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Full Length Research Paper

Efficacy of fungicides, botanicals and bioagents against *Rhizoctonia solani* inciting leaf blight on turmeric (*Curcuma longa* L.)

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Leaf blight caused by *Rhizoctonia solani* is one of the most serious diseases that cause damage to turmeric crop. Fifty percent disease incidence was observed in turmeric growing areas of Andhiyur (Erode district) and Mettur (Salem district) of Tamil Nadu, India. The antifungal efficacy of botanicals, bioagents and fungicides were tested against *R. solani in vitro*. It was observed that seed extract and oil cake of *Madhuca longifolia* suppressed the mycelia growth (34.81; 49.63%), sclerotial formation (100%) and production of biomass (2.18; 2.41 mg) of *R. solani* followed by leaf extracts of *Azadirachta indica* and *Littorina littorea*. Among the bioagents native isolate of BSoya1 of *Bacillus* spp., Cg1 of *Chaetomium* spp., Pf1 of *Pseudomonas* spp. and Tv1 of *Trichoderma* spp. shown more than 50% antagonistic effect on mycelia growth and 100% on sclerotial formation. The complete fungal growth inhibition was observed in Nativo and Bavistin fungicides amended medium. Then, the basal soil application of mahua cake and *T. viride* in the ratio of 1:2 (1 g of mahua cake mixed with 2 g *T. viride* per kg of pot soil) followed by foliar spray of Nativo @ 0.5% significantly mitigated the leaf blight (8.6%) over the inoculated control (50.51%) in pot culture experiments.

Key words: Turmeric, efficacy, fungicides, bioagents, botanicals, Rhizoctonia solani.

INTRODUCTION

Turmeric (*Curcuma longa* L.) known as the "golden spice", is one of the most important herbs in tropical and subtropical countries. It is a rhizomatous perennial plant of the ginger family, Zingiberaceae (Dixit et al., 2002). Turmeric is used as condiment, dye, drug and cosmetic in addition in religious ceremonies (Gescher et al., 2005). India is the leading producer, consumer and exporter of turmeric in the world and

contributes about 80% of the world production and 60% to the total trade (Parthasarthy et al., 2007).

Several fungal, bacterial and viral diseases are reported in turmeric (Hudge and Ghugul, 2010). Among the diseases, leaf blight caused by *Rhizoctonia solani* is an emerging disease to the turmeric crop (Roy, 1992). The pathogen is known to be soil borne and sclerotia are often found in

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Table 1. Standard evaluation system for screening rice sheath blight.

Grade	Description
0	No incidence
1	Less than 1% sheath area affected
3	1-5% sheath area affected
5	6-25% sheath area affected
7	26-50% sheath area affected
9	51-100% sheath area affected

the soil. Limited information is available on sustainable management and the disease control is generally with chemical applications. The increasing awareness of fungicide related hazards has emphasized the need for adopting biological methods as an alternative disease control method, which is also ecofriendly (Khare et al., 2010).

Biological control appears to be the best solution for long term sustainability and effective management of soil borne disease which can considerably minimize the disease. Successful management of *R. solani* on various crops by bioagents has been previously reported (Lahlali and Hijri, 2010; Seema and Devaki, 2012; Srinivas et al., 2014).

Hence, considering economic importance of the crop and the disease, the present investigation was undertaken to conduct the disease survey, to evaluate the efficacy of fungicide, botanicals and bioagents against *R. solani* and to find out the suitable management practice to mitigate the disease.

MATERIALS AND METHODS

Survey

Turmeric growing villages of Tamil Nadu, India were surveyed to assess the leaf blight disease incidence and severity on turmeric. The disease scoring was done in 50 plants at three different locations in a single village. The disease rating scale (0 to 9) was followed for the assessment of incidence and severity of leaf blight disease. The blight disease incidence was assessed and expressed in percent disease index (PDI) and also relative lesion height (RLH) was assessed based on the lesion height (Sharma et al., 1990). The following formula was used to calculate PDI.

$$PDI = \frac{Sum \ of \ individual \ ratings}{Total \ number \ of \ plants \ observed} \times \frac{100}{Maximum \ grade}$$

Standard evaluation system for screening rice sheath blight (0-9 scale) is shown in Table 1

The following formula was used to calculate relative lesion height (RLH):

Isolation of pathogen

R. solani was isolated from the naturally infected leaf samples through tissue segment method collected from different places of Tamil Nadu. Small portions of sheath showing typical lesions were cut into small bits of 0.5 - 1 cm, surface sterilized with 0.1% mercuric chloride and rinsed three times with sterile distilled water. Then, they were transferred to Petri dish containing potato dextrose agar (PDA) medium aseptically (Riker and Riker, 1936). The pure culture of the pathogen was obtained by single hyphal tip technique and the culture was maintained on PDA slants to carry out further studies (Taheri et al., 2007).

