs fed with alfalfa than lambs fed with wheat straw (Ciftci et al., 2010).

Total *trans* fatty acid (TFA) was found as 5.11, 11.76 and 3.18% of total fatty acids in omental fat, 4.61, 11.57 and 7.41% of total fatty acids in perirenal fat and 4.87, 10.12 and 5.68% of total fatty acids in tail fat from maternal milk, concentrate and pasture-fed lambs, respectively. Major TFA was C 18:1 t11 *trans*-vaccenic acid in omental, perirenal and tail fat of maternal milk, concentrate and pasture-fed lambs. *Trans* vaccenic acid

Table 5. Fatty acid composition (%) of omental fat of lambs fed diets containing maternal milk, concentrate and pasture^x.

Fatty acid	Maternal milk group (n=15)	Concentrate group (n=15)	Pasture group (n=15)
C 10:0*	0.45 ± 0.08 ^{a, y}	0.22 ± 0.05 ^b	0.29 ± 0.08 ^b
C 11:0	0.03 ± 0.01 ^{a, z}	0.01 ± 0.00 ^b	0.01 ± 0.00 ^b
C 12:0	1.01 ± 0.26 ^a	0.15 ± 0.04 ^c	0.35 ± 0.19 ^b
C 13:0	0.09 ± 0.03 ^a	0.02 ± 0.01 ^b	0.04 ± 0.02 ^b
C 14:0	8.89 ± 1.30 ^a	3.15 ± 0.50 ^c	4.76 ± 1.19 ^b
C 15:0	0.88 ± 0.12 ^a	0.56 ± 0.10 ^b	0.90 ± 0.13 ^a
C 16:0	26.13 ± 1.52 ^a	21.58 ± 1.02 ^c	23.55 ± 1.35 ^b
C 17:0	1.45 ± 0.16 ^b	2.04 ± 0.44 ^a	1.71 ± 0.23 ^b
C 18:0	15.31 ± 2.46 ^c	20.92 ± 3.73 ^b	31.30 ± 3.05 ^a
C 19:0	0.32 ± 0.08 ^a	0.12 ± 0.02 ^b	0.12 ± 0.04 ^b
C 20:0	0.07 ± 0.02 ^c	0.11 ± 0.02 ^b	0.57 ± 0.06 ^a
C 21:0	0.02 ± 0.01 ^b	0.05 ± 0.02 ^a	0.04 ± 0.01 ^a
C 22:0	0.01 ± 0.01 ^b	0.02 ± 0.01 ^b	0.11 ± 0.04 ^a
Σ SFA ^t	54.67 ± 4.43 ^b	48.95 ± 3.35 ^c	63.75 ± 3.53 ^a
C 14:1ω-5	0.29 ± 0.07 ^b	0.20 ± 0.04 ^c	0.64 ± 0.12 ^a
C 15:1ω-5	0.21 ± 0.02 ^b	0.08 ± 0.03 ^c	0.46 ± 0.07 ^a
C 16:1ω-7	2.85 ± 0.51 ^a	1.27 ± 0.17 ^c	1.92 ± 0.12 ^b
C 17:1ω-8	0.73 ± 0.25 ^a	0.60 ± 0.18 ^a	0.37 ± 0.07 ^b
C 18:1ω-9	30.29 ± 3.39 ^a	30.49 ± 3.12 ^a	26.46 ± 3.58 ^b
C 18:1ω-7	1.09 ± 0.26 ^a	1.50 ± 0.62 ^a	0.52 ± 0.09 ^b
C 20:1ω-9	0.03 ± 0.02 ^a	0.01 ± 0.01 ^b	0.01 ± 0.00 ^b
C 22:1ω-9	0.01 ± 0.00 ^a	0.01 ± 0.01 ^a	0.01 ± 0.00 ^a
Σ MUFA ^t	35.51 ± 4.28 ^a	34.16 ± 3.34 ^{ab}	30.40 ± 3.45 ^b
C 18:2ω-6	2.88 ± 0.43 ^b	4.13 ± 0.68 ^a	1.64 ± 0.07 ^c
C 18:3ω-6	0.09 ± 0.02 ^a	0.02 ± 0.01 ^b	0.01 ± 0.00 ^b
C 18:3ω-3	0.26 ± 0.04 ^a	0.20 ± 0.03 ^b	0.24 ± 0.05 ^a
C 20:2ω-6	0.06 ± 0.01 ^b	0.07 ± 0.02 ^a	0.02 ± 0.00 ^c
C 20:3ω-6	0.04 ± 0.01 ^a	0.03 ± 0.01 ^b	0.02 ± 0.01 ^c
C 20:3ω-3	0.02 ± 0.01 ^a	0.02 ± 0.01 ^a	0.03 ± 0.02 ^a
C 20:4ω-6	0.14 ± 0.04 ^a	0.08 ± 0.02 ^b	0.06 ± 0.02 ^b
C 20:5ω-3	0.02 ± 0.01 ^a	0.02 ± 0.01 ^a	0.02 ± 0.01 ^a
C 22:2ω-6	0.02 ± 0.01 ^a	0.02 ± 0.01 ^a	0.01 ± 0.01 ^a
C 22:3ω-3	0.07 ± 0.06 ^a	0.05 ± 0.04 ^a	0.04 ± 0.03 ^a
C 22:4ω-6	0.05 ± 0.02 ^a	0.04 ± 0.02 ^a	0.01 ± 0.01 ^b
C 22:5ω-6	0.02 ± 0.01 ^a	0.02 ± 0.01 ^a	0.01 ± 0.01 ^a
C 22:5ω-3	0.09 ± 0.02 ^a	0.03 ± 0.02 ^b	0.08 ± 0.01 ^a
C 22:6ω-3	0.02 ± 0.01 ^a	0.02 ± 0.01 ^a	0.01 ± 0.00 ^a
Σ PUFA ^t	3.75 ± 0.48 ^b	4.74 ± 0.71 ^a	2.20 ± 0.12 ^c
C 18:2 c9, t11	0.94 ± 0.24 ^a	0.34 ± 0.07 ^b	0.45 ± 0.07 ^b
C 18:2 t10, c12	0.01 ± 0.00 ^b	0.04 ± 0.01 ^a	0.01 ± 0.00 ^b
C 18:2 c11, t13	0.01 ± 0.00 ^b	0.02 ± 0.02 ^a	0.01 ± 0.00 ^b
Σ CLA ^t	0.96 ± 0.24 ^a	0.40 ± 0.08 ^b	0.47 ± 0.07 ^b
C 14:1t9	0.25 ± 0.05 ^b	0.08 ± 0.05 ^c	0.40 ± 0.06 ^a
C 16:1t9	0.50 ± 0.05 ^a	0.16 ± 0.04 ^b	0.49 ± 0.05 ^a
C 18:1 t9	0.02 ± 0.01 ^a	0.01 ± 0.00 ^b	0.02 ± 0.01 ^a

