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Full Length Research Paper

Phytochemical and pharmacological studies of *Citharexylum quadrangulare* Jacq. leaves

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Chromatographic investigation of the *Citharexylum quadrangulare* leaves led to isolation of nine compounds viz., stigmasterol (1), β -sitosterol (2), oleanolic acid (3), duranterectoside B (4), durantoside I (5), cirsimaritin 4'-O- β -D-glucopyranoside (6), 5-deoxypulchelloside (7), lamiide (8) and cirsimaritin 4'-O- β -D-glucopyranoside 4"-sodium sulphate (9). The compounds (1, 3 and 4) were isolated for the first time from genus *Citharexylum*. The structures of the isolated compounds were determined by interpretation of their spectroscopical data and comparison with published literature. The aqueous fraction exhibited the most significant anti-inflammatory activity. Its effect was more potent than the reference drug. While, the most significant anti-pyretic sample was the total methanol extract. Also, it showed a gastroprotective activity with preventive index (78%). Finally, the ethyl acetate fraction demonstrated a significant decrease in blood glucose level on hyperglycemic alloxaned rats.

Key words: Citharexylum quadrangulare, Verbenaceae, anti-inflammatory, anti-pyretic, gastroprotective, anti-diabetic.

INTRODUCTION

Verbenaceae is a large family, which contanis 100 genera and around 3000 species (Datta, 1988; Dahiya, 1979; Datta, 1970). It is considered as a potential source of natural products particularly, flavonoids, essential oils (Rizk and Al-Nowaihi, 1989; Rizk, 1986), iridoids, anthocyanins, quinones and caffeic acid derivatives, while alkaloids are rare (Kenner and Requena, 1996; Evans, 1996; Hall, 1976). One of these genera is *Citharexylum*. It includes 115 species and is distributed in South Florida, Guyana, Suriname and Venezuela (Wagner et al., 1999; Datta, 1988; Dahiya, 1979; Datta, 1970). One of these plants is *Citharexylum quadrangulare*

Jacq., which is known as fiddlewood (Wagner et al., 1999). It has a synonym; *C. spinosum* L. (Bedevian, 1994). It was reported in the folkloric medicine as antiarthritic, anti-pyretic, diuretic and liver disorders (Wagner and Wolf, 1977). The previous phytochemical study of *C. quadrangulare* resulted in the isolation of flavonoids viz., cirsiliol 4'-O- β -D-galactopyranoside, cirsimaritin 4'-O- β -Dgluco-pyranoside and cirsimaritin 4'-O- β -Dglucopyranoside 4"-sodium sulfate (Shalaby and Bahgat, 2003). Further investigation on *C. spinosum* and *C. quadrangulare* led to isolation of iridoids such as lamiidoside, lamiide, duranterectoside C, durantoside I,

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Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> License 4.0 International License citharone, phlomiol, 5-deoxypulchelloside I (Balazs et al., 2006; Shalaby and Bahgat, 2003; Khalifa et al., 2002).

The available biological literature showed that C. guadrangulare was evaluated for many activities as gastrointestinal antihypertensive, tract disorders, immunomodulatory, hepatoprotective and anti Schistosoma mansoni cercariae (Bahgat et al., 2005; Shalaby and Bahgat, 2003; Khalifa et al., 2002). On these bases and computer survey employing different data bases including Scifinder, few literature have been reported. This provoked us to carry out further phytochemical and pharmacological investigations on this plant.

METHODOLOGY

General

The NMR spectra were measured using a Jeol JNM-LA 400 and 600 MHz. FT NMR spectrometer (Japan), using tetramethylsilane (TMS) as internal standard and chemical shifts were expressed in δ ppm. Column chromatography (CC) was performed by using Normal Phase-Silica gel (NP-Silica, 70-230 mesh, FLUCA, Germany), Sephadex LH-20 (GE Health Care, Sweden) and Diaion HP-20 (Mitsubishi Chemical Corp., Japan). Fractions were monitored by using thin layer chromatography (TLC) Silica gel 60 precoated plates F₂₅₄ (Merck, Germany) and spots were visualized by heating Silica gel plates and sprayed with 10% H₂SO₄ in MeOH (v/v). The TLC plates were allowed to dry at room temperature and then heated at 110°C till the colors develop and reach their maxima (Stahl, 1970).