Pathogenicity tests

Diseases free (healthy) turmeric rhizomes (Erode local 8) were planted in 30 cm diameter earthen pots containing pot mixture at two rhizomes per pot. Actively growing plants at 30 days after planting (DAP) were inoculated with *R. solani* by inserting young immature sclerotia (two sclerotia per sheath) and incubated for seven days for the development of typical blight symptoms on the plants (Sriram et al., 1997).

In vitro effect of botanicals on radial mycelial growth of R. solani

Leaves of nine plants viz., Lawsonia inermis L. (Maruthani), Ocimum tenuiflorum L. (Thulasi), Azadirachta indica L. (Neem), Morinda citrifolia L. (Noni), Vinca major (Periwinkle), Gloriosa suberba (Glory lily), Justicia adhatoda (Adathoda), Vitex nigundo (Nochi) and Madhuca longifolia (Mahuva seed extract) and seven oilcakes viz., mahuva (Mahua longifolia L.), pungam (Pongamia glabra L.), sesamum (Sesamum indicum L.), groundnut (Arachis hypogaea L.), castor (Ricinus communis L.), neem (Azadirachta indica L.) and sunflower (Helianthus annuus L.) were collected and tested for their efficacy against R. solani by poisoned food technique. Fresh plant leaves and seed extracts were prepared by cold water extraction method (Shekhawat and Prasad, 1971). The leaves were first washed with distilled water and finally with sterile water and the oilcakes were first soaked in sterile distilled water at the rate of one g in 1 ml of water and kept overnight. Then, ground in pestle and mortar by adding sterile water at the ratio of 1:1 (w/v). The macerate was squeezed using cotton to get the extract. The extract was strained through two layers of muslin cloth and finally through Whatman No.1 filter paper and this formed the standard plant extract solution (100%). This was further diluted with sterilized distilled water (v/v) to have the required concentrations (10, 15, and 20%). The PDA medium was mixed with different concentrations viz., 10, 15 and 20% of plant extracts and oil cakes. Mycelial plug of the pathogens (7 mm) was placed at the centre of each Petri plate and incubated at 28 ± 1°C and three replications were maintained for each treatment. The experiment was arranged in a completely randomized block design. The fungus grown on PDA without any plant extracts and oil cakes served as control. The growth diameter was recorded and the percent inhibition was calculated.

In vitro efficacy of botanicals on biomass production of R. solani

The effect of plant products on mycelia growth of the pathogen in liquid medium was done by the method suggested by Neslihan et al. (2008). Different concentrations (10, 15, and 20%) of plant extracts and oil cakes were amended with the 100 ml of potato dextrose broth. A mycelial plug (7 mm) was transferred to the flasks which contain different concentrations of plant extracts and incubated at

28±2°C for one week. Three replications were maintained for each treatment. The experiment was arranged in a completely randomized block design. The fungal mycelial mat was collected, dried and mycelium weight recorded.

In vitro efficacy of bioagents on mycelial growth and sclerotial formation of R. solani

The native isolates of bio-agents viz., Bacillus isolates (EPCO8, EPCO44, EPCO26, BG1, Bsoya1, Bsoya2, BG2, BC3, EPCO81, EPCO PF, BC1, EPCO78, BC9, BC8, BC7, BC6, EPCO P6); Pseudomonas isolates (Pf1, KAU46, APF6, Pf3, PC3, 18P, 6P, PC4 and Pf26); Chaetomium isolates (Cg1, Cg2, Cg3, Cg4, Cg5, Cg6 and Cg249) and Trichoderma viride isolates (Tv1, Tv2, Tv3, Tv4, Tv5, Tv6 and Tv7) were used for testing their efficacy against R. solani by dual plate technique (Dennis and Webster, 1971). The bioagents as well as the pathogen were inoculated equidistant on PDA medium aseptically and incubated at 28 ± 1°C. In each case, three replications were maintained for each isolates along with control. The experiment was arranged in a completely randomized block design. After obtaining the maximum growth in the control, the observation on radial growth of pathogen was taken and percent inhibition was worked out using Vincent's formula. The sclerotial formation was recorded on 7th and 14th days after inocu-lation.

In vitro efficacy of bioagents on biomass production of R. solani

The effect of culture filtrates of the native isolates of bioagents viz., Bacillus isolates (EPCO8, EPCO44, EPCO26, BG1, Bsoya1, Bsoya2, BG2, BC3, EPCO81, EPCO PF, BC1, EPCO78, BC9, BC8, BC7, BC6, EPCO P6); Pseudomonas isolates (Pf1, KAU46, APF6, Pf3, PC3, 18P, 6P, PC4 and Pf26); Chaetomium isolates (Cg1, Cg2, Cg3, Cg4, Cg5, Cg6 and Cg249) and Trichoderma viride isolates (Tv1, Tv2, Tv3, Tv4, Tv5, Tv6 and Tv7) on the biomass production of R. solani was studied as per method given by Dennis and Webster (1971). Sterilized potato dextrose (PD) broth (100ml) was taken in 250 ml flask and inoculated with mycelial plugs (7 mm) of the biocontrol native isolates taken from the edge of four day old culture. Inoculated flasks were incubated at 28 ± 2°C for one week and the cultural filtrate was extracted by centrifuging the content at 10000 rpm for 30 min and the culture filtrate was collected. The culture filtrate of bioagents were mixed with PD broth at a rate of 1:4 (v/v) and inoculated with mycelial plugs of pathogen (7 mm), then incubated at 28±2°C for seven days. Three replications were maintained for each isolates. The experiment was arranged in a completely randomized block design. After the incubation period, fungal mycelial mat was collected and dried. The dried mycelial weight was recorded.