Table 5. Continue.

C 18:1 <i>t</i> 11	3.90 ± 1.00 ^b	11.36 ± 3.46 ^a	2.04 ± 0.22 ^b
C 18:2 <i>t</i> 9, <i>t</i> 12	0.27 ± 0.06 ^a	0.08 ± 0.04 ^b	0.12 ± 0.10 ^b
C 18:2 <i>t</i> 9, <i>c</i> 12	0.18 ± 0.08 ^a	0.07 ± 0.02 ^b	0.11 ± 0.04 ^b
Σ TFA [†]	5.11 ± 1.02 ^b	11.76 ± 3.50 ^a	3.18 ± 0.14 ^b
Σ ω-3	0.48 ± 0.11 ^a	0.33 ± 0.09 ^b	0.41 ± 0.09 ^{ab}
Σ ω-6	3.29 ± 0.45 ^b	4.40 ± 0.69 ^a	1.79 ± 0.06 ^c
ω-3/ω-6	0.14 ± 0.04 ^b	0.08 ± 0.02 ^c	0.23 ± 0.05 ^a
ω-6/ω-3	6.85 ± 1.63 ^b	13.33 ± 3.12 ^a	4.37 ± 1.11 ^c

[‡]Average of three lots analyzed; [†]values reported are mean ± SD; [‡]abc values for each sample with different letters in the same fraction are significantly different at $p < 0.05$. [†]SFA: Saturated fatty acid; MUFA: monounsaturated fatty acid; PUFA: polyunsaturated fatty acid; TFA: *trans* fatty acid; CLA: conjugated linoleic acid.

Table 6. Fatty acid composition (%) of perirenal fat of lambs fed diets containing maternal milk, concentrate and pasture^x.

