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Short Communication

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10 **Polymorphism of calpastatin gene in Arabic sheep**
11 **using PCR- RFLP**

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19 **Calpastatin has been known as candidate gene in muscle growth efficiency and meat quality. This**
20 **gene has been located to chromosome 5 of sheep. In order to evaluate the calpastatin gene**
21 **polymorphism, random blood sample were collected from 111 Arabic ram sheep from different**
22 **regions. The DNA extraction was based on Boom et al. (1989) method. Exon and intron I from L**
23 **domain of the ovine calpastatin gene was amplified to produce a 622 bp fragment. The PCR products**
24 **were electrophoresed on 1.2% agarose gel and stained by etidium bromide. Then, they were digested**
25 **with restriction enzyme *MspI* and then electrophoresed on 2.5% agarose gel with ethidium bromide**
26 **and revealed two alleles, allele A and allele B. Data were analysed using PopGene32 package. In this**
27 **population, AA, AB, BB genotype have been identified with the 70.27, 28.82, 0.9% frequencies. A and**
28 **B allele's frequencies were 0.85, 0.15, respectively. The population was found to follow Hardy-**
29 **Weinberg equilibrium.**

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31 **Key words:** Calpastatin gene, polymorphism, PCR-RFLP.

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34 **INTRODUCTION**

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36 Producing meat tenderness that consumers desire is one
37 of the major problems facing the industry, because meat
38 tenderization during the postmortem period is highly
39 variable between carcasses. Therefore, studies of bio-
40 chemical mechanisms for muscle breakdown are essen-
41 tial at the molecular level. The rate and extent of skeletal
42 muscle growth ultimately depends mainly on three
43 factors: rate of muscle protein synthesis, rate of muscle
44 protein degradation, and the number and size of skeletal
45 muscle cells. The calpain activity is required for myoblast
46 fusion (Kuryl et al., 2003; Barnoy et al, 1997) and cell
47 proliferation in addition to cell growth (Mellegren, 1997).
48 The calpain system may also affect the number of skele-
49 tal muscle cell (fibres) in domestic animals by altering
50 rate of myoblast proliferation and modulation myoblast
51 fusion. The calpain system is also important in normal
52 skeletal muscle growth. Increased rate of skeletal muscle
53 growth can result from a decreased rate of muscle pro-
54 tein degradation, and this is associated with a decrease
55 in activity of the calpain system, due principally to a large
56 increase in calpastatin activity (Goll et al., 1998).

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59 Calpastatin, which is an endogenous inhibitor (Ca²⁺ de-
60 pendent cysteine proteinase), plays a central role in re-
61 gulation of calpain activity in cells (Murachi et al., 1981;
62 Murachi, 1983; Forsberg et al., 1989) and is considered
63 to be one of the major modulators of the calpains. There-
64 fore, calpastatin may affect proteolysis of myofibrils due
65 to regulation of calpain, which can initiate postmortem
66 degradation of myofibril proteins (Goll et al., 1992; Huff-
67 long et al., 1996). At the protein structural level, cal-
68 pastatin is a five-domain inhibitory protein (Figure 1)
69 (Kilber and Koohmaraie, 1994).

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71 Calpastatin is present in all tissues expressing cal-
72 pains and in skeletal muscle. Calpastatin is expressed at
73 a higher level of activity than the calpains themselves. Of
74 the five domains, the N-terminal leader (L) domain does
75 not appear to have any calpains inhibitory activity, but
76 may be involved in targeting or intracellular localization
77 (Tavano et al., 1999), while the other domains (I-IV) are
78 highly homologous and are each independently capable
79 of inhibiting calpains (Cong et al., 1998). This indicates
80 that the inhibitory domains of calpastatin contain three
81 highly conserved regions, A, B and C, of which A, and C,
82 bind calpain in a strictly Ca²⁺-dependent manner but have
83 no inhibitory activity, whereas region B inhibits calpain on
84 its own. It is also found that the removal of the XL domain

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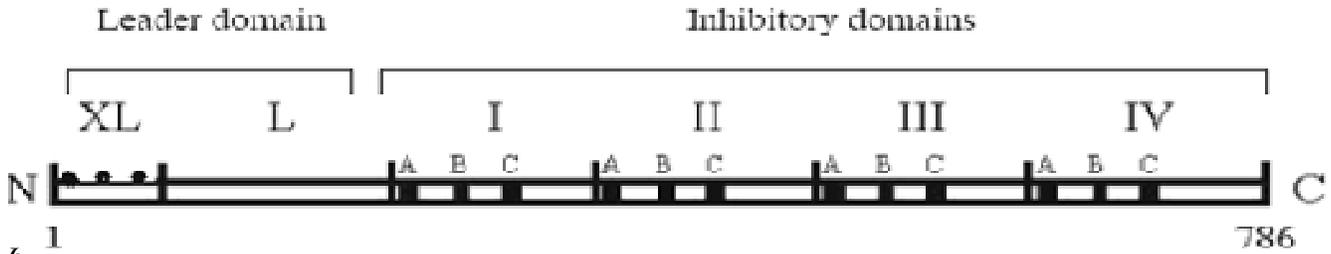


Figure 1. Structure of calpastatin's polypeptide domains.