In vitro efficacy of fungicides on mycelial growth of R. solani

Evaluation of fungicides was carried out for their efficacy to inhibit the mycelial growth of R. solani isolate by "poisoned food technique" as described by Sharvelle (1961). The fungicides viz., azoxystrobin (Mirador 250 SC), tebuconazole (Orius 25.9% EC), tridemorph (Calixin 80% EC), fosetyl-Aluminium (Aliette WP 80), trifloxystrobin (Flint 50 WG), fenamidone 10% + mancozeb 50% (Sectin 60 WG), Carbendazim (Bavistin 50WP), tebuconazole 50% + trifloxystorbin 25% (Nativo WG 75) were used at 10, 50, 100, 250, 500, 750 and 1000 ppm concentration. Mycelial plugs of the pathogens (7 mm) taken from a seven day old culture was placed at the centre of each Petri plate and incubated at $28 \pm 1^{\circ}$ C. Three

replications were maintained for each treatment. The experiment was arranged in a completely randomized block design. The fungus grown on PDA without any fungicides served as control. The radial growth of the colony was recorded when maximum growth was observed in the control and it was calculated by using the following formula.

$$I = \frac{C - T}{C} \times 100$$

Where, I = Percent inhibition; C = radial growth in control; T = radial growth in treatment.

The sclerotial formation was recorded on 7th and 14th days after inoculation.

Management of leaf blight of turmeric incited by *R. solani* – Greenhouse studies

Studies were conducted to test the efficacy of effective treatment against leaf blight disease of turmeric under greenhouse condition with nine treatments with fungicide, bioagents and botanical. The Erode local 8 was sown in the pots containing 5 kg/pot of sterilized soil. The pathogen was inoculated 30 DAP. Three replications were maintained for each treatment. The experiment was arranged in a completely randomized block design. The blight incidence was recorded on 40 days after sowing and the effectiveness of the treatments on the intensity of leaf blight disease was observed seven days after inoculation, with a 0-9 scale of the Standard Evaluation System for rice, IRRI (2002) and expressed as percent disease index (PDI).

RESULTS AND PLUGUSSION

Survey and pathogenicity studies of *R. solani* in turmeric

Among the fifteen villages surveyed, the turmeric plants at Andhiyur, Erode district recorded the maximum of 54.5% disease index (PDI) of leaf blight which was followed by Mettur in Salem district which recorded 44.3 PDI. The disease incidence was prevalent in only four villages viz., Annur (Rs1), Andhiyur (Rs2), Mettur (Rs3) and Paramathivellur (Rs4) and the incidence ranged from 6.4 to 54.5 PDI (Table 2). The results revealed that placing of sclerotia in the injured sheath region exhibited the maximum lesion length of 16.89 cm and leaf blight lesion length. The symptom observed under field condition was very typical when compared with the artificially inoculated plants. Though the symptoms produced by all the isolate were similar, the most virulent Andhiyur, Erode isolate was highly virulent as compared to other isolates and produced larger lesion length.

Efficacy of fungicides on turmeric leaf blight pathogen (*R.* solani)

Nativo and Bavistin were individually effective against the pathogen even at the lowest concentration of 10 ppm by completely inhibiting the mycelia growth and sclerotia

Table 2. Survey of leaf blight of turmeric incited by *R. solani*.

Isolates	Village	District	*PDI	*Relative Lesion Height (RLH)
Rs1	Annur	Coimbatore	6.4	7.14
Rs2	Andhiyur	Erode	54.5	16.67
Rs3	Mettur	Salem	44.3	11.63
Rs4	Paramathivellur	Namakkal	28.6	8.8
	SEd	l	0.5279	0.2606
	CD(.0	5)	1.2173	0.6009

^{*} Mean of three replications.

Table 3. Efficacy fungicides on the mycelia growth of *Rhizoctonia solani*.