Fatty acid	Maternal milk group (n=15)	Concentrate group (n=15)	Pasture group (n=15)
C 10:0*	0.36 ± 0.08 ^{a,y}	0.21 ± 0.03 ^b	0.42 ± 0.26 ^a
C 11:0	0.02 ± 0.01 ^{a,z}	0.01 ± 0.01 ^b	0.01 ± 0.01 ^b
C 12:0	0.73 ± 0.16 ^a	0.12 ± 0.03 ^b	0.61 ± 0.40 ^a
C 13:0	0.06 ± 0.02 ^a	0.03 ± 0.02 ^c	0.04 ± 0.02 ^b
C 14:0	6.94 ± 1.18 ^a	2.87 ± 0.55 ^b	6.76 ± 3.62 ^a
C 15:0	0.65 ± 0.15 ^b	0.53 ± 0.30 ^b	0.94 ± 0.23 ^a
C 16:0	21.59 ± 2.04 ^{ab}	19.95 ± 1.97 ^b	22.80 ± 3.29 ^a
C 17:0	1.59 ± 0.15 ^b	2.00 ± 0.43 ^a	1.59 ± 0.13 ^b
C 18:0	23.18 ± 4.98 ^a	23.24 ± 4.53 ^a	27.05 ± 4.60 ^a
C 19:0	0.25 ± 0.11 ^a	0.11 ± 0.05 ^b	0.24 ± 0.06 ^a
C 20:0	0.07 ± 0.05 ^c	0.15 ± 0.03 ^b	0.48 ± 0.12 ^a
C 21:0	0.02 ± 0.01 ^c	0.05 ± 0.01 ^a	0.04 ± 0.02 ^b
C 22:0	0.01 ± 0.00 ^b	0.02 ± 0.01 ^b	0.10 ± 0.05 ^a
Σ SFA [†]	55.48 ± 3.56 ^b	49.29 ± 2.91 ^c	61.06 ± 3.32 ^a
C 14:1ω-5	0.20 ± 0.14 ^b	0.19 ± 0.06 ^b	0.52 ± 0.11 ^a
C 15:1ω-5	0.18 ± 0.03 ^b	0.08 ± 0.04 ^c	0.29 ± 0.05 ^a
C 16:1ω-7	2.11 ± 0.79 ^a	1.19 ± 0.35 ^b	1.52 ± 0.18 ^b
C 17:1ω-8	0.60 ± 0.20 ^a	0.55 ± 0.27 ^a	0.33 ± 0.08 ^b
C 18:1ω-9	31.05 ± 2.50 ^a	30.44 ± 3.01 ^a	24.19 ± 1.21 ^b
C 18:1ω-7	1.09 ± 0.21 ^b	1.54 ± 0.46 ^a	0.60 ± 0.21 ^c
C 20:1ω-9	0.03 ± 0.01 ^a	0.01 ± 0.00 ^b	0.01 ± 0.00 ^b
C 22:1ω-9	0.01 ± 0.01 ^a	0.01 ± 0.00 ^a	0.01 ± 0.00 ^a
Σ MUFA [†]	35.27 ± 3.40 ^a	34.01 ± 3.23 ^a	27.47 ± 1.46 ^b
C 18:2ω-6	3.00 ± 0.43 ^b	4.21 ± 0.70 ^a	1.84 ± 0.31 ^c
C 18:3ω-6	0.14 ± 0.06 ^a	0.02 ± 0.01 ^b	0.02 ± 0.00 ^b
C 18:3ω-3	0.25 ± 0.05 ^b	0.19 ± 0.04 ^b	0.87 ± 0.25 ^a
C 20:2ω-6	0.06 ± 0.01 ^b	0.07 ± 0.02 ^a	0.06 ± 0.02 ^b
C 20:3ω-6	0.05 ± 0.02 ^a	0.03 ± 0.01 ^b	0.02 ± 0.00 ^b
C 20:3ω-3	0.02 ± 0.01 ^b	0.02 ± 0.01 ^b	0.04 ± 0.03 ^a
C 20:4ω-6	0.11 ± 0.05 ^a	0.06 ± 0.02 ^b	0.03 ± 0.01 ^b
C 20:5ω-3	0.02 ± 0.01 ^a	0.02 ± 0.01 ^a	0.02 ± 0.01 ^a

Table 6. Continue.

