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REPRODUCTION AND TRYPANOSOMA CONGOLENSE IN NIGERIAN WEST AFRICAN DWARF EWES: II. GENITAL AND ENDOCRINE LESIONS

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Abstract

Aim: The study was designed to determine the effect of *Trypanosoma congolense* on the genital and endocrine organs of West African Dwarf (WAD) ewes.

Methods: Ten WAD ewes obtained for the study were divided into two groups comprising 5 ewes each. Group A was infected with *Trypanosoma congolense*, while group B was uninfected and allowed to run for eight weeks. At the end of the study period, three infected and two control ewes were sacrificed. The weights of the ovaries were determined and sections taken for histopathological examination. Sample sections from other parts of the reproductive tract, brain, pituitary gland, and hypothalamus were also taken.

Results: The mean ovarian weight, number of follicles and number of corpera lutea of the infected ewes decreased compared to the control ewes but were not statistically significant (p > 0.05). No parasites were seen following impression smears conducted on the ovaries. However, there were slight areas of necrosis and mild degeneration of the ovarian stroma. These were absent in the control ewes. No detectable gross lesions were seen in the adenohypophysis of both the infected and control ewe, although slight areas of focal necrosis were seen at histology. There were no detectable gross and histological lesions in the uterus, cervix, vagina, brain and hypothalamus of both infected and control ewes.

Conclusion: The findings from this study are of great importance for the economic exploitation of WAD sheep in tsetse infected area.

Keywords: Adenohypophysis, Ovaries, Trypanosoma congolense, WAD Ewe.

INTRODUCTION

Trypanosoma congolense is a haemoparasite affecting cattle, pigs, goats, sheep, horses, and dogs (OIE, 2013). It is a pathogenic parasite with two known strains, one from West Africa and the other from East Africa. However, the West African strain is more pathogenic (Osaer et al., 1994). T. congolense is transmitted biologically (Mbaya et al, although mechanical and congenital 2012), transmissions have been reported (Griffin, 1983; Desquesnes & Dia, 2003). Pregnant animals infected with T. congolense may abort or give birth to weak neonates (Faye et al., 2004). There is anorexia, anaemia, lacrimation, weight loss, weakness and death of the dam in some instances (Llewelyn et al,

1987). In non-pregnant females, there is anestrus (Llewelyn et al., 1988) and genital lesions (Ogwu and Njoku, 1991). Infected males show pathological changes characterized by testicular degeneration, penile protrusion, haemorrhage, prepucial inflammation, decrease testosterone levels, increase cortisol concentration and depressed pituitary and adrenocortical functions (Adeyemo et al., 1990; Sekoni et al. 1990; Raheem et al., 2009; Victor et al. 2012; Okubanjo et al. 2014; Okubanjo et al. 2015). pathogenesis of trypanosomosis-induced reproductive losses has been the subject of numerous researches (Ogwu et al., 1986; Edeghere et al., 1992; Faye et al., 2004; Leigh and Fayemi, 2013; Silver et al., 2013; Allam et al., 2014; Adeyeye et al., 2016a), with a few reviews available on the disease (Ikede et al., 1988; Sekoni, 1994; Raheem, 2014). The mechanism responsible for these losses is not widely known. However, Bawa (2000) suggested fetal hypoxia and stress, pyrexia, anemia and direct invasion of body tissues as possible mechanisms. The attack on the body tissues is characterized by pathological changes. T. congolense mainly resides in plasma (Seifert, 1996) and is believed to cause injury by anaemia without major histological changes on the tissue (Ikede and Loses, 1972). However, Ogwu and Njoku (1991) reported histopathological changes in T. congolense infected heifers. To the best of our knowledge, no study has been designed to ascertain this position in West African Dwarf ewe which is a trypanotolerant breed (Geerts et al., 2009). This study was carried out to determine the gross and histopathological changes associated with T. congolense infection in the genital and endocrine organs of WAD ewes.

MATERIALS AND METHODS

Experimental animals

Ten matured non-pregnant but cycling West African Dwarf (WAD) ewes obtained from the Small Ruminant Research Program, National Animal Production Research Institute, Shika-Zaria, Nigeria. They were selected from the sheep stock of the institute, and were initially used to study the effect of *Trypanosoma congolense* on the oestrous cycle of WAD, their management has therefore been described in Abubakar *et al.* (2015).

Study design

They were divided into two groups comprising 5 ewes each. Group A was infected with *Trypanosoma congolense* while group B was uninfected, and the study ran for eight weeks. The parasite used was obtained from the Department of Veterinary Parasitology and Entomology, Ahmadu Bello University Zaria, Nigeria.