Formulation a		Мус	elia growth inl	nibition over co	ontrol (%)* (ppr	n)	
Fungicides	10	50	100	250	500	750	1000
Mirador 250 SC	4.21	100.00	100.00	100.00	100.00	100.00	100.00
Orius 25.9% EC	27.4	100.00	100.00	100.00	100.00	100.00	100.00
Calixin 80 % EC	27.44	100.00	100.00	100.00	100.00	100.00	100.00
Aliette WP 80	0.00	0.00	0.74	3.70	19.26	19.83	48.15
Flint 50 WG	59.63	87.04	94.44	100.00	100.00	100.00	100.00
Sectin 60 WG	0.00	41.85	72.59	72.89	87.41	100.00	100.00
Bavistin 50WP	100.00	100.00	100.00	100.00	100.00	100.00	100.00
Nativo WG 75	100.00	100.00	100.00	100.00	100.00	100.00	100.00
Control	0.00	0.00	0.00	0.00	0.00	0.00	0.00
	Т	ppm					
SEd	0.04292	0.03591					
CD (0.05)	0.08487	0.07101					

^{*} Mean of three replications.

production. It was significantly superior over other fungicides and on par with each other. It was followed by Mirador, Orius and Calixin which exhibited total inhibition at 50 ppm (Table 3). The least effective fungicides were Aliette, Flint and Sectin (48.15% inhibition at 1000 ppm). The same trend was observed in the liquid culture technique in which the two fungicides *viz.*, Nativo and Bavistin completely inhibited the mycelia growth at the lowest concentration of 10 ppm (Table 3). Aliette had recorded sclerotial formation of 86 numbers on 14 DAI at 100 ppm. As the concentration of fungicides increased, the sclerotial formation was arrested (Table 4).

Fungicides *viz.*, trifloxystrobin + tebuconazole, tebuconazole and propiconazole showed higher level of efficacy against *R. solani* of rice in laboratory conditions (Hunjan et al., 2011). Among the new formulations, Filia and Nativo were equally effective in controlling sheath blight (Swamy et al., 2009). In the present study, among different fungicides screened for, *R. solani* Nativo and Bavistin were individually effective against the pathogen in inhibiting the mycelia growth and sclerotial production even at the lowest concentration of 10 ppm.

In vitro antagonistic effect of bioagents against R. solani

Bacillus sp.

Bacillus isolates, BSoya1 and BC3 respectively recorded 32 and 33 mm colony diameter which accounted for 58 and 57% mycelia growth inhibition over the control. BSoya1 and BC3 Bacillus isolates were significantly superior and on par with each other in controlling the mycelia growth of R. solani. It was interesting to note that the sclerotial formation was completely inhibited in eleven isolates. The remaining six isolates of Bacillus viz., EPCO26, BG1, BSoya2, BG2, BC7 and BC8 had sclerotial formation. Among the seventeen isolates a maximum number of 84 sclerotia were observed in the BG2 as against the control which recorded 228.67 sclerotia (Table 5). All the Bacillus isolates reduced the mycelial weight of the pathogen over the control. The least biomass production was seen with the Bacillus isolate EPCO26 (0.52 mg) followed by Bsoya1 (0.65 mg)

Table 4. Effect of fungicides on the *Rhizoctonia solani* sclerotial formation.

Fungicides	*Sclerotial formation (DAI)	10 ppm	50 ppm	100 ppm	250 ppm	500 ppm	750 ppm	1000 ppm
Miradar 250 CC	7	2.67	-	-	-	-	-	-
Mirador 250 SC	14	8	-	-	-	-	-	-
Orius 25.9% EC	7	-	-	-	-	-	-	-
Olius 25.9% EC	14	-	-	-	-	-	-	-
Calixin 80 % EC	7	6.33	-	-	-	-	-	-
Calixili 60 % EC	14	13.67	-	-	-	-	-	-
Aliette WP 80	7	20	40	78.67	-	-	-	-
Allette WP 60	14	25	50	86.00	-	-	-	-
Flint FO MC	7	-	-	-	-	-	-	-
Flint 50 WG	14	-	-	-	-	-	-	-
Sectin 60 WG	7	96.00	-	-	-	-	-	-
Secun 60 WG	14	101.67	-	-	-	-	-	-
Bavistin 50WP	7	-	-	-	-	-	-	-
Davistili SUVVP	14	-	-	-	-	-	-	-
Nativo WG 75	7	-	-	-	-	-	-	-
Nativo VVG 75	14	-	-	-	-	-	-	-
Control	7	164	154.9	134	158	191	145	169
Control	14	189	162	173	169	204	163	182

^{-:} Sclerotia not formed. DAI: Days after inoculation.

and BC3 (0.78 mg) (Table 6). Similar antagonistic effect of Bacillus spp. against R. solani infecting many other crops was reported by several workers. Calvo et al. (2010) reported that, Bacillus spp. from rhizosphere region of potato showed high antagonistic effect against R. solani causing various diseases in crops plants. Padaria and Kapoor (2011) reported that, Bacillus pumillus obtained from rice field exhibited antagonistic activity against R. solani infecting various crops. Bacillus sp. strain 916, isolated from the soil showed strong activity against R. solani causing rice sheath blight (Wang et al., 2012). Bacillus sp. shows inhibition against the growth of a wide range of plant fungal pathogens like R. solani and Fusarium verticillioides by producing morphological modifications on the pathogenic fungi hyphae like total collapse of the cell. In the present study, among the 17 Bacillus isolates, BSoya1 and BC3 were significantly superior over other antagonists and it also reduced the mycelial weight and sclerotial production of the pathogen.

