

Full Length Research Paper

Phylogenetic analysis of dematiaceous fungi isolated from the soil of Guangdong, China

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To search for the dematiaceous fungi in nature, clarify their habitat and the environmental circumstances in which they may infect man, we studied the phylogenetic relationships of these isolates. 60 dematiaceous fungal strains out of 367 soil samples were isolated. They were further identified by molecular biological method. The phylogenetic relationships were demonstrated by using internal transcribed spacer (ITS) region of ribosomal DNA sequences. In the neighbor-joining (NJ) tree, *Phialophora* sp., *Cladophialophora chaetospira*, *Exophiala spinifera*, *Phaeococcomyces* sp. and *Exophiala eucalyptorum* formed cluster A, *Didymella bryoniae*, *Leptosphaeriaceae* sp., *Ascomycete* sp., *Microdiplodia hawaiiensis* and *Cochliobolus lunatus* formed cluster B, *Staninwardia suttonii*, *Cladosporium oxysporum*, *Cladosporium cladosporioides*, *Cladosporium* sp. and *Melanized limestone ascomycete* formed cluster C, and *Scolecobasidium tereum*, *Scolecobasidium humicola* formed cluster D. The phylogenetic relationships between cluster B, cluster C and cluster D were closer than that of cluster A. Dematiaceous species were found widely in Guangdong soil, and the distribution amounts were not in a specific pattern, the phylogenetic method based on the rDNA ITS sequence was proven to be a quick and accurate fungi identify method. There is a relationship between genetic distances and some biological habits of some strains, but lacking of connection was found between the genetic distances and the geographical factors.

Key words: Dematiaceous fungi, isolation, phylogenetic tree, internal transcribed spacer (ITS) region, ribosomal DNA (rDNA).

INTRODUCTION

With the increasing of invasive fungal infection (Marr et al., 2002; Pfaller and Diekema, 2004; Trick et al., 2002; Wisplinghoff et al., 2004; Baddley et al., 2001) emerging pathogens have been increasingly recognized as important pathogens. We face a marked shift in the epidemiological profile of fungal infections: new and emerging pathogens including some dematiaceous fungi (e.g. *Alternaria* sp., *Bipolaris* spp, *Curvularia* spp., *Cladophialophora* spp., *Exophiala* spp., *Phialophora*) are

increasingly being reported (Malani and Kauffman, 2007; Richardson and Lass-Flörl, 2008). Because risk-factors for these infections continue to increase in frequency, it is likely that the incidence of the emerging pathogens infections will continue to increase in the coming decades.

Dematiaceous or darkly pigmented fungi are uncommon causes of human disease but can be responsible for life-threatening infections in both immunocompromised and immunocompetent individuals (Revankar and Sutton, 2010; Schell, 1995). They are a heterogeneous group. The distinguishing characteristic common to all these various species is the presence of melanin in their cell walls, which imparts dark color to their conidia or spores

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Table 1. Localities, sites and number of soil samples collected in Guangdong, PR china.

Site	Localities					Total
	North *	Medio - region †			South ‡	
		Eastern§	Central ¶	Western #		
Field	14	64	79	32	42	231
Forest	4	9	17	2	3	35
Farm	2	1	11			14
Waterside		3	9	5	5	22
Road side	5	6	4	14	2	31
Factory		4	9			13
Others	1	7	9	2		19
Total	26	94	138	55	52	365

*: indicates the north region of Guangdong with the medio-subtropical region; †: the middle region of Guangdong with the south subtropical region; ‡: the south region of Guangdong with the north tropical region; §: the east of the middle region of Guangdong; ¶: the middle region of Guangdong; #: the west of the middle region of Guangdong. The west of the middle region of Guangdong with the south subtropical region is Zhaoqing Gaoyao West; the central of the middle region of Guangdong with the south subtropical region is Zhaoqing Gaoyao East to Heyuan; the east of the middle region of Guangdong with the south subtropical region is Heyuan East.

and hyphae. The colonies are typically brown to black in color as well. Dematiaceous fungi are commonly found in the soil and generally distributed worldwide (Montenegro et al., 1996; Dixon et al., 1980; Lopez et al., 2004). They are the etiologic agents of phaeohyphomycosis, chromoblastomycosis and mycetoma. Over 100 species and 60 genera of dematiaceous fungi have been implicated in human disease (Matsumoto et al., 1994).