Plant material

The leaves of *C. quadrangulare* Jacq. were collected (June 2007) from El-Orman Garden (Giza, Egypt). It was identified by Agricultural Engineer/Trease Labeeb (El-Orman Garden). A voucher specimen has been deposited at Pharmacognosy Department, Faculty of Pharmacy, Minia University under the registration number (Mn-Ph-Cog-005).

Preliminary phytochemical screening

The air-dried powdered leaves of *C. quadrangulare* Jacq. were macerated in 70% methanol (MeOH). The total methanolic extract of the leaves (TMEL) was subjected to preliminary phytochemical screening for its constituents (Sofowora, 1993; Trease and Evans, 1985; Harborne, 1973; Schmidt, 1964; Clause, 1961).

Extraction and isolation

The powdered leaves (1.8 kg) were exhaustively extracted with 70% MeOH (3x, 5 L each) and yielded (250.0 g of TMEL). Physical partitioning of TMEL with petroleum ether and water, followed by chloroform (CHCl₃) and water and finally ethyl acetate (EtOAc) and water, was performed and yielded four main fractions viz., petroleum ether (30.0 g), CHCl₃ (29.6 g), EtOAc (45.8 g) and aqueous (99.6 g).

The petroleum ether fraction was fractionated on NP-Silica column (Φ =7, *L*=200 cm, 900 g), employing gradient technique, increasing polarity from petroleum ether to EtOAc and 42 fractions

were collected (500 ml each). The similar fractions were pooled together. Sedimentation from fraction 18 (0.5 g, eluted by petroleum ether-EtOAc, 80:20) was a mixture of two compounds (1 and 2), which was further crystallized by MeOH (78 mg, colorless needles, R_f=0.59, system; petroleum ether-EtOAc, 7:3). Fractions 19-23 (2.6 g, eluted by petroleum ether-EtOAc, 80:20) was further purified on NP-Silica gel for column (Φ =2, *L*=100 cm, 100.0 g), employing gradient technique with increasing polarity from petroleum ether to EtOAc and 40 fractions were collected (10 ml each). Compound **3** (11.8 mg, white amorphous powder, R_f=0.45, system; petroleum ether-EtOAc, 7:3) was eluted in fractions (8-12) by (petroleum ether-EtOAc, 65:35).

The EtOAc fraction was purified on NP-Silica gel column (ϕ =10, *L*=200 cm, 1.4 kg), employing gradient technique, increasing polarity from CHCl₃ to MeOH and 36 fractions were collected (500 ml each). The similar fractions were collected together.

Fractions 17-20 (5.94 g, eluted by CHCl₃-MeOH, 80:20) were purified on Diaion HP-20 (ϕ =4, *L*=100 cm) and eluted with gradient elution from H₂O to MeOH (H₂O, 20% MeOH, 50% MeOH, 80% MeOH and MeOH, 1.5 L each). The 80% MeOH fraction (3.3 g) was left for 2 h, then filtered. The filtrate was concentrated under reduced pressure to yield (0.7 g). It was purified on NP-Silica (ϕ =2, *L*=80 cm, 28.0 g) employing gradient elution from CHCl₃ to MeOH and 120 fractions were collected (10 ml each). The similar fractions were pooled together. The fractions 39-42 (350 mg, eluted by CHCl₃-MeOH, 95:5) were further purified on Sephadex LH-20 (ϕ =2, *L*=60 cm), eluted with MeOH and led to a mixture of two compounds **4** and **5** (141 mg, faint yellow residue, R₁=0.34, system; CHCl₃-MeOH, 90:10). Compound **6** (90.3 mg, yellow amorphous powder, R₁=0.76, system; CHCl₃-MeOH-H₂O, 15:6:1) was precipitated from the MeOH fraction (1.4 g).