Pseudomonas spp.

Among nine isolates of *Pseudomonas* spp., Pf1 strain was significantly superior and recorded 59.22% mycelial growth inhibition and 1.18 mg biomass production. This was followed by KAU46 which recorded 56.7 mm colony diameter which accounted for 37% mycelial growth

inhibition over control (Table 6). After 14 days of inoculation, the sclerotial count was recorded and it was observed that except three Pseudomonas strains viz., KAU46, Pf3 and Pf26, all the other strains supported sclerotia formation of R. solani but the production was drastically reduced. Among the various strains of Pseudomonas, the maximum number of 142 sclerotia was recorded in PC3 strain as against the control which recorded 198.67 numbers of sclerotia. Though the Pseudomonas isolate KAU 46 exhibited 56.7 mm mycelial growth, it had completely inhibited the sclerotial formation in R. solani. Pf 26 has recorded full growth of the test pathogen but it had also inhibited the sclerotial formation (Table 7). Hence the mycelial growth could not be correlated with the sclerotial formation or inhibition. Similarly, Zachow et al. (2010) found that P. fluorescens L13-6-12 applied to the R. solani infected soil in sugar beet, formed large microcolonies consisting of hundreds of cells.

Chaetomium sp.

On studying the interaction between the *Chaetomium* and the pathogen, the isolate Cg1 showed the maximum percent inhibition (61.89%) which was significantly higher than other isolates (Table 6). It was followed by Cg249 (46.67%) and Cg2 (45.22%) and they were on par with each other. The reduction in mycelial biomass was also

Table 5. Antagonistic effect of *Bacillus* isolates against *Rhizoctonia solani*.

Bacillus	*Colony diameter	*Mycelial growth inhibition over	*Biomass production	*Sclerotial formation (nos.)		
isolates	(mm)	control (%)	(mg)	7 DAI	14 DAI	
EPCO8	43.00	47.00	1.02	-	-	
EPCO44	50.70	39.30	1.19	-	-	
EPCO26	43.30	46.70	0.52	-	2	
BG1	45.00	45.00	1.08	-	3	
BSoya1	32.00	58.00 ^a	0.65	-	-	
BSoya2	41.70	48.30	0.88	13	19	
BG2	41.70	48.30	0.97	38	84	
BC3	33.00	57.00	0.78	-	-	
EPCO81	48.70	41.30	1.19	-	-	
EPCO Pf	73.00	17.00	2.52	-	-	
BC1	74.30	15.70	2.31	-	-	
EPCO78	55.00	35.00	1.28	-	-	
BC9	43.00	47.00	1.02	-	-	
BC8	90.00	0.00	2.79	-	3	
BC7	90.00	0.00	2.56	36	73	
BC6	63.30	26.70	1.29	-	-	
EPCO P6	52.70	37.30	1.25	-	-	
Control	90.00	0.00	3.65	175.67	228.67	
SEd	1.7825	0.0978	0.0648			
CD(.05)	3.6155	0.1985	0.1314			

^{*}Mean of three replications. Means in a column followed by same superscript letters are not significantly different and the means are compared with LSD. Sclerotia not formed. DAI- Days after inoculation.

Table 6. Antagonistic effect of Pseudomonas isolates against Rhizoctonia solani.

Pseudomonas	*Colony diameter	*Mycelial growth inhibition over	*Biomass production	*Sclerotial formation (nos.)		
isolates	(mm)	control (%)	(mg)	7 DAI	14 DAI	
Pf1	36.7	59.22	1.18	1	2	
KAU46	56.7	37.00	1.26	-	-	
APF6	62 [.] 0	31.11	2.12	3	3	
Pf3	76.7	14.78	2.52	-	-	
PC3	78.7	12.56	2.49	45	142	
18P	90.0	0.00 ^f	3.59	43	50	
6P	90.0	0.00	3.65	3	5	
PC4	90.0	0.00	3.52	10	14	
Pf26	90.0	0.00	3.63	-	-	
Control	90.0	0.00	3.65	122.67	198.67	
SEd	1.9772	0.0843	0.2708			
CD (0.5)	4.1244	0.1759	0.5649			

^{*}Mean of three replications. Means in a column followed by same superscript letters are not significantly different and the means are compared with LSD. - Sclerotia not formed. DAI- Days After Inoculation.

recorded with Cg1 (1.39 mg) and the isolate was individually significantly superior when compared with other isolates. It was followed by Cg249 (1.99 mg) and

Cg2 (2.01 mg). All the isolates drastically reduced the number of sclerotial formation over control. The isolates Cg2 completely inhibited the sclerotial formation. The

Table 7. Antagonistic effect Chaetomium isolates against Rhizoctonia solani.