As these diseases usually occur by the penetration of the causative agent through skin wounds, it is significant to search for the agents in nature, clarify their habitat and the environmental circumstances in which they may infect man. Agents (Dixon et al., 1980; Yegres et al., 1991) have been isolated such as *Phialophora* spp., *Cladosporium* spp., *Exophiala* spp., *Sporothrix* sp., *Wangiella dermatitidis*, *Bispora betulina*, and *Scytalidium lignicola*, which demonstrated the presence of pathogenic dematiaceous fungi in nature, although the identity of most of these strains has not been verified by molecular data. Nishimura (Nishimura, 1994) and Nishimura et al. (Nishimura et al. 1989) investigated the ecology of pathogenic fungi in natural and living environments in Colombia, Venezuela, Brazil, China and Japan and succeeded in isolating various species of pathogenic dematiaceous fungi including *Fonsecaea pedrosoi*, *Phialophora verrucosa* and *Exophiala spinifera*. They did not find the fungus *Cladophialophora* spp., which are the mainly causative agents of chromoblastomycosis in China.

The taxonomy and identification of dematiaceous fungi are difficult due to a lack of phenetic characters and high degree of morphological plasticity. In the present study, we isolated 60 dematiaceous fungal strains out of 367 soil samples; these were further identified by molecular biological method. If the phylogenetic relationship and the geographical distribution of dematiaceous fungi from soil

of Guangdong, PR china, are revealed, it will be useful for future study.

MATERIALS AND METHODS

Sample collection

There are three climatic zones in Guangdong: the central subtropical (Nanxiong, Lianshan, Lianxian and Shaoguan), the southern subtropical (Yingde, Meixian, Shantou, Guangzhou, Yangjiang), and the northern tropical (Zhanjiang, Xuwen). 365 samples were collected from the three climatic zones where four to ten collecting sites were set up randomly (Table 1). The work was done during autumn and winter (October 2006 to January 2007), the dry season in Guangdong. Samples were collected from the surface soil upward of 15 cm. Utilizing a spoon which was rinsed with sterile water after each use, approximately 25 g sample were placed in 100 ml plastic bottles containing a small crystal of paradichlorobenzene (for arthropod control), and returned to the laboratory for processing on the same day.

Isolation of dematiaceous

3 g of soil were transferred to a sterile 15 ml glass tube; 10 ml of sterile saline were then added, mixed by agitation for 1 min and set for 20 min, after which it was diluted to the concentration of the proportion of 1:100, when 0.2 ml was collected from the middle part of the soil suspension and placed on two plates. Then, the solution was poured on two media; potato dextrose agar (PDA) Rose Bengal respectively, both containing antibiotics (50 mg of chloramphenicol, 10⁶ units of penicillin, 200 mg of streptomycin and 200 mg of cyclohexamide per liter). Pulled a medium to 3 plates, sealed the plates with plastic film, left a hole of 3 to 5 mm. The plates were incubated at 26°C for 2 to 3 weeks. The suspected colonies were subculture on Sabouraud dextrose agar (SDA) at 25°C and checked grossly and microscopically. All isolates were further evaluated by molecular biology methods.

DNA extraction, PCR amplification

DNA was extracted using 6% In Sta Gene TM Matrix (BioRad,

Table 2. List of reference sequences.

GenBank accession	Name	Geography
EU035406.1	<i>C. chaetospora</i>	Germany
AB456580.1	<i>E. spinifera</i>	Japan: Osaka
AB456578.1	<i>P. europaea</i>	Japan: Osaka
AJ972801.1	<i>Phaeococcomyces</i> sp.	Turkey: Mediterranean
DQ092530.1	<i>Leptosphaeriaceae</i> sp.	USA: Kuli'ou'ou Beach Park in Hawaii Kai, Oahu, Hawaii
AF297228.1	<i>D. bryoniae</i>	USA
DQ923535.1	<i>S. suttonii</i>	Australia
EU272531.1	<i>C. tenuissimum</i>	Colombia: Andean paramo ecosystem
EU714392.1	<i>Scolecobasidium</i> sp.	Thailand: southern Thailand, Gulf of Thailand and Andaman Sea