C 22:2 ω -6	0.02 \pm 0.01 ^a	0.02 \pm 0.01 ^a	0.02 \pm 0.01 ^a
C 22:3 ω -3	0.04 \pm 0.03 ^a	0.03 \pm 0.02 ^a	0.04 \pm 0.02 ^a
C 22:4 ω -6	0.03 \pm 0.01 ^a	0.03 \pm 0.02 ^a	0.01 \pm 0.01 ^b
C 22:5 ω -6	0.01 \pm 0.00 ^a	0.02 \pm 0.01 ^a	0.02 \pm 0.01 ^a
C 22:5 ω -3	0.06 \pm 0.02 ^b	0.02 \pm 0.01 ^c	0.09 \pm 0.02 ^a
C 22:6 ω -3	0.02 \pm 0.01 ^a	0.02 \pm 0.01 ^a	0.02 \pm 0.01 ^a
Σ PUFA ^t	3.81 \pm 0.54 ^b	4.75 \pm 0.70 ^a	3.07 \pm 0.42 ^c

^xAverage of three lots analyzed; ^yvalues reported are mean \pm SD; ^zabc values for each sample with different letters in the same fraction are significantly different at $p < 0.05$. ^tSFA: Saturated fatty acid; MUFA: monounsaturated fatty acid; PUFA: polyunsaturated fatty acid; TFA: trans fatty acid; CLA: conjugated linoleic acid.

Table 7. Fatty acid composition (%) of tail fat of lambs fed diets containing maternal milk, concentrate and pasture^x.

Fatty acid	Maternal milk group (n=15)	Concentrate group (n=15)	Pasture group (n=15)
C 10:0*	0.48 \pm 0.07 ^{a, y}	0.24 \pm 0.06 ^b	0.28 \pm 0.11 ^b
C 11:0	0.04 \pm 0.01 ^{ab, z}	0.05 \pm 0.04 ^a	0.02 \pm 0.01 ^b
C 12:0	0.76 \pm 0.27 ^a	0.31 \pm 0.16 ^b	0.36 \pm 0.17 ^b
C 13:0	0.09 \pm 0.03 ^a	0.09 \pm 0.04 ^a	0.05 \pm 0.03 ^b
C 14:0	7.38 \pm 1.65 ^a	2.99 \pm 0.70 ^c	5.18 \pm 1.28 ^b
C 15:0	0.99 \pm 0.17 ^b	1.52 \pm 0.41 ^a	0.99 \pm 0.17 ^b
C 16:0	24.85 \pm 2.06 ^a	20.74 \pm 2.55 ^c	22.98 \pm 0.82 ^b
C 17:0	1.65 \pm 0.33 ^b	2.99 \pm 0.61 ^a	1.76 \pm 0.22 ^b
C 18:0	10.66 \pm 3.30 ^b	9.38 \pm 1.99 ^b	15.13 \pm 4.70 ^a
C 19:0	0.44 \pm 0.09 ^a	0.34 \pm 0.09 ^b	0.45 \pm 0.09 ^a
C 20:0	0.07 \pm 0.03 ^b	0.09 \pm 0.02 ^b	0.25 \pm 0.14 ^a
C 21:0	0.02 \pm 0.01 ^b	0.05 \pm 0.03 ^a	0.03 \pm 0.01 ^b
C 22:0	0.01 \pm 0.00 ^b	0.02 \pm 0.01 ^a	0.02 \pm 0.01 ^a
Σ SFA ^t	47.42 \pm 5.49 ^a	38.82 \pm 3.63 ^b	47.49 \pm 3.25 ^a
C 14:1 ω -5	0.39 \pm 0.07 ^b	0.42 \pm 0.17 ^b	0.59 \pm 0.17 ^a
C 15:1 ω -5	0.22 \pm 0.03 ^b	0.24 \pm 0.12 ^{ab}	0.31 \pm 0.04 ^a
C 16:1 ω -7	3.73 \pm 0.67 ^a	2.80 \pm 0.70 ^b	3.10 \pm 1.00 ^b
C 17:1 ω -8	1.24 \pm 0.35 ^b	2.30 \pm 0.91 ^a	1.01 \pm 0.39 ^b
C 18:1 ω -9	35.81 \pm 4.81 ^a	37.84 \pm 4.71 ^a	36.66 \pm 3.45 ^a
C 18:1 ω -7	1.41 \pm 0.35 ^a	1.68 \pm 0.52 ^a	0.75 \pm 0.23 ^b
C 20:1 ω -9	0.06 \pm 0.02 ^a	0.05 \pm 0.03 ^a	0.01 \pm 0.00 ^b
C 22:1 ω -9	0.01 \pm 0.00 ^a	0.02 \pm 0.01 ^a	0.01 \pm 0.00 ^a
Σ MUFA ^t	42.86 \pm 5.93 ^a	45.34 \pm 6.19 ^a	42.43 \pm 4.88 ^a
C 18:2 ω -6	2.90 \pm 0.46 ^b	4.16 \pm 0.76 ^a	1.86 \pm 0.60 ^c
C 18:3 ω -6	0.09 \pm 0.03 ^a	0.03 \pm 0.01 ^b	0.04 \pm 0.01 ^b
C 18:3 ω -3	0.27 \pm 0.05 ^b	0.24 \pm 0.05 ^b	0.75 \pm 0.47 ^a
C 20:2 ω -6	0.06 \pm 0.01 ^b	0.09 \pm 0.02 ^a	0.05 \pm 0.02 ^b
C 20:3 ω -6	0.04 \pm 0.01 ^a	0.04 \pm 0.02 ^a	0.02 \pm 0.00 ^b
C 20:3 ω -3	0.02 \pm 0.01 ^b	0.03 \pm 0.02 ^a	0.04 \pm 0.02 ^a
C 20:4 ω -6	0.13 \pm 0.06 ^a	0.09 \pm 0.04 ^b	0.08 \pm 0.04 ^b
C 20:5 ω -3	0.02 \pm 0.01 ^b	0.03 \pm 0.02 ^a	0.02 \pm 0.01 ^b
C 22:2 ω -6	0.02 \pm 0.01 ^b	0.03 \pm 0.02 ^a	0.02 \pm 0.01 ^b
C 22:3 ω -3	0.05 \pm 0.04 ^b	0.09 \pm 0.06 ^{ab}	0.12 \pm 0.07 ^a
C 22:4 ω -6	0.05 \pm 0.03 ^{ab}	0.07 \pm 0.07 ^a	0.02 \pm 0.01 ^b