Chaetomium	*Colony	*Mycelial growth inhibition	*Piomocs production (mg)	*Sclerotia	*Sclerotial formation (nos.)		
isolates	diameter (mm)	over control (%)	*Biomass production (mg)	7DAI	14 DAI		
Cg1	34.30	61.89	1.39	28	54		
Cg2	49.30	45.22	2.01	-	-		
Cg3	51.70	42.56	2.10	3	4		
Cg4	56.70	37.00	2.30	3	6		
Cg5	65.30	27.44	2.58	5	12		
Cg6	52.70	41.44	2.12	6	10		
Cg249	48.00	46.67	1.99	36	74		
Control	90.00	0.00	3.65	113.02	132		
SEd	1.4269	0.0979	0.0624				
CD(.05)	3.0250	0.2075	0.1322				

^{*}Mean of three replications. Means in a column followed by same superscript letters are not significantly different and the means are compared with LSD. Sclerotia not formed. DAI- Days after inoculation.

Table 8. Antagonistic effect Trichoderma viride isolates against Rhizoctonia solani.

Trichoderma	*Colony	*Mycelial growth	*Biomass	*Sclerotial fo	ormation (nos.)
isolates	diameter (mm)	inhibition over control (%)	production (mg)	7 DAI	14 DAI
Tv1	19.7	78.11	0.40	-	-
Tv2	43.3	51.89	1.42	-	-
Tv3	38.7	57.00	1.10	-	-
Tv4	41.0	54.44	1.74	-	-
Tv5	35.0	61.11	1.41	-	-
Tv6	39.7	55.89	1.61	-	-
Th	49.7	44.78	2.01	-	-
Control	90.0	0.00	3.65	97	121
SEd	1.0212	0.1080	0.0577		
CD(.05)	2.1648	0.2290	0.1224		

^{*}Mean of three replications. Means in a column followed by same superscript letters are not significantly different and the means are compared with LSD. Sclerotia not formed. DAI- Days after inoculation.

maximum sclerotial production was recorded in the control (132 numbers), where as the minimum sclerotial production was seen in the isolate Cg 3 (4 numbers) (Table 7). Kaushik et al. (2010) reported that *Chaetomium globosum* showed good activity against *Sclerotinia sclerotiorum*, *Fusarium oxysporum* and *R. solani*.

Trichoderma viride

Among the seven isolates of *Trichoderma* spp. tested, maximum percentage of inhibition was recorded with the isolate Tv1. All the seven isolates inhibited the growth of *R. solani* in dual culture. The isolate Tv1 showed maximum percent inhibition of mycelial growth (78.11%) and it was individually significantly superior, followed by

Tv5 (61.11%) and Tv3 (57.00%). The least percent inhibition was observed with the isolate *T. harzianum* (44.78%) (Table 8). The Tv1 culture filtrate significantly reduced the mycelial biomass of the pathogen by recording 0.40 mg biomass production. It was followed by Tv3 (1.10 mg) and Tv5 (1.41 mg). All the isolates of *Trichoderma* inhibited the formation of sclerotia over control which recorded 121 sclerotia after 14 days of inoculation (Table 8).

Similarly, Khan and Sinha (2007) reported that, *T. harzianum* (rice leaf sheath isolate) was found most effective against *R. solani* in *in vitro* and glasshouse conditions. *T. harzianum* is well known biocontrol agents against several soil borne pathogens. Osman et al. (2011) showed that, *T. harzianum* was found effective in inhibiting the *R. solani* causing soya beans root rot. In the

present study, T. viride was found to be effective among the other antagonists. Among the seven isolates of Trichoderma spp tested, maximum percentage of inhibition was recorded with the isolate Tv1. All the seven isolates inhibited the growth, mycelial biomass and sclerotial production of R. solani in dual culture. Alamri et al. (2012) proved that the mechanism of T. harzianum JF419706 to suppress the pathogenic fungi viz., Alternaria alternate, Fusarium oxysporum, Exserohilum rostratum, Macrophomina phaseolina, Pythium ultimum, and R. solani was though competition for nutrients, mycoparasitizing and lysis of pathogen's cell walls. Harman et al. (2012) mentioned that Trichoderma spp. parasitize a range of phytopathogenic fungi.

In vitro effect of botanicals against R. solani

Madhuca longifolia (mahua) seed extract showed maximum inhibition of 34.81% at 20% concentration and it was found significantly superior to other extracts (Plate 1). This was followed by A. indica leaf extract which showed an inhibition of 20% at 20% concentration and O. tenuiflorum (thulasi) leaf extract, 18.15%. All the nine aqueous plant extracts were ineffective at 10% concentration (Table 9). The botanicals reduced the mycelial biomass of the pathogen over control. Extract from the M. longifolia seed significantly reduced the mycelial biomass (2.18 mg) over other treatments. It was followed by O. tenuiflorum (2.73 mg) and A. indica (2.95 mg) and they were on par with each other. The leaf extracts of A. indica, L. littorea and M. longifolia seed extract completely inhibited the sclerotial formation at all the three concentrations tested viz., 10, 15 and 20% (Table 8).