Fractions 21-24 (8.5 g, eluted by CHCl₃-MeOH, 75:25) were fractionated on Diaion HP-20 column (ϕ =4, L=150 cm) using gradient elution with H2O-MeOH (H2O, 20% MeOH, 50% MeOH, 80% MeOH and MeOH, 1.5 L each). The 20% MeOH fraction (2.4 g) was purified on NP-Silica (ϕ =2, L=80 cm, 100.0 g), using gradient elution from CHCl₃ to MeOH and 70 fractions were collected (10 ml, each). The fraction 18 (410 mg, eluted by CHCl₃-MeOH, 80:20) was further purified on Sephadex LH-20 column $(\Phi=2, L=80 \text{ cm})$, eluted with MeOH and led to compound 7 (90.9 mg, faint yellow residue, R_f=0.48, system; CHCl₃-MeOH-H₂O, 80:20:1). Moreover, fraction 26 (380 mg, eluted by CHCl₃-MeOH, 70:30) was further purified on Sephadex LH-20 column (ϕ =2, L=60 cm), eluted with MeOH and led to compound 8 (47.6 mg, faint vellow residue, R_f=0.31, system; CHCl₃-MeOH-H₂O, 80:20:1). Finally, 50% MeOH fraction (3.2 g) was purified on NP-Silica column (ϕ =2, L=150 cm, 130.0 g), isocratically eluted with CHCl₃-MeOH-H₂O (80:20:1) and 200 fractions were collected (10 ml each). The fractions (105-144, 130 mg) were purified on Sephadex LH-20 column (ϕ =2, L=60 cm), eluted with MeOH and led to compound 9 (3 mg, yellow amorphous powder, R_f=0.42, system; CHCl₃-MeOH, 80:20).

Animals

Adult male Sprague-Dawley albino rats (220-250 g) were used. They were purchased from the animal house, Faculty of Agriculture, Minia University. The animals were kept under identical environmental circumstances, fed with standard nutrition and water *ad libitum* and left to adapt to the environment for at least 7 days prior to the experiments at 22±2 °C under a 12/12 h light/dark cycle. They were handled only at the time of experiments and during cage washing. All conditions were ensured to reduce animal distress. The care and procedures involving animals were conducted in conformity with the institutional guidelines of the Pharmacology and Toxicology Department, Faculty of Pharmacy, Minia University and in agreement with the provisions of the Declaration of laboratory

No.	Compounds; δ_{H} (Integration, Multiplicity, J in Hz)							
NO.	(1)	(2)	(3)	(4)				
1				5.77 (1H, br.s)				
3	3.54 (1H, m)	3.54 (1H, m) 3	.20 (1H, br.d, 10.4)	7.42 (1H, s)				
4	2.24 (2H, m)	2.24 (2H, m)						
6	5.34 (1H, m)	5.34 (1H, m)		2.29 (2H, m)				
9				2.81 (1H, s)				
10				1.10 (3H, s)				
12			5.20 (1H, br.s)					
18	0.68 (3H, s)	0.68 (3H, s)						
19	0.99 (3H, s)	0.99 (3H, s)						
22/23	4.98,5.71 (2H, m	n)						
23			1.10 (3H, s)					
24			0.91 (3H, s)					
25			0.88 (3H, s)					
26			0.76 (3H, s)					
27			0.97 (3H, s)					
29			0.73 (3H, s)					
30			0.90 (3H, s)					
Me gps	0.77-0.97 (m)	0.77-0.97 (m)						
COOMe				3.70 (3H, s)				
1'				4.61 (1H, d, 7.8)				
7"				7.00 (1H, d, 12.4				
8"				6.07 (1H, d, 12.4				

Table 1. Important signals in ¹H-NMR spectral data of compounds 1-4.

Compounds 1-3 were measured in CDCI3: compound 4 was measured in MeOD. All compounds were measured by using 400 MHz.

animals of the National Institutes of Health (NIH publication No. 85-23, revised 1985).

Determination of acute toxicity (LD₅₀)

The acute toxicity of the TMEL of C. quadrangulare was determined by the following experimental model described by Schapoval et al. (1998). This was carried out by measuring the lethal dose for 50% of the laboratory animals (LD₅₀ method). Different single oral doses (0.5, 1, 2, 4 and 8 g/kg, per oral) of TMEL of the plant [suspended in 0.5% carboxymethylcellulose solution (CMC)] were administered to 6 groups (5 animals each) of rats (240±10 g). The control group received an equivalent dose of TMEL vehicle (0.5% CMC solution, 10 ml/kg, per oral). Both the test and control groups were observed for 48 h under normal environmental conditions, with free access to food and water.

Pharmacological activities

All groups were orally given the tested samples by using gavage. In all experiments, rats were randomly divided into 7 groups (5 animals each) as follows:

1- The 1st group (negative control) non-treated one, administered 10 ml/kg (0.5% CMC).

2- The 2nd group (positive control) treated with the standard drugs (indomethacin or acetylsalicylic acid or ranitidine or glibenclamide).