Table 7. Continued.

C 22:5 ω -6	0.02 \pm 0.01 ^b	0.03 \pm 0.02 ^a	0.02 \pm 0.01 ^b
C 22:5 ω -3	0.07 \pm 0.03 ^a	0.03 \pm 0.01 ^b	0.09 \pm 0.07 ^a
C 22:6 ω -3	0.02 \pm 0.01 ^a	0.03 \pm 0.01 ^a	0.02 \pm 0.01 ^a
Σ PUFA ^t	3.73 \pm 0.54 ^b	4.98 \pm 0.85 ^a	3.14 \pm 0.93 ^b
C 18:2 <i>c</i> 9, <i>t</i> 11	1.09 \pm 0.21 ^a	0.69 \pm 0.19 ^b	1.21 \pm 0.13 ^a
C 18:2 <i>t</i> 10, <i>c</i> 12	0.01 \pm 0.00 ^b	0.03 \pm 0.02 ^a	0.01 \pm 0.01 ^b
C 18:2 <i>c</i> 11, <i>t</i> 13	0.01 \pm 0.00 ^b	0.02 \pm 0.02 ^a	0.04 \pm 0.02 ^a
Σ CLA ^t	1.11 \pm 0.21 ^a	0.74 \pm 0.19 ^b	1.26 \pm 0.14 ^a
C 14:1 <i>t</i> 9	0.22 \pm 0.08 ^b	0.26 \pm 0.14 ^b	0.35 \pm 0.04 ^a
C 16:1 <i>t</i> 9	0.51 \pm 0.07 ^a	0.35 \pm 0.18 ^b	0.51 \pm 0.06 ^a
C 18:1 <i>t</i> 9	0.04 \pm 0.02 ^a	0.02 \pm 0.01 ^b	0.04 \pm 0.02 ^a
C 18:1 <i>t</i> 11	3.60 \pm 1.02 ^b	9.21 \pm 3.22 ^a	4.28 \pm 2.01 ^b
C 18:2 <i>t</i> 9, <i>t</i> 12	0.28 \pm 0.05 ^a	0.08 \pm 0.04 ^b	0.22 \pm 0.17 ^a
C 18:2 <i>t</i> 9, <i>c</i> 12	0.23 \pm 0.07 ^b	0.20 \pm 0.05 ^b	0.28 \pm 0.04 ^a
Σ TFA ^t	4.87 \pm 1.03 ^b	10.12 \pm 3.13 ^a	5.68 \pm 1.96 ^b
Σ ω -3	0.44 \pm 0.09 ^b	0.44 \pm 0.12 ^b	1.04 \pm 0.59 ^a
Σ ω -6	3.30 \pm 0.51 ^b	4.53 \pm 0.78 ^a	2.11 \pm 0.63 ^c
ω -3/ ω -6	0.13 \pm 0.03 ^b	0.10 \pm 0.02 ^b	0.49 \pm 0.26 ^a
ω -6/ ω -3	7.50 \pm 1.56 ^b	10.30 \pm 2.18 ^a	2.03 \pm 1.13 ^c