U.S.A.). rDNA ITS domains (including part of 18S, ITS I, 5S, ITS II, part of 26S) were amplified in a Biometra T-Gradient Thermoblock (Germany) using primers ITS-4 (5'-TCCTCCGCTTATTGATATGC-3') and ITS-5 (5'-GGAAGTAAAAGTCGTAACAAGG-3'), which were described previously (Abliz et al., 2004). Each PCR mixture contained 5 µl of 10 × reaction buffer (Pharmacia), 4 µl 10 × dNTP, 0.2U Taq polymerase, 1 µl primer, and 2 µl DNA template solution. Ultrapure water was added to increase the volume to 50 µl. Each reaction mixture was heated to 95°C for 4 min, followed by 30 cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 60 s., followed by incubation for 10 min at 72°C.

Direct sequencing and phylogenetic analysis

Direct sequencing of PCR products was done with an ABI PRISM 3100 sequencer (ABI, America) after labeling with BigDye Terminator Cycle Sequencing Ready Reaction (Applied Biosystems, Foster City, California). The ITS sequences of reference sequence from GenBank collection (Table 2) and isolated dematiaceous fungal (Table 3) in this study were aligned by using Clustal W software. Phylogenetic tree was then constructed by the neighbor-joining (NJ) method in the Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0 (Tamura, 2007). Bootstrap analysis with the MEGA program was performed by taking 500 random samples from the multiple alignments (values > 50 are shown with the branches). The evolutionary distance between organisms is indicated by the horizontal branch length, which reflects the number of nucleotide substitutions per site along that branch from node to the endpoint.

RESULTS

Within 367 soil samples, 15 genera, 33 species and 60 dematiaceous fungal strains (including 14 pathogenic dematiaceous fungal strains) were isolated. All isolates (Table 3) were identified by molecular biological method. The rDNA ITS regions (including part of 18S, ITS I, 5S, ITS II, part of 26S) were successfully amplified from all the dematiaceous fungi by universal primers. The size of acquired PCR products ranged from 500 bp to 650 bp. Each strain of dematiaceous fungi tested was shown to have unique ITS base sequences, although some of them were very similar.

In the phylogenetic tree (Figure 1) constructed from data of 60 strains and 9 reference strains, *Phialophora*

sp., *Cladophialophora*, *Chaetospora*, *E. spinifera*, *Phaeococcomyces* sp. and *Exophiala eucalyptorum* formed cluster A, *Didymella bryoniae*, *Leptosphaeriaceae* sp., *Ascomycete* sp., *Microdiplodia hawaiiensis* and *Cochliobolus lunatus* formed cluster B, *Staninwardia suttonii*, *Cladosporium oxysporum*, *Cladosporium cladosporioides*, *Cladosporium* sp. and *Melanized limestone ascomycete* formed cluster C, and *Scolecobasidium tereum*, *Scolecobasidium humicola* formed cluster D. Obviously, the phylogenetic relationships between cluster A, cluster B and cluster C were more closer than that of cluster D.

DISCUSSION

Studies of an infectious disease usually approach from pathogens, so isolations and identifications of pathogens are the most important parts in the approach. The usual identification of fungi by the morphological method, combining with some biochemical approaches, processes the observation of colonial textures, shapes, and colors in culture mediums, and the inspection of conidiophores, morphologies and generations of conidia. However, it is difficult because many dematiaceous fungi appear in multi-morphologies, which the isolate may generate more than one kind of conidium or may be generated by various conditions. Therefore, it is not easy to determine whether a conidium is yielded by the multi-morphological fungi or by a mixture of fungi. To obtain an isolate with high purity, it requires sub-cultures from the similar culture medium. Even with the required conditions of sub-culture, some colonies are complicated to isolate. Furthermore the members of mitosporic fungi are taxonomically closely related, morphological identification of mitosporic fungi becomes more difficult. Early studies have shown that the results from the molecular biological identification of fungi are in accordance with of the morphology identification (Pechere et al., 1999). In this study, a series of colony complexes are involved. We identified all the strains by the molecular biological method and processed the morphology method to classify some isolates.

Table 3. List of strains isolated and source.