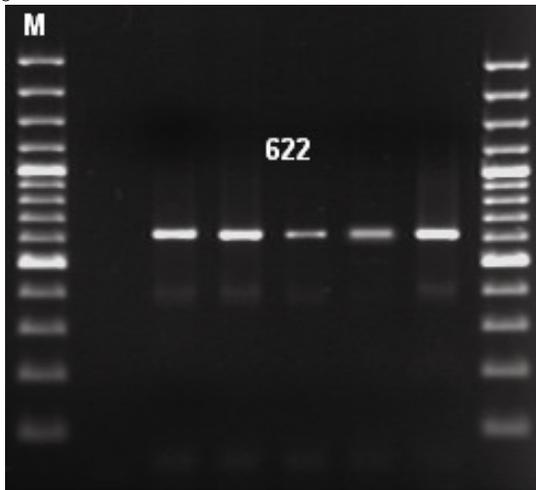


Figure 2. PCR product analyzed by electrophoresis (622 bp).

played a regulatory role by altering phosphorylation patterns on the protein (Takano et al., 1999). The purpose of the present investigation was to analyse the polymorphism of the CAST gene in Arabic sheep and evaluate its association with daily weight gain traits.

MATERIALS AND METHODS

In this study random blood samples were collected from 111 (Arabic) ram, sheep from different regions in southwestern of Iran. Approximately, 5 ml blood sample was gathered from vena in EDTA tube and was transferred to -20°C freezer. Genomic DNA was isolated by using DNA Extraction Kit and was based on Boom et al. (1996) method. Exon and intron region from a portion of the first repetitive domain of the ovine calpastatin gene were amplified to a product of 622 bp using primers based on the sequence of the bovine (Killefer and Koochmarraie, 1994; Gen bank accession no L13450) and ovine calpastatin genes. Spectrophotometer was used for investigating quality and quantity of DNA. The full sequence of primer:

CAST 1C 5'- TGGGGCCCAATGACGCCATCGATG - 3'
 CAST 1D 5'- GGTGGAGCAGCACTTCTGATCACC - 3'

Method of detection

The polymerase chain reaction (PCR) was performed using a buffer PCR1X, 200 µM dNTPs, 1.5 µM MgCL₂, 10 pmol each primer, 1.25

U 47 DNA polymerase, 50 ng ovine genomic DNA and H₂O up to a total volume of 25 µl. 33 cycle of preliminary denaturation at 95°C (5 min), denaturation at 94°C (1 min), annealing at 60°C (1 min), extension at 72°C (2 min) and final extension at 72°C (8 min). The PCR products were separated by 1.2% (w/v) agarose gel electrophoresis. The amplified fragment of calpastatin was digested with MspI 15 µl of PCR production with 2 µl buffer, 5U (0.5) of MspI and 11.5 µl H₂O up to a total volume of 29 µl, following the manufacturer's instruction for 12-16 h at 37°C. The digestion products were electrophoresed on 2.4% agarose gel in 1X TBE and visualized by ethidium bromide staining for 1 h at 85 V.

Statistical analysis

Estimates genotype and alleles frequencies and Hardy-Weinberg equilibrium was analysis with Pop Gene 32 package (Yeh et al., 1999).

RESULT AND DISCUSSION

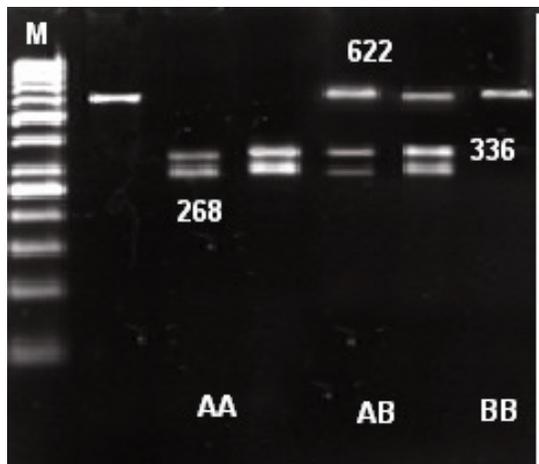
The amplified calpastatin resulted in a DNA fragment with 622 bp including the sequences of Exon and intron regions from a portion with PCR technique (Figure 2). Within the analyzed, two alleles (A and B) were observed resulting in three genotypes. The MspI digests the allele A amplicon, but not allele B. The animals with both alleles were assigned as AB genotype, whereas those possessing only A or B alleles as AA or BB genotypes, respectively. Genotype AA showed the two- band pattern (bands of approximately 339 and 286 bp). Genotype BB – one- band pattern (approximately 622), while AB animals displayed a pattern with all three- band (622, 336, 286) (Figure 3). This result shows that the polymorphism was detected in CAST I segment, as previously observed by Palmer et al. (1998) and chung et al. (2001).

A and B allele frequencies were 0.85 and 0.15, respectively. The genotype frequencies within 111 animals examined were 70.27 for AA, 0.9 for BB and 28.82 for AB (in 78,1,32 ram, sheep, respectively). χ^2 in this population showed Hardy – Weinberg equilibrium. The observed and expected heterozygosity were 0.26 and 0.28, respectively.

Effective allele and true allele are estimated 1.38 and 2, respectively. This different between effective all and true allele number and low diversity is due to more frequency of allele A compare to allele B, that reduced frequency in any locus. This number is more, if there are

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7 **Figure 3.** Genotype AA, AB and BB digestion with
8 *M*spl on 2.5% agarose gel. The band sizes indicated
9 in the figure are in base pairs (bp).

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13 more loci with same combination of alleles.

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