^xAverage of three lots analyzed; ^yValues reported are mean \pm SD; ^zabc values for each sample with different letters in the same fraction are significantly different at $p < 0.05$. TFA: *trans* fatty acid; CLA: conjugated linoleic acid; ^tSFA: Saturated fatty acid; MUFA: monounsaturated fatty acid; PUFA: polyunsaturated fatty acid.

which is the predominant *trans* monounsaturated fatty acid in ruminant milk and tissue fat (Parodi, 1976; Precht et al., 2001) is formed as an intermediate during the biohydrogenation of dietary linoleic acid to stearic acid (Kepler and Tove, 1967; Jiang et al., 1996).

In the present study, pasture feeding increased total ω 3 and ω 3/ ω 6 ratio in perirenal and tail fat. Significant differences were observed between pasture and concentrate groups in tail and perirenal fat for total ω 3 and ω 3/ ω 6 ratio ($P < 0.05$). From the nutritional aspect, fat from lambs raised on pasture seems to be more adequate, than that of lambs raised in confinement with concentrate because of their higher proportion on ω 3 PUFA and CLA and lower ω 6/ ω 3 ratio (Santos-Silva et al., 2002). In the present study, omental, perirenal and tail fat of concentrate-fed lambs had higher ω 6/ ω 3 ratio and pasture feeding decreased this ratio. The ratio of ω 6/ ω 3 fatty acid was significantly lower in grass fed lamb adipose tissue fat compared with lambs fed with concentrate (Nuernberg et al., 2008). The ω 6/ ω 3 ratio, in perirenal fat (1.87) and tail fat (2.03) from pasture fed lambs was below the recommended level of 4 for human consumption (Department of Health, 1994).

In conclusion, fatty acid composition of omental, perirenal and tail fat from Akkaraman lambs were affected

by feeding regime. Omental, perirenal and tail fat of lambs fed pasture contained more CLA, total ω 3, ω 3/ ω 6 ratio which is beneficial to human health compared with concentrate-fed lambs. Moreover, omental, perirenal and tail fat of lambs fed concentrate had higher ω 6/ ω 3 ratio and pasture feeding decreased this ratio. Fatty acid composition of omental, perirenal and tail fat from lambs can be improved by including pasture in the feeding regime.

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REFERENCES

- Aharoni Y, Nachtomi E, Holstein P, Brosh A, Holzer Z, Nitsan Z (1995). Dietary effects on fat deposition and fatty acid profiles in muscle and fat depots of Friesian bull calves. *J. Anim. Sci.* 73: 2712-2720.
- Akman N, Emiroğlu M, Tavmen A (2001). *Koyunculuk, Dünya'da- Avrupa Birliği'nde- Türkiye'de Hayvansal Üretim ve Ticareti.* (In Turkish). Çamlica Kültür ve Yardım Vakfı, İstanbul, p. 159.