Strain name	Source	Climate zone
<i>E. pisciphila</i>	The bamboo grove	Medio-subtropical
<i>E. eucalyptorum</i>	The grassland	Western of south subtropical
<i>E. xenobiotica</i>	The drainage	Middle of south subtropical
<i>E. oligosperma</i>	The stream	Eastern of south subtropical
<i>E. mesophila</i>	The rice field	Eastern of south subtropical
<i>Exophiala</i> sp.	The roadside	Eastern of south subtropical
<i>E. dermatitidis</i>	The sideway	Western of south subtropical
<i>Phaeococcomyces</i> sp.	The roadside	Western of south subtropical
<i>Phaeococcomyces</i> sp.	The roadside	Western of south subtropical
<i>Phaeococcomyces</i> sp.	The hillside	Western of south subtropical
<i>Phaeococcomyces</i> sp.	The root of tree	Eastern of south subtropical
<i>Phaeococcomyces</i> sp.	The taro field	North-tropical
<i>Phialophora</i> sp.	The rice field	Western of south subtropical
<i>Phialophora</i> sp.	The rice field	Western of south subtropical
<i>C. chaetospora</i>	The bamboo grove	Medio-subtropical
<i>C. chaetospora</i>	The taro field	Medio-subtropical
<i>C. carrionii</i>	The root of tree	Medio-subtropical
<i>C. chaetospora</i>	The factory zone	Middle of south subtropical
<i>C. chaetospora</i>	The hill	Eastern of south subtropical
<i>C. devriesii</i>	The rice field	Eastern of south subtropical
<i>C. chaetospora</i>	The nursery	North-tropical
<i>C. chaetospora</i>	The sweet potato field	North-tropical
<i>C. chaetospora</i>	The roadside	Middle of south subtropical
<i>C. chaetospora</i>	The hillside	Western of south subtropical
<i>C. oxysporum</i>	The vegetable field	Middle of south subtropical
<i>C. cladosporioides</i>	The drainage	Middle of south subtropical
<i>Cladosporium</i> sp.	The vegetable field	Eastern of south subtropical
<i>C. cladosporioides</i>	The farmland	Eastern of south subtropical
<i>C. oxysporum</i>	The pinking field	North-tropical
<i>C. oxysporum</i>	The sweet potato field	North-tropical
<i>C. oxysporum</i>	The cucumber field	North-tropical
<i>S. tereum</i>	The pigsties	Medio-subtropical
<i>S. tereum</i>	The pineapple field	Western of south subtropical
<i>S. tereum</i>	The roadside	Western of south subtropical
<i>S. tereum</i>	The rice field	Western of south subtropical
<i>S. tereum</i>	The rice field	Medio-subtropical
<i>S. tereum</i>	The vegetable field	Middle of south subtropical
<i>S. tereum</i>	The crown field	Middle of south subtropical
<i>S. tereum</i>	The grassland	Middle of south subtropical
<i>S. tereum</i>	The drainage	Middle of south subtropical
<i>S. tereum</i>	The onion field	Middle of south subtropical
<i>S. tereum</i>	The Chinese eggplant field	Middle of south subtropical
<i>S. humicola</i>	The papaya field	Eastern of south subtropical
<i>S. humicola</i>	The chicken shed	Eastern of south subtropical
<i>S. tereum</i>	The grass land	Eastern of south subtropical
<i>S. tereum</i>	The vegetable field	Eastern of south subtropical
<i>S. tereum</i>	The vegetable field	Eastern of south subtropical
<i>S. tereum</i>	The vegetable field	Eastern of south subtropical
<i>S. suttonii</i>	The grassland	Western of south subtropical
<i>S. suttonii</i>	The roadside	Middle of south subtropical
<i>D. bryoniae</i>	The orchard	Western of south subtropical

Table 3. Contd.

<i>Capnocylium</i> sp.	The duck shed	Middle of south subtropical
<i>M. limestone ascomycete</i>	The factory zone	Middle of south subtropical
<i>Leptosphaeriaceae</i> sp.	The vegetable field	Middle of south subtropical
<i>Leptosphaeriaceae</i> sp.	The grassland	Eastern of south subtropical
<i>Ascomycete</i> sp.	The root of tree	Middle of south subtropical
<i>Ascomycete</i> sp.	The banana field	Middle of south subtropical
<i>M. Alistairii</i>	The rice field	North-tropical
<i>C. lunatus</i>	The Chinese potato field	Eastern of south subtropical
<i>M. hawaiiensis</i>	The drainage	Middle of south subtropical