- 3- The 3rd group treated with TMEL.
- 4- The 4th group treated with petroleum ether fraction.

- 5- The 5th group treated with period an and a final field of the second seco

The groups from 3rd to 7th were orally given doses of 100 mg/kg (the extract and the different fractions were suspended in 0.5% CMC).

Anti-inflammatory activity

The TMEL and its fractions were evaluated by the carrageenaninduced paw edema method in the rats (Winter et al., 1962). The tested samples were suspended in 0.5% CMC solution and administered orally to the rats (230±10 g), one hour prior to the subcutaneous injection of 0.1 ml carrageenan suspension (1% w/v in normal saline solution) into the sub-plantar area of the right hind paw (Vogel and Vogel, 1997). The 2nd group was given indomethacin at a dose of 8 mg/kg (Sawadogo et al., 2006; Petrovic et al., 2003). Edema measurements were determined in mm with a plethysmometer (Ferreira, 1979) after 0, 1, 2, 3 and 4 h carrageenan injection. The results are listed in (Table 5).

Anti-pyretic activity

Anti-pyretic effect was evaluated by employing yeast-induced fever

N	Compounds; δ_{H} (Integration, Multiplicity, J in Hz)								
No.	(5)	(6)	(7)	(8)	(9)				
1	5.80 (1H, br.s)		5.48 (1H, d, 2.3)	5.81(1H, s)					
3	7.35 (1H, s)	6.80 (1H, s)	7.38 (1H, s)	7.43 (1H, s)	6.94 (1H, s)				
5		12.78 (1H, s)	2.87 (1H, d, 9.2)		12.60 (1H, s)				
6	2.29 (2H, m)		2.26 (1H, m)	2.37 (1H, dd, 15.1, 5.2) 2.25 (1H, dd, 15.1, 3.4)					
OMe-6		3.72 (3H, s)			3.73 (3H, s)				
7			3.49 (1H, m)	3.52 (1H, m)					
OMe-7		3.87 (3H, s)			3.93 (3H, s)				
8		6.81 (1H, s)	2.26 (1H, m)		6.98 (1H, s)				
9	2.92 (1H, s)		2.80 (1H, m)	2.78 (1H, s)					
10	1.13 (3H, s)		1.05 (3H, d, 7.3)	1.00 (3H, s)					
COOMe	3.70 (3H, s)		3.67 (3H, s)	3.73 (3H, s)					
1'	4.61 (1H, d, 7.8)		4.55 (1H, d, 7.9)	4.59 (1H, d, 7.9)					
2',6'		7.99 (2H, d, 8.6)			8.08 (1H, d, 8.9)				
3',5'		7.19 (2H, d, 8.6)			7.21 (1H, d, 8.9)				
1''		5.06 (1H, d, 7.0)			5.10 (1H, d, 7.7)				
7"	7.72 (1H, d, 16.1)								
8"	6.55 (1H, d, 16.1)								

 Table 2. Important ¹H-NMR spectral data of compounds (5-9).

Compounds (5, 7 and 8) were measured in MeOD. Compounds (6 and 9) were measured in DMSO-*d*₆. All compounds were measured by using 400 MHz, except compound 8 was measured by using 600 MHz.

according to Teotino et al. (1963). It was performed on rats (225±5 g) by subcutaneous injection (in the back, below the nape of the neck) of 20% aqueous suspension of yeast in a dose of 10 ml/kg to induce pyrexia (Kang et al., 2008). The rectal temperature of each animal was recorded, using digital thermometer, which was inserted 2 cm into the rectum, before and 18 h after the yeast injection, when the temperature was at the peak (Panthong et al., 2003). The animals that did not show a minimum increase of 0.5 °C in temperature after 18 h were discarded. The rectal temperature of each animal was recorded again at 30 min interval for 3 h following the drug administration. The positive control group was given acetylsalicylic acid (100 mg/kg) (Gege-Adebayo et al., 2013). The results are displayed in (Table 6).

Gastroprotective activity

The procedure of gastroprotective activity was started by feeding the rats (235±10 g) with a standard diet of commercial rat chow and tap water. They were left to acclimatize to the environment for at least one week prior the experiment. Rats fasted for 24 h prior to the experiment in mesh-bottomed cages to decrease coprophagia, but the animals had free access to water except for the last hour before the experiment (Inas et al., 2011). All the experiments were performed during the same time of the day to avoid variations due to diurnal rhythms of putative regulators of gastric functions (Garrik et al., 1986).