- Anonymous (2000). Statistical Yearbook of Turkey. State Institute of Statistics Prime Ministry Republic of Turkey, Ankara.
- Arana A, Mendizabal JA, Alzon M, Eguinoa P, Beriain MJ, Purroy A (2006). Effect of feeding lambs oleic acid calcium soaps on growth, adipose tissue development and composition. *Small Ruminant Res.* 63: 75-83.
- Aurousseau B, Bauchart D, Galot AL, Prache S, Micol D, Priolo A (2007a). Indoor fattening of lambs raised on pasture: 2. Influence of stall finishing duration on triglyceride and phospholipid fatty acids in the *longissimus thoracis* muscle. *Meat Sci.* 76: 417-427.
- Aurousseau B, Bauchart D, Faure X, Galot AL, Prache S, Micol D, Priolo A (2007b). Indoor fattening of lambs raised on pasture: (1) Influence of stall finishing duration on lipid classes and fatty acids in the *longissimus thoracis* muscle. *Meat Sci.* 76: 241-252.
- Ciftci M, Cerci IH, Kilinc U, Yilmaz O, Gurdogan F, Seven PT, Bahsi M, Benzer F, Ozcelik M (2010). Effects of alfalfa (fresh, silage, hay) on the fatty acid and conjugated linoleic acid amounts in lamb muscles and fats. *Rev. Med. Vet-Toulouse*, 161: 432-437.
- Dannenberger D, Nuernberg K, Nuernberg G, Scollan N, Steinhart H, Ender K (2005). Effect of pasture vs. concentrate diet on CLA isomer distribution in different tissue lipids of beef cattle. *Lipids*, 40: 589-598.
- Demirel G, Ozpinar H, Nazli B, Keser O (2006). Fatty acids of lamb meat from two breeds fed different forage: concentrate ratio. *Meat Sci.* 72: 229-235.
- Department of Health (1994). Report on Health and Social Subjects No. 46. Nutritional Aspects of Cardiovascular Disease. London: HMSO.
- Enser M (1991). Animal carcass fats and fish oils. In: Rossel JB, Pritchard, JLR (Eds.). *Analysis of Oilseeds, Fats and Fatty Foods.* Elsevier Appl. Sci. London.
- French P, Stanton C, Lawless F, O'Riordan EG, Monahan FJ, Caffrey PJ, Moloney AP (2000). Fatty acid composition, including conjugated linoleic acid, of intramuscular fat from steers offered grazed grass, grass silage or concentrate-based diets. *J. Anim. Sci.* 78: 2849-2855.
- Folch J, Lees M, Sloane Stanley GH (1957). A simple method for the isolation and purification of total lipids from animal tissues. *J. Biol. Chem.* 226: 497-509.
- Griinari JM, Bauman DE (1999). Biosynthesis of conjugated linoleic acid and its incorporation into meat and milk in ruminants. In Yurawecz MP, Mossoba M, Kramer JK, Nelson G and Pariza MW (Eds). *Advances in Conjugated Linoleic Acid Research.* 1: 180-200. AOCS Press, Champaign, IL.
- Ha YL, Storkson J, Pariza MW (1990). Inhibition of benzo(α)pyrene-induced mouse forestomach neoplasia by conjugated dienoic derivatives of linoleic acid. *Cancer Res.* 50(4): 1097-1101.
- Ip C, Singh M, Thompson HJ, Scimeca JA (1994). Conjugated linoleic acid suppresses mammary carcinogenesis and proliferative activity of the mammary gland in the rat. *Cancer Res.* 54: 1212-1215.
- ISO-International Organization for Standardization (1978). *Animal and Vegetable Fats and Oils – Preparation of Methyl Esters of Fatty Acids.* ISO. Geneve, Method ISO 5509. pp. 1-6.
- Jiang J, Bjoerck L, Fonden R, Emanuelson M (1996). Occurrence of conjugated cis-9, trans 11-octadecadonic acid in bovine milk: effects of feed and dietary regimen. *J. Dairy Sci.* 79: 438-445.
- Kemp JD, Mahyuddin M, Ely DG, Fox JD, Moody WG (1981). Effect of feeding systems, slaughter weight and sex on organoleptic properties, and fatty acid composition of lamb. *J. Anim. Sci.* 51: 321-330.
- Kepler CR, Tove SB (1967). Biohydrogenation of unsaturated fatty acids. *J. Biol. Chem.* 242: 5686-5692.
- Kritchevsky D (2003). Conjugated linoleic acids in experimental atherosclerosis. In: Sebedio JL, Christie WW and Adolf RO (eds). *Advances in Conjugated Linoleic Acid Res.* 2: 292-301, AOCS Press, Champaign.
- Ledoux M, Chardigny JM, Darbois M, Soustre Y, Sebedio JL, Laloux L (2005). Fatty acid composition of French butters, with special emphasis on conjugated linoleic acid (CLA) isomers. *J. Food Compos. Anal.* 18: 409-425.
- Lee KN, Kritchevsky D, Pariza MW (1994). Conjugated linoleic acid and atherosclerosis in rabbits. *Atherosclerosis*, 108(1): 19-25.