Results indicated that dematiaceous species were found widely in Guangdong soil. However, the distribution amounts were not in a pattern. Defined by climatic zones, the western species of the southern subtropical are with the highest abundance and density; the eastern species of the southern subtropical are with the lowest abundance and density. The most found genus was *Scolecobasidium*. There were 17 samples found and it was 28% (17/60) of the positive results. There is no detailed report found on its pathogenicity. Ten strains of *Cladophialophora* were found from 10 samples, which was 17% (10/60) of the positive results. Among them, the isolates of *Cladophialophora carrionii* were from the garden of Yuanshan Qingyuan. It has been the first time that *C. carrionii* which is the common pathogen for chromoblastomycosis in China was isolated from samples of Guangdong environment. Seven strains of *Exophiala* were recognized, including *E. dermatitidis*, *E. xenobiotica*, *E. oligosperma*, *E. pisciphila*, *E. mesophila*, *Exophiala* sp., *E. eucalyptorum*. Except *E. eucalyptorum*, the rest strains were common pathogens. *Phaeococcomyces* is a member of black yeast, which is hard to identify because its confusion on the morphology. Five strains were found from the experiment, but the species was not determined. Two isolates of *Phialophora parasitica* were found. There are 25 members in *Phialophora* of which five species are human pathogens, including *P. verrucosa*, *P. richardsiae*, *P. repens*, *P. parasitica*, and *P. cyanescens* (Park et al., 2005). The common pathogen *P. verrucosa* of chromoblastomycosis was not found in this experiment which may be because it intends to distribute in the cold zone (Liu et al., 2004). *Cladosporium*, dispensing widely, is the saprophyte found usually in soil and on plants, and is also the pathogen for plants. Some of *Cladosporium* are relative to human infections (Chew et al., 2009; Gugnani et al., 2006). Seven strains were found in this experiment, including four strains of *C. oxysporum*, two strains of *C. cladosporioides*, and one strain of *C. sphaerospermum*. The three species can cause human phaeohyphomycosis. *C. oxysporum* presents in the warm condition (Mckemy and Morgan, 1991). It is compliant with the four strains that have been from farms of Zhuhai and

Zhanjiang Leizhou. The locations are warmer than the Chinese lettuce field in Meizhou from which the mould of the *C. sphaerospermum* strain is Meizhou is in the further northern region. One strain of *D. bryoniae*, plant pathogen, was isolated. Some studies have shown the fungi of *Didymella* are related to asthma (Pulimood et al., 2007), because the amount of the fungi increases during the storm season, which may lead to asthma.

There is no human disease reported for the rest eleven findings, including two strains of *S. suttonii*, one strain of *Capnocylium* sp., one strain of *Melanized limestone ascomycete*, two strains of *Leptosphaeriaceae* sp., two strains of *Ascomycete* sp., one strain of *Mycosphaerella Alistairii*, one strain of *C. lunatus*, one strain of *Micro-diplodia hawaiiensis*. The major pathogen *Fonsecaea pedrosoi* of chromoblastomycosis was not found in the experiment. Are the substances foci suitable for *Fonsecaea pedrosoi* growing not the soil but other material, such as plant, litter and wheat stalk? May the condition factors of the experiment, such as culture medium, temperature or other involved fungi also play the role to inhibit the species survive? Further studies will be needed to address the above questions.

Using rDNA ITS sequences from 60 newly isolated strains and 9 reference strains, we constructed phylogenetic tree of dematiaceous fungi. The phylogenetic method based on the ITS rDNA sequence to identify fungi agreed with the morphological method, even if more quick and accurate. The NJ tree indicates the relationships between genetic distances and some biological habits of strains, for example, the strains that can cause diseases in human and animal often group together (*Cladophialophora carrionii*, *Cladophialophora devriesii*, *Cladophialophora oxysporum*, *Cladophialophora cladosporioides*, *E. dermatitidis*, *E. xenobiotica*, *E. oligosperma*, *E. pisciphila*, *E. mesophila*, *D. bryoniae*), on the other hand, the strains that can cause diseases in plants usually get into other crowd. There is no evidence showing that the sorting relates to the geographical location, while the same kind strains from different regions reveal similar genetic distances. Therefore, the strains within the crowd or with closer genetic distance would be also with potential pathogenicity. Development