All these treatments were given one hour before the induction of gastric ulceration by a large oral dose of indomethacin (40 mg/kg) (Choudhary et al., 2014; Raji et al., 2011; Mishra et al., 2009).

The reference drug (ranitidine, 50 mg/kg) was administered (Choudhary et al., 2014). After one hour of administration of the drugs, the rats were sacrificed by cervical dislocation. Their

stomachs were removed and opened along the greater curvature and then washed with serum physiological solution and macroscopic gross mucosal lesion were counted and scored to determine the severity of these lesions (Inas et al., 2011). The protective effect of TMEL and its fractions were compared with indomethacin and ranitidine groups.

The assessment of gastric mucosal lesions in each stomach was measured. Moreover, the ulcer score for each stomach was expressed as the total length of gastric lesions in that stomach. Afterwards, the mean ulcer score for each group was calculated. The preventive index (P.I.) of a given drug was calculated from the following equation (Hano et al., 1976). The results are shown in Table 7.

$$P.I. = \frac{(U.I. \text{ of ulcerated group}) - (U.I. \text{ of treated group})}{(U.I. \text{ of ulcerated group})} \times 100$$

Where, U.I. is the ulcer index.

Anti-diabetic activity

The rats weighing 230±10 g were allowed to fast for 24 h prior to experiment and diabetic rats were obtained by the administration of a single dose of intraperitoneal injection of alloxan 120 mg/kg body weight (Vogel and Gang, 2002). After 72 h of alloxan injection, diabetes was confirmed by testing blood sugar level by using Accu-Chek[®] Glucometer to monitor the blood sample from the tail vein. When, blood glucose level (BGL) above 200 mg/dl was considered diabetic. The 2nd group treated with standard drug (glibenclamide, 0.5 mg/kg) (Hazra et al., 2011). The blood samples were taken and measured. They were taken at zero, 1, 2, 3 and 4 h following drug

Na	Compou	inds (δ _c , Multij	olicity)	Na	Compounds (δ_c , Multiplicity)			
No. —	(1)	(2)	(3)	No.	(1)	(2)	(3)	
1	37.3,t	37.3,t	38.5,t	16	28.8,t	28.2,t	23.0 ^d ,t	
2	31.7 ^a ,t	31.7 ^a ,t	27.2 ^a ,t	17	56.1 ^b ,d	56.0 ^b ,t	46.5,s	
3	71.8,d	71.8,d	79.0,d	18	12.0 ^c ,q	12.0 ^c ,q	41.0,d	
4	42.4,t	42.4,t	38.8 ^b ,s	19	19.4 ^d ,q	19.4 ^d ,q	45.9,t	
5	140.8,s	140.8,s	55.3,d	20	40.4,d	36.1,d	30.7,s	
6	121.7,d	121.7,d	18.3,t	21	21.2,q	18.7,q	33.8,t	
7	31.9 ^a ,t	31.9 ^a ,t	32.7 ^c ,t	22	138.2,d	34.0,t	32.5 [°] ,t	
8	31.9 ^ª ,d	31.9 ^a ,d	39.3 ^b ,s	23	129.3,d	26.2,t	28.1,q	
9	50.2,d	50.2,d	47.7,d	24	51.2,d	45.9,d	15.5,q	
10	36.5,s	36.5,s	37.1,s	25	31.9,d	29.2,d	15.3,q	
11	21.1,t	21.1,t	23.4 ^d ,t	26	19.0 ^d ,q	19.8 ^d ,q	17.2,q	
12	39.7,t	39.8,t	122.7,d	27	21.2,q	19.1 ^d ,q	25.9,q	
13	42.4,s	42.4,s	143.6,s	28	25.3,t	23.1,t	183.1,s	
14	56.9 ^b ,d	56.8 ^b ,d	41.6,s	29	12.0 ^c ,q	12.0 ^c ,q	33.1,q	
15	24.4,t	24.3,t	27.7 ^a ,t	30			23.6,q	

 Table 3.
 ¹³C-NMR spectral data of compounds (1-3).

^{a, b, c, d}values may be interchangeable within the same compound (CDCl₃, 100 MHz).

treatment. The results are demonstrated in (Table 8).