Plant extract are not only easy to prepare but also non-polluting and low priced as compared to commercial fungicides. This is supported by the work of Alabi and Olorunju (2004). In their studies, plants sprayed with neem seed extract gave yields higher than the plants sprayed with black soap and cow dung extract (Alabi and Olorunju, 2004). Gujar et al. (2012) reported that *A. indica* and *A. vera* showed inhibition of mycelial growth of the pathogen and can be utilized for the management of fungal diseases caused by the *Aspergillus niger*, *Aspergillus flavus*, *R. solani*, *Rhizoctonia bataticola*. In this present study, *M. longifolia* seed extract showed maximum inhibition and it was significantly superior to other plant extracts. All these plant extracts inhibited the mycelial weight and sclerotial formation completely.

All the oil cake extracts tested were not inhibitory at the lowest concentration of 10%. Mahua oil cake was the only extract which was inhibitory at 15% concentration recording 14.81% mycelial growth inhibition over the control. Among the seven extracts, mahua oil cake extract exhibited the maximum mycelial growth inhibition of 49.63% against the pathogen *R. solani* at 20% concentra-

tion (Plate 2). It was followed by neem oil cake (14.07%) and castor oil cake (8.15 %) (Table 11). Mahua oil cake and neem oil cake extract completely inhibited the formation of sclerotia at all concentrations. Among the treatments, maximum sclerotial formation was seen in the Petri plate poisoned with gingelly oil cake extract (117 numbers) at 20% concentration after 14 days of inoculation. The control recorded 199 numbers of sclerotia. It was interesting to note that the lower concentration of the seven oil cake extracts *viz.*, 10 and 15% had inhibited sclerotial formation in solid medium (Table 10). Alice et al. (1998) reported that the presence of antifungal principles present in the mahua cake extract (10%) was effective in combating the jasmine wilt incidence caused by *Sclerotium rolfsii*.

Management of leaf blight of turmeric under greenhouse studies

The effective treatments were culled out from the present research findings and used to perform the pot culture experiments to find out its efficacy against R. solani causing leaf blight of turmeric. Among the seven treatments, the basal soil application of mahua cake and T. viride in the ratio of 1:2 (1 g of mahua cake mixed with 2 g T. viride per kg of pot soil) followed by foliar spray of Nativo @ 0.5% significantly reduced the percent disease incidence over other treatments. This effective treatment recorded 8.6 PDI and a plant height of 160.8 cm on 180 DAP as against the inoculated control which recorded 50.51 PDI with a plant height of 138.6 cm. The disease reduction over control was 83.84% and the increase in plant height accounted for 11.00% (Table 11). The reduction in the turmeric sheath blight incidence and increased plant height in the treated plants in the present study is due to the biocontrol nature of all the treatments imposed.

Similarly, application of *T. harzianum* amended with organic fertilizer was more efficient alone in managing damping off disease (*R. solani*) in cucumber (Huang et al., 2011). Clove extract at a concentration of 4% as well as the chemical fungicide (Rizolex-T) significantly reduced the incidence of *R. solani* in pea (Al-Askar and Rashad, 2010). Addition of mustard oil cake in French bean (*Phaseolus vulgaris*) along with the inoculation of arbuscular mycorrhizal fungi (AMF) and *P. fluorescens* was found highly effective in reducing the root rot caused by *R. solani* (Neeraj and Kanchan, 2011).

Conclusion

Research presented here has expanded knowledge of the association of *R. solani* with turmeric crops in Tamil Nadu, India and about disease symptomology. Fungicide, botanical and bioagents were shown to vary in effectiveness at controlling mycelia growth, sclerotial

Table 9. In vitro effect of plants extracts on growth and sclerotial production of Rhizoctonia solani.

Plant products/	*Mycelial	growth inhibition o	ver control	*Biomass prod	luction	(mg)	*Sclerotial production				
concentration	10%	15%	20%	10%	15%	20%	DAI	10%	15%	20%	
Lawsonia inermis	0.00	0.00 ^j	2.59	3.65	3.65	3.54	7	3.00	6.33	97.33	
Lawsonia iriennis	0.00	0.00	2.59	3.05	3.00	3.54	14	5.33	9.00	100.33	
Ocimum tenuiflorum	0.00	0.00 ^j	18.15	3.62	3.61	2.73	7	-	194.33	-	
Ocimani tendinorum	0.00	0.00	10.10	3.02	5.01	2.70	14	-	205.00	-	
Azadirachta indica	0.00	6.30 ^f	20.00	3.62	3.56	2.95	7	-	-	-	
712aanaona maioa	0.00	0.00	20.00	0.02	0.00	2.00	14	-	-	-	
Morinda citrifolia	0.00	0.00	12.96	3.60	3.43	3.20	7	189.00	-	-	
oaa ooa	0.00	0.00		0.00	00	0.20	14	195.00	-	-	
Vinca major	0.00	0.00	3.70	3.63	3.60	3.58	7	-	-	-	
,							14	-	-	-	
Gloriosa superba	0.00	0.00	0.00	3.65	3.65	3.65	7	93	95	84	
•							14	98	100	89	
Justicia adhatoda	0.00	0.00	0.00	3.65	3.65	3.60	7	-	103	37	
							14 7	-	117	43 98.67	
Vitex nigundo	0.00	0.00	1.85	3.65	3.61	3.58	, 14	2	- 5	104.33	
							7	_	-	104.33	
Mahuva seed extract	0.00	14.81	34.81	3.62	3.50	2.18	, 14	_	_	_	
							7	192	186	163	
Control	0.00	0.00	0.00 ^J	3.65	3.65	3.65	, 14	232	227	263	
							17	202	<i>_</i> 1	200	
	Т	Concentration	Т	Concentration							
SEd	0.0284	0.01559	0.06410	0.03511							
CD(0.05)	0.0569	0.03118	0.12823	0.07023							