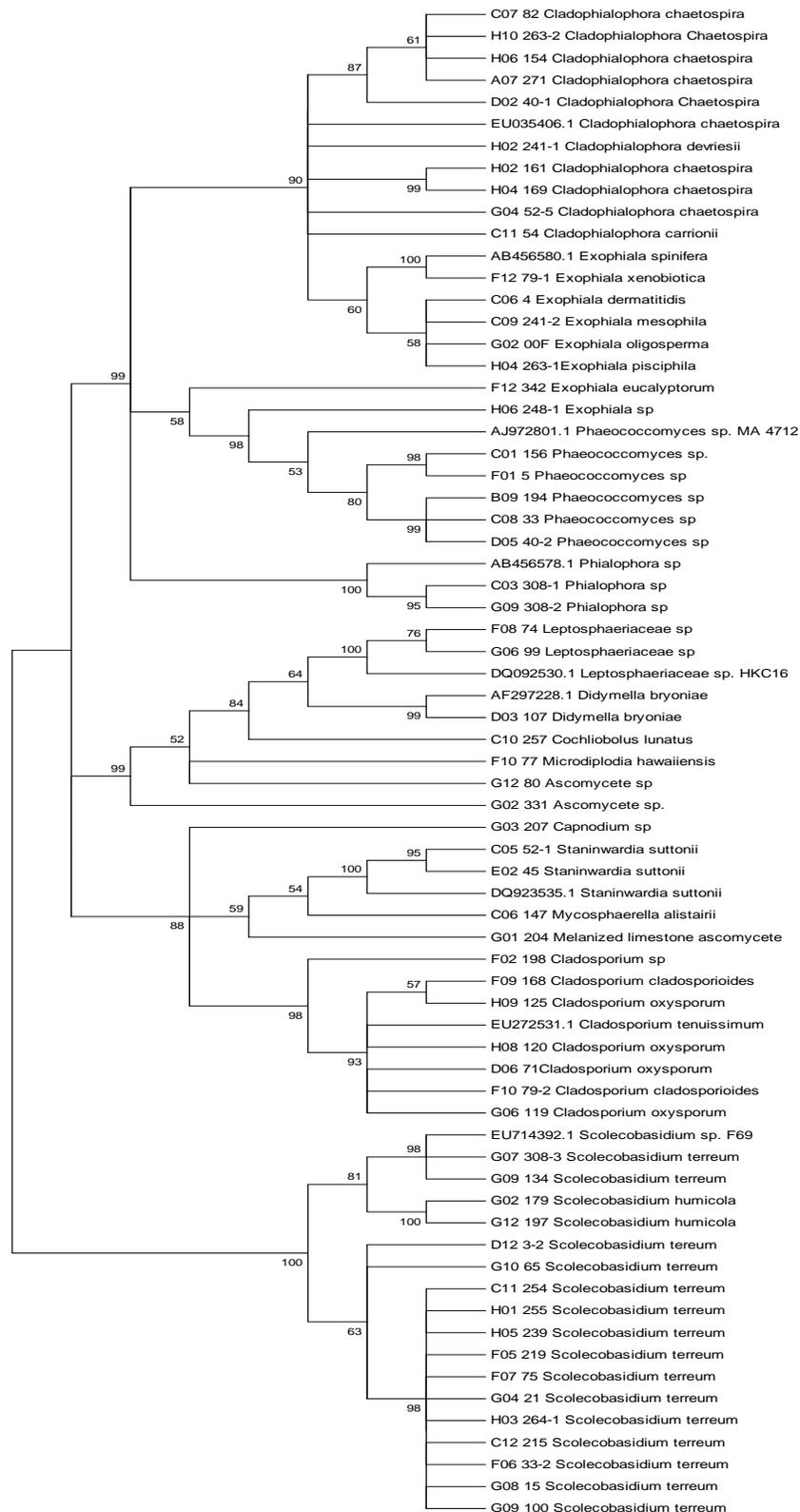


Figure 1. Phylogenetic tree constructed by neighbor-joining (NJ) method. The tree was constructed using 500 bootstrap replications (values > 50 are shown with the branches). The evolutionary distance between organisms is indicated by the horizontal branch length, which reflects the number of nucleotide substitutions per site along that branch from node to the endpoint.

of NJ tree would not only effect on identifications of fungal strains but would also play a role on guiding the studies on some biological habits of the strains.

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