Statistical analyses

The statistical analyses of the obtained results were done using GraphPad Prism 5 (Graphpad Software, San Diego California, USA). The obtained outcomes were expressed in terms of mean \pm SEM. Differences between the mean values for individual groups were assessed by one-way analysis of variance. In all analyses, (**P*<0.05) or (***P*<0.01) or (***P*<0.001) were taken to indicate statistical significance.

RESULTS AND DISCUSSION

The preliminary phytochemical screening of *C. quadrangulare* TMEL showed the presence of various metabolites such as carbohydrates and/or glycosides, flavonoids, unsaturated sterols and/or triterpenes, saponins and tannins. While, it was free from many constituents such as crystalline sublimate substances, volatile oils, alkaloids and/or nitrogenous substances, cardenolides and anthraquinones.

Furthermore, the petroleum ether and EtOAc fractions of TMEL were purified by employing diversity of chromatographic techniques; including open columns of Silica gel, Diaion HP-20 and Sephadex LH-20 to afford nine compounds (Figure 1) viz., compounds (1-3) from petroleum ether fraction and compounds (4-9) from EtOAc fraction. They belong to different classes; two steroloidal aglycones: stigmasterol (1) (Maima et al., 2008) and β -sitosterol (2) (Maima et al., 2008); triterpenes: oleanolic acid (3) (Chang et al., 2009); four iridoids: duranterectoside B (4) (Takeda et al., 1995), durantoside I (5) (Takeda et al., 1995), 5deoxypulchelloside (7) (Shalaby and Bahgat, 2003; Khalifa et al., 2002) and lamiide (8) (Yalcin et al., 2007; Khalifa et al., 2002) and finally two flavonoids: cirsimaritin 4'-O- β -D-glucopyranoside (6) (Shalaby and Bahgat, 2003) and cirsimaritin 4'-O-B-D-glucopyranoside 4"sodium sulphate (9) (Shalaby and Bahgat, 2003). The structures of the isolated compounds were elucidated by comparing spectroscopical data (Tables 1 to 4) with the reported literature and also comparing their physical properties with the authentic samples. Three of them (compounds 1, 3 and 4) were isolated for the first time from genus Citharexylum. This is very important in the chemotaxonomical study of the plant.

The TMEL of *C. quadrangulare* was evaluated against the lethality effect on rats up to 8 g/kg (exceeded ten times of the therapeutic dose 100 mg/kg) during the first 48 h. Moreover, no toxic manifestation has been observed such as paw-licking, stretching, respiratory distress, diarrhea (Arthur et al., 2011). Thus, *C. quadrangulare* has a wide margin of safety.

TMEL and its fractions exhibited anti-inflammatory activities due to its inhibition of the carrageenan induced edema. The significant decrease of paw edema has been obtained with the aqueous and petroleum ether fractions, after 4 h, comparing with indomethacin as positive control (Table 5). The phytochemical investigation of *C. quadrangulare* indicated that it has considerable contents of sterols, triterpenes and flavonoids. Most of these compounds are responsible for the anti-inflammatory effects (Lalitha and Gayathiri, 2013; Perez, 2001). Moreover, the previous studies indicated that flavonoids such as rutin, quercetin, luteolin, biflavonoids, steroids