- Mahgoub O, Khan AJ, Al-Maqbaly RS, Al-Sabahi JN, Annamalai K, Al-Sakry NM (2002). Fatty acid composition of muscle and fat tissues of Omani Jebel Akhdar goats of different sexes and weights. *Meat Sci.* 61: 381-387.
- Mitchell GE, Reed AW, Rogers SA (1991). Influence of feeding regimen on the sensory qualities and fatty acid contents of beef steaks. *J. Food Sci.* 56: 1102-1103.
- Moharrey A (2007). Effect of docking and energy of diet on carcass fat characteristics in fat-tailed Badghisian sheep. *Small Rumin. Res.* 69: 208-216.
- Nuernberg K, Fischer A, Nuernberg G, Ender K, Dannenberger D (2008). Meat quality and fatty acid composition of lipids in muscle and fatty tissue of Skudde lambs fed grass versus concentrate. *Small Rumin. Res.* 74: 279-283.
- Nürnberg K, Wegner J, Ender K (1998). Factors influencing fat composition in muscle and adipose tissue of farm animals. *Livest. Prod. Sci.* 56: 145-156.
- Oriani G, Maiorano G, Filetti F, Di Cesare C, Manchisi A, Salvatori G (2005). Effect of age on fatty acid composition of Italian Merino suckling lambs. *Meat Sci.* 71: 557-562.
- Osorio MT, Zumalacarregui JM, Alaiz-Rodriguez R, Guzman-Martinez R, Engelsen SB, Mateo J (2009). Differentiation of perirenal and omental fat quality of suckling lambs according to the rearing system from Fourier transforms mid-infrared spectra using partial least squares and artificial neural networks analysis. *Meat Sci.* 83: 140-147.
- Pariza MW, Park Y, Cook M, Albright K, Liu W (1996). Conjugated linoleic acid (CLA) reduces body fat. *FASEB J.* 10: p. 3227 (Abstract).
- Pariza MW, Park Y, Cook ME (2001). The biologically active isomers of conjugated linoleic acid. *Prog. Lipid Res.* 40: 283-298.
- Park Y, Pariza MW (2007). Mechanisms of body fat modulation by conjugated linoleic acid (CLA). *Food Res. Int.* 40(3): 311-323.
- Parodi PW (1976). Distribution of isomeric octadecenoic fatty acids in milk fat. *J. Dairy Sci.* 59: 1870-1873.
- Parodi PW (1997). Cow's milk fat components as potential anticarcinogenesis agents. *J. Nutr.* 127: 1055-1060.
- Precht D, Molkentin J, Destaillets F, Wolff RL (2001). Comparative studies on individual isomeric 18:1 acids in cow, goat, and ewe milk fats by low-temperature high-resolution capillary gas-liquid chromatography. *Lipids*, 36: 827-832.
- Rule DC, Smith SB, Romans JR (1995). Fatty acid composition of muscle and adipose tissue of meat animals. In Smith SB, Smith DR (Eds.). *Biol. Fat Meat Anim.*, Champaign, IL, pp. 144-165.
- Salva BK, Zumalacarregui JM, Figueira AC, Osorio MT, Mateo J (2009). Nutrient composition and technological quality of meat from alpacas reared in Peru. *Meat Sci.* 82: 450-455.
- Santos-Silva J, Bessa RJB, Santos-Silva F (2002). Effect of genotype, feeding system and slaughter weight on the quality of light lambs II. Fatty acid composition of meat. *Livest. Prod. Sci.* 77: 187-194.
- Serra A, Mele M, La Comba F, Conte G, Buccioni A, Secchiari P (2009). Conjugated Linoleic Acid (CLA) content of meat from three muscles of Massese suckling lambs slaughtered at different weights. *Meat Sci.* 81: 396-404.
- Ünsal M, Aktaş N (2003). Fractionation and characterization of edible sheep tail fat. *Meat Sci.* 63: 235-239.
- Ünsal M, Yanlic KO (2005). Fractionation and characterization of tail fats from Morkaraman lambs fed with diets containing *Rosa canina* L. seed at different levels. *Int. J. Food Prop.* 8: 301-312.
- Vanschoonbeek K, de Maat MPM, Heemskerk JWM (2003). Fish oil consumption and reduction of arterial disease. *J. Nutr.* 133: 657-660.
- Whigham LD, Cook ME, Atkinson RL (2000). Conjugated linoleic acid: implications for human health. *Pharmacol. Res.* 42: 503-510.
- Wood JD, Enser M (1997). Factors influencing fatty acids in meat and the role of antioxidants in improving meat quality. *Br. J. Nutr.* 78: 49-60.
- Wood JD, Richardson RI, Nute GR, Fisher AV, Campo MM, Kasapidou E, Sheard PR, Enser M (2004). Effects of fatty acids on meat quality: A review. *Meat Sci.* 66: 21-32.

Yılmaz MT, Karakaya M (2010). Thermal analysis of lipids isolated from various tissues of sheep fats. *J. Therm. Anal. Calorim.* 101: 403-409.