^{*} Mean of three replications. Means in a column followed by same superscript letters are not significantly different and the means are compared with LSD. - Sclerotia not formed. DAI- Days after inoculation.

 Table 10. In vitro effect of oil cake extracts on growth and sclerotial production of Rhizoctonia solani.

Oil cake/concentration	*Mycelial gr	owth inhibition (over control (%)	*Biom	ass prod (mg)	luction	*Sclerotial production			
	10%	15%	20%	10%	15%	20%	DAI	10%	15%	20%
Mahua	0.00	14.81 ^a	49.63 ^a	3.50 ^a	3.45 ^a	2.41 ^a	7 14	-	-	-
Neem	0.00	0.00 ^e	14.07 ^c	3.58 ^{ab}	3.53 ^{ab}	3.50 ^b	7 14	-	-	-
Groundnut	0.00	0.00 ^e	0.00 ^e	3.68 ^c	3.68 ^c	3.68 ^c	7 14	-	-	21 26
Gingelly	0.00	0.00 ^e	0.00 ^e	3.68 ^c	3.68 ^c	3.68 ^c	7 14	-	-	108 117
Sunflower	0.00	0.00 ^e	0.00 ^e	3.68 ^c	3.68 ^c	3.68 ^c	7 14	-	-	50 100
Castor	0.00	0.00 ^e	8.15 ^d	3.68 ^{bc}	3.58 ^{bc}	3.53 ^{bc}	7 14	-	-	15.67 24.6
Coconut	0.00	0.00 ^e	0.00 ^e	3.68 ^c	3.68 ^c	3.68 ^c	7	-	-	45.1

Table 10. Contd

control	0.00	0.00°	0.00 ^e	3.68°	3.68 ^c	3.68 ^c	14 7 14	- 167 184	- 154 179	50.1 176 199
	Т	Concentration	Т	Concentration						
SEd	0.02219	0.01359	0.06211	0.03803						
CD(0.05)	0.04461	0.02732	0.12488	0.07647						

^{*} Mean of three replications. Means in a column followed by same superscript letters are not significantly different and the means are compared with LSD. - Sclerotia not formed; DAI- Days after inoculation.

Table 11. Management of leaf blight of turmeric under greenhouse conditions.

		Plant he	eight* (cm)		L	eaf blight*
Treatment	45 DAP	90 DAP	135 DAP	180 DAP	(PDI)	Reduction over control (%)
Mahua cake	50.9 ^b	92.3 ^b	141.2 ^b	153.8 ^b	35.49 ^b	29.74 ^{ef}
T. viride	47.6 ^{de}	87.3 ^{de}	138.1 ^{de}	142.0 ^{de}	33.52 ^{de}	33.64 ^e
Mahua cake + T. viride	52.3 ^{bc}	89.6 ^{bc}	136.5 ^{bc}	148.6 ^{bc}	20.15 ^{bc}	60.11 ^d
Nativo WG 75 @ 0.5 %	48.02 ^e	84.12 ^e	135.6 ^e	138.6 ^e	18.39 ^e	63.59 ^{cd}
Mahua cake + Nativo WG 75 @ 0.5 %	49.9 ^{bc}	88.3 ^{bc}	141.2 ^{bc}	147.8 ^{bc}	13.04 ^{bc}	74.18 ^{bc}
T. viride + Nativo WG 75 @ 0.5 %	48.6 ^{cd}	87.88 ^{cd}	138.98 ^{cd}	143.34 ^{cd}	15.02 ^{cd}	70.26 ^c
T. viride: Mahua cake (2:1) + Nativo WG 75 @ 0.5%	55.9 ^a	98.3 ^a	145.2 ^a	160.8 ^a	8.16 ^a	83.84 ^a
Control (uninoculated)	49.02 ^{de}	85.2 ^{de}	137.6 ^{de}	140.6 ^{de}	_	_
Control (Inoculated)	38.3 ^f	79.92 ^f	132.3 ^f	138.6 ^f	50.51 ^{de}	0.00 ^g

^{*}Mean of three replications.

formation and biomass production of *R. solani*. Basal application of *T. viride* + mahua cake and also foliar application of Nativo 0.5% were explored as a means for integrated disease management of leaf blight disease in turmeric crops.

Conflict of Interests

The authors have not declared any conflict of interests.

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