N.,	Compounds (δ _c , Multiplicity)					N.			Compounds	(δ _c , Multiplicity)			
No. ((4) ¹	(5)1	(6) ²	(7) ¹	(8) ¹	(9) ²	No.	(4) ¹	(5) ¹	(6) ²	(7) ¹	(8) ¹	(9) ²
1	94.0,d	94.1,d		96.0,d	94.5,d		5'	77.3,d	77.3,d	116.6,d	78.1 ^ь ,d	78.4,d	116.6,d
2			163.3,s			163.3,s	6'	62.7,t	62.7,t	128.2,d	62.7,t	62.8,d	128.1,d
3	152.2,d	152.2,d	103.6,d	153.8,d	152.5,d	103.6,d	1"	136.2,s	135.7,s	100.0,d			99.2,d
4	115.5,s	115.5,s	182.1,s	111.0,s	115.4,s	182.2,s	2"			73.2,d			73.2,d
5	69.0,s	68.9,s	152.0ª,s	38.7ª,d	69.2,s	152.0ª,s	2",6"	129.1,d	129.2,d				
6	45.3,t	45.6,t	132.0,s	77.5 ^b ,d	46.7,t	131.9,s	3"			76.6 ^b ,d			75.0⁵,d
7	80.4,d	80.7,d	158.6,s	79.9,d	77.8ª,d	158.7,s	3",5"	131.4 ^ь ,d	131.0 ^ь ,d				
8	78.7,s	78.7,s	91.4,d	39.0ª,d	79.1,s	91.7,s	4"	130.1 ^b ,d	130.0 ^b ,d	69.8,d			75.1⁵,d
9	58.1,d	58.3,d	152.5ª,s	40.2,d	58.1,d	152.6ª,d	5"			77.2 ^b ,d			75.3⁵,d
10	21.3,q	21.3,q	105.1, s	13.9,q	21.3,q	105.1,s	6"			60.7,t			60.7,t
11	167.9ª,s	167.9ª,s		169.1,s	168.0,s		7"	144.7,d	146.4,d				
							8"	120.6,d	119.1,d				
1'	99.6,d	99.6,d	124.0,s	99.7,d	99.6,d	123.9,s	9"	167.1ª,s	168.1ª,s				
2'	74.4,d	74.4,d	128.1,d	74.5,d	74.4,d)	128.1,d	OMe-6			60.0,q			60.0,q
3'	78.2,d	78.2,d	116.6,d	77.8 ^b ,d	77.4ª,d	116.6,d	OMe-7			56.3,q			56.4,q
4'	71.6,d	71.6,d	160.3,s	71.5,d	71.7,d	160.2,s	COOMe	51.8,q	51.8,q		51.8,q	51.7,q	

 Table 4.¹³C-NMR spectral data of compounds (4-9).

^{a, b} values may be interchangeable within the same compound. ¹(MeOD) and ²(DMSO-*d*₆). All compounds were measured by using 100 MHz, except compound (**8**) was measured by using 150 MHz NMR.

Group	Thickness of the paw (mm)/h								
Group	0	1	2	3	4				
Control	5.55±0.12	5.87±0.17	5.03±0.23	4.87±0.10	4.90±0.12				
Indomethacin	5.19±0.17	4.32±0.10***	4.33±0.15*	3.99±0.11**	3.94±0.14**				
TMEL	5.42±0.19	4.91±0.26**	4.29±0.24*	4.29±0.23*	3.90±0.23**				
Petroleum ether fr.	5.18±0.23	4.70±0.22***	4.42±0.25	4.16±0.95**	3.80±0.18***				
CHCl₃ fr.	5.30±0.28	4.60±0.25***	4.23±0.20*	4.24±0.16*	3.90±0.16**				
EtOAc fr.	5.39±0.23	4.80±0.10**	4.41±0.21	4.02±0.16**	4.23±0.16*				
Aqueous fr.	4.95±0.22	4.60±0.14***	4.04±0.18*	3.80±0.18***	3.66±0.20***				

Values represent Mean±SEM, (n=5). Significant difference (*P<0.05, **P<0.01 and ***P<0.001). Indomethacin (8 mg/kg) and the other drugs (100 mg/kg).

and triterpenoids produced significant antiinflammatory activities (Silva et al., 2005). The obtained anti-inflammatory results are in line with other previous studies in family Verbenaceae (Shukla et al., 2011; Amir et al., 2011; Krishnaraju et al., 2009; Monteiro et al., 2007; Penido et al.,

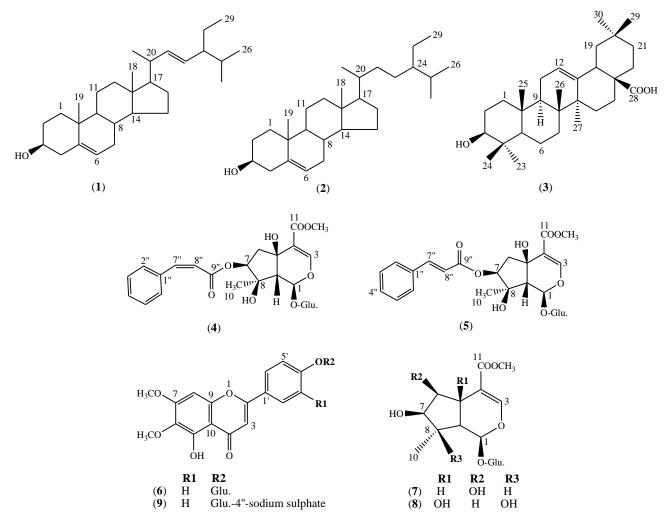


Figure 1. Structures of the isolated compounds 1-9.