Pathological examination

At the end of the study period, three infected and two control ewes were humanly euthanized and necropsied. The necropsy was carried out to examine for gross lesions on the reproductive tract and the endocrine glands. The weights of the ovaries were determined and sections taken for histopathological examination. Sample sections from other parts of the reproductive tract, brain, pituitary gland, and hypothalamus were also taken. All these were fixed in Bouin's solution and used for histopathological evaluation.

RESULTS

The infected ewes had a pre-patent period of $10.2 \pm$ 1.2 days. Other clinical signs were undulating parasitemia, intermittent pyrexia, anemia and emaciation. The mean ovarian weight, number of follicles and number of corpera lutea of the infected ewes is presented in Table 1. These parameters decreased compared to the control ewes but were not statistically significant (p > 0.05). Petechial haemorrhage was observed on the left ovary of one of the infected ewe. No parasites were seen following impression smears conducted on the ovaries. However, they had slight areas of necrosis and mild degeneration of the ovarian stroma. These were absent in the infected ewes. No detectable gross lesions were seen in the adenohypophysis of both infected and control ewe (Figure 1), although some slight areas of focal necrosis were seen histologically in the infected ewes (Figure 2). There were no detectable gross and histological lesions in the uterus, cervix, vagina, brain and hypothalamus of both infected and control ewes.

DISCUSSION

The ovarian weights of the infected ewes decreased but were not substantially different from the uninfected ewes. Similarly, the number of follicles and corpera lutea of the infected ewes did not differ from ewes in the control. Our observation contradicts the report of Isoun and Anosa (1974) as well as Adenowo *et al.* (2005) in *T. vivax* infected ewes, where substantial decreases were observed. Likewise, ovarian atrophy, reduction in number of follicles and corpera lutea has also been reported in *T. congolense* infected goats (Mutayoba *et al.*, 1988) and heifers (Ogwu and Njoku, 1991), as well as in

Table 1: Mean±SEM ovarian weight, number of follicles and number of corpora lutea in *Trypanosoma congolense* infected WAD ewes

	Mean ovarian weight	Mean number of follicles	Mean number of corpora lutea
Infected ewes (n=3)	1.33 ± 0.06	2.00 ± 0.29	1.50 ± 0.09
Control ewes (n=2)	1.45 ± 0.05	3.25 ± 0.43	1.75 ± 0.31

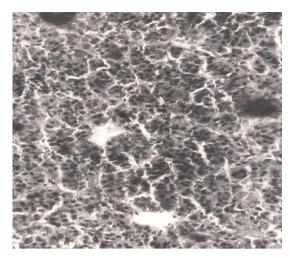


Fig 1: Adenohypophysis of a the uninfected WAD ewe showing normal parenchymal cells (H&E x 400).

T. vivax infected goats (Rodrigues et al., 2013) and cattle (Ige and Amodu, 1975). The contradictions may be attributed to the trypanotolerant nature of WAD ewes. There was petechial haemorrhage on the left ovary of one of the infected ewe. Since this was not generalized nor found in other infected ewes, it maynot be attributed to T. congolense infection in this study. Slight areas of necrosis and mild degeneration of the ovarian stroma were observed histologically on the ovaries of the infected ewes. In other trypanosomosis susceptible animals, lesions ranging from fibrosis degeneration of ovarian stroma and follicular cyst with atretic follicles have been reported (Mutayoba et al., 1988; Ogwu and Njoku, 1991; Adenowo et al., 2005), probably due to the breed variation suggested earlier. In the adenohypophysis, no gross lesions were seen among the infected ewes, although slight areas of focal necrosis were observed at histology. In T. vivax infected Yankasa ewes, Adenowo et al. (2005) reported mononuclear cell infiltration in the capsules and the parenchyma along with necrosis of the parenchyma cells of the adenohypophysis. These changes were observed in T. evansi infected ewes (Adeyeye, 2016b) as well as in goats infected with T. congolense (Mutayoba et al., 1988) and T. brucei (Leigh et al., 2015). Lesions on the adenohypophysis will lead to impairment in the release of gonadotropin-releasing hormone (GnRH) which is responsible for stimulating the production of follicular stimulating hormone (FSH) and luteinizing hormone (LH). This leads to impaired estrus cycle which was observed in our earlier report (Abubakar et al., 2015). The uterus, cervix, vagina, brain and hypothalamus had no detectable gross or histological changes. This is similar to the reports of (Adeyeye et al, 2016b) in T. evansi infected ewes, except for the hypothalamus which they reported had neuronal degeneration and

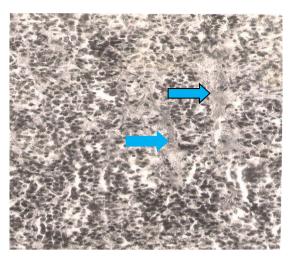


Fig 2: Adenophysis of a *T. congolense* infected WAD ewe showing focal necrosis (blue arrows) of the parenchymal cells (H&E x 400).

infiltrated by microglial cells. Our observations disagree with Adenowo et al. (2005), who reported lesions in the hypothalamus of T. vivax infected ewes. Leigh et al. (2015) also reported pathological lesions in the uterus and hypothalamus of WAD does infected with *T. brucei*. The variation in sheep breeds maybe responsible for this difference with Adenowo et al. (2005). Although WAD goats are also trypanotolerant, Sheep have been observed to be more trypanotolerant than goats (Boid et al., 1981). This probably explains our variation with Leigh et al. (2015). In conclusion, infection of WAD ewes with T. congolense showed no marked genital and endocrine lesions, in contrast with observations made in previous studies using other sheep breeds and trypanosoma species. It is therefore suggested that WAD ewes are capable of controlling the pathological effect of *T. congolense*. The phenomenon of trypanotolerance in WAD sheep might have contributed to the relatively mild pathological lesion on the genital and endocrine organs. This finding is of great importance for the economic exploitation of this breed of sheep in tsetse infected area.

REFERENCES

Abubakar YU, Oyedipe EO, Eduvie LO, Ogwu DO, Adeyeye AA (2015). Reproduction and *Trypanosoma congolense* in Nigerian West African Dwarf Ewes: I. Effects on the Oestrous Cycle. J. Protoz. Res, 25: 1-7.

Adenowo TK, Njoku CO, Oyedipe EO, Sannusi A (2005). Lesions of the hypothalamus, adenohypophysis and the ovaries in *Trypanosoma vivax* infected Yankasa ewes. Nig. Vet. J, 26: 56-62.

Adeyemo O, Oyejide A, Agbedana O (1990). Plasma testosterone in *Trypanosoma congolense* and

Trypanosoma brucei infected West African dwarf rams. Anim. Reprod. Sci, 22:21-26.

Adeyeye AA, Ate IU, Lawal AI & Adamu S (2016a). Effects of experimental *Trypanosoma evansi* infection on pregnancy in Yankasa ewes. Theriogenology, 85(5): 862–869.

Adeyeye AA, Ate IU, Lawal AI & Adamu S (2016b). Postpartum pathology in Yankasa ewes experimentally infected with *Trypanosoma evansi* during pregnant. Comp. Clin. Pathol, 10.1007/s00580-016-2236-y.

Allam L, Ogwu D, Agbede RIS & Sackey AKB (2014). Abortion and its probable cause in gilts experimentally infected with *Trypanosoma brucei*. J Protozool Res, 24: 26-32.

Bawa EK (2000). Studies on the effects of *Trypanosoma vivax* on ovine pregnancy. Unpublished Ph.D Dissertation. Ahmadu Bello University, Zaria, Nigeria.

Boid R, Amin EA, Mahmoud MM, Luckins AG (1981). *Trypanosoma evansi* infections and antibodies in goats, sheep and camels in the Sudan. Trop. Anim. Hlth. Prod, 13: 141-146.

Desquesnes M, Dia ML (2003). Mechanical transmission of *Trypanosoma congolense* in cattle by the African tabanid *Atylotus agrestis*. Exp. Parasitol, 105: 226–231.

Edeghere H, Elhassan E, Abenga J, Osue HO, Lawani FAG, Falope O (1992). Effects of infection with *Trypanosoma brucei brucei* on different trimesters of pregnancy in ewes. Vet. Parasit. 43:203-209.

Faye D, Sulon J, Kane Y, Beckers JF, Leak S, Kaboret Y, Sousa NM, Losson B, Geerts S (2004). Effects of an experimental *Trypanosoma congolense* infection on the reproductive performance of West African Dwarf goats. Theriogenology, 62: 1438-1451.

Geerts S, Osaer S, Goossens B, Faye D (2009). Trypanotolerance in small ruminants of sub-Saharan Africa. Trends Parasitol, 25:132-138.

Griffin L. (1983). Congenital transmission of *Trypanosoma congolense* in mice. J. Comp. Pathol, 93:489-92.

Ige K, Amodu AA (1975). Studies on the pathogenesis of trypanosomiasis for N'dama cattle. Proceedings of the 14th meeting of the OAU/ISCRT, Dakar, Senegal.

Ikede BO, Losos GJ (1972). Pathology of the disease in sheep produced experimentally *T. brucei* infection in sheep. Vet. Path, 9: 278-289.

with *Trypanosoma congolense*. Asian Pac. J. Trop. Dis, 4: 185-189.

Ikede BO, Elhassan E, Akpavie SO (1988). Reproductive disorders in African Trypanosomiasis. Acta Trop, 45: 5-10.

Isoun TT, Anosa VA (1974). Lesions in the reproductive organs of sheep and goats infected with *T. vivax. Z.*Tropenmed. Parasit, 25: 469-476.

Leigh OO, Emikpe BO, Ogunsola JO (2015). Histopathological changes in some reproductive and endocrine organs of *Trypanosoma brucei* infected West Africa Dwarf does. BJVM, 18: 31-39.

Leigh OO, Fayemi OE (2013). The effect of experimental *Trypanosoma brucei* infection on hormonal changes during the oestrous cycle, pregnancy and pregnancy outcome in West Africa dwarf does. Wayamba J. Anim. Sci., ID 1365263763.

Llewelyn CA, Luckins AG, Munro CD, Perrie J (1987). The effect of *Trypanosoma congolense* infection on the oestrous cycle of goat. Brit. Vet. J, 143: 432-431.

Llewelyn CA, Munro CD, Luckins AG, Jordt T, Murray M, Lorenzini E. (1988). The effects of *Trypanosoma congolense* infection on the oestrous cycle of Boran cow. Brit. Vet. J, 144: 379-387.

Mbaya AW, Kumshe H, Nwosu CO (2012). The mechanisms of anaemia in trypanosomosis: A Review. *In: Anemia*, (D. Silverberg editor.). Published by InTech.

Mutayoba BM, Gombe S, Kaaya GP, Waindi EN (1988). Effect of chronic experimental *Trypanosoma congolense* infection on the ovaries, pituitary, thyroid and adrenal glands in female goats. Res. Vet. Sci, 44: 140-146.

Ogwu, D. and Njoku, C. O. 1991.Genital lesions in experimental *Trypanosoma congolense* infection in heifers. Anim. Reprod. Sci, 26: 1-11.

Ogwu D, Njoku CO, Osori DIK (1986). Effects of experimental *Tryanosoma vivax*infection on first, second and third trimester pregnancy in heifers. Theriogenology, 25(3): 383-398.

OIE (2013). *Trypanosomosis* (*Tsetse-transmitted*). *Retrieved May 18*, 2015 from http://www.oie.int/fileadmin/Home/eng/Animal_Healt h_in_the_World/docs/pdf/Disease_cards/TRYPANO_TSETSE.pdf.

Okubanjo OO, Sekoni VO, Ajanusi OJ, Nok AJ, Adeyeye AA (2014). Testicular and epididymal pathology in Yankasa rams experimentally in infected

Okubanjo OO, Sekoni VO, Ajanusi OJ, Adeyeye AA (2015). Effects of experimental *Trypanosoma*

congolense infection on sperm morphology in Yankasa rams. Mac, Vet. Rev. 38 (2): 203-208.

Osaer S, Goossens B, Clifford DJ, Kora S, Kassama M (1994). A comparison of the susceptibility of Djallonké sheep and West African Dwarf goats to experimental infection with two different strains of *Trypanosoma congolense*. Vet. Parasitol, 51:191-204.

Raheem KA (2014). A review of trypanosomosisinduced reproductive dysfunctions in male animals. Agrosearch, 14: 30-38

Raheem AK, Fayemi EO, Leigh OO, Ameen SA (2009) Selected fertility parameters of West African Dwarf bucks experimentally infected with *Trypanosoma congolense*. Folia Vet, 53: 68-71.

Rodrigues CMF, Olinda RG, Silvaa TMF, Valea RG, Da Silvaa AE, Limaa GL, Garciab HA, Teixeirab MMG, Batista JS (2013). Follicular degeneration in the ovaries of goats experimentally infected with *Trypanosoma vivax* from the Brazilian semi-arid region. Vet. Parasit, 191: 146–153.

Seifert HSH (1996) Tropical Animal Health. Kluwer Academic publication.

Sekoni VO (1994). Reproductive disorders caused by animal trypanosomiasis: A Review. Theriogenology. 42: 557-570.

Sekoni VO, Njoku CO, Kumi-Diaka J, Saror DI (1990). Pathological changes in male genitalia of cattle infected with *Trypanosoma vivax* and *Trypanosoma congolense*. Brit. Vet. J, 146:175-180.

Silva TMF, Olinda RG, Rodrigues CMF, Camara ACL, Lopes FC, Coelho WAC, Ribeiro MFB, Frieitas CIA, Teixeira MMG, Batista JS (2013). Pathogenesis of reproductive failure induced by *Trypanosoma vivax* in experimentally infected pregnant ewes. Vet. Res, 44:1

Victor I, Sackey AKB, Natala AJ (2012). Penile protrusion with hemorrhages and prepucial inflammation in pigs experimentally infected with *Trypanosoma congolense*. J. Anim. Prod. Adv, 2: 297-302