2006; Abena et al., 2003).

The anti-pyretic activity of TMEL and its fractions have been evaluated by causing reduction in yeast-induced pyrexia (Table 6). The rectal temperature reached normal level after 1.5, 2, 1, 3 and 3 h with TMEL, petroleum ether, CHCl₃, EtOAc and aqueous fractions, respectively. The CHCl₃ fraction showed a weak anti-pyretic effect, based on decreasing the rectal temperature without reaching the normal level after 3 h. Aforementioned information about the chemical profile of the C. quadrangulare showed the presence of the major constituents as sterols, triterpenes and flavonoids, can explain the anti-pyretic activity (Hossain et al., 2011; Boakye-Gyasi et al., 2011; Achuta et al., 2011). This deduction is concentrated in TMEL. Our findings are in line with previous studies in family Verbenaceae (Rohit et al., 2012; Shukla et al., 2011; Abena et al., 2003).

Table 7 shows that the TMEL has a significant gastroprotective effect, while the petroleum ether and $CHCl_3$ fractions have a moderate gastro-protective

activity. On the other hand, the EtOAc and aqueous fractions displayed a weak gastro-protective activity. The preliminary phytochemical investigation of TMEL of C. a positive quadrangulare exhi-bited result with Liebermann-Burchard's test (Schmidt, 1964) indicating the presence of sterols. Moreover, from petroleum ether fraction, sterols and triterpenes compounds were isolated; hence gastroprotective activity of these fractions may be due to presence of terpenoidal or steroidal compounds (Tovey et al., 2011; Subhadhirasakul and Pechpongs, 2005). The aforementioned gastroprotective results are in line with other previous studies in family Verbenaceae (Tajik et al., 2015; Chellappan and Pemiah 2014; Jothi et al., 2010; Monteiro et al., 2007; Penido et al., 2006).

The anti-diabetic activity was concentrated in the EtOAc fraction. This clearly appeared in decreasing the blood glucose level, while the TMEL and $CHCl_3$ fraction have moderate effects. But, the petroleum ether fraction increases the blood glucose level (13%). The previous

Crown	Rectal temperature (°C)/min									
Group	Pre-yeast	Pre-drug	30	60	90	120	150	180		
Control	36.67±0.23	38.03±0.33	37.47±0.53	37.37±0.23	37.90±0.20	37.60±0.70	38.33±0.17	37.83±0.07		
Acetylsalicylic acid	35.00±0.39	38.40±0.49	37.13±0.38	37.57±0.28	35.47±0.23***	35.40±0.15**	35.33±0.24***	35.30±0.32***		
TMEL	35.83±0.13	38.40±0.15	36.53±0.12	35.60±0.17***	35.80±0.10***	36.03±0.24*	35.90±0.10***	35.80±0.17***		
Petroleum ether fr.	36.00±0.28	38.47±0.28	37.10±0.10	36.20±0.15**	35.97±0.18***	36.00±0.40*	36.27±0.20***	36.03±0.15***		
CHCl₃ fr.	35.67±0.36	38.17±0.37	37.80±0.51	36.50±0.23*	36.33±0.29***	36.20±0.36*	36.03±0.41***	36.27±0.49**		
EtOAc fr.	35.87±0.29	38.13±0.29	37.23±0.12	36.13±0.24**	36.17±0.12***	36.33±0.29*	36.40±0.23***	35.97±0.35***		
Aqueous fr.	35.90±0.40	39.13±0.47	37.30±0.50	36.20±0.46**	36.57±0.35***	36.07±0.52*	36.07±0.33***	35.90±0.36***		

Table 6. Anti-pyretic activities of C. quadrangulare.

Values represent Mean±SEM, (n=5). Significant difference (*P<0.05, **P<0.01 and ***P<0.001). All drugs (100 mg/kg).

Table 7. Gastroprotective activities of C. quadrangulare.

Group	Mean ulcer score (mm)	P.I. (%)
Control	25.0±8.38	
Ranitidine	3.0±1.08	88
TMEL	5.5±1.85	78
Petroleum ether fr.	8.5±1.55	66
CHCl₃ fr.	9.5±4.66	62
EtOAc fr.	15.8±5.66	38
Aqueous fr.	22.5±2.66	10

Values represent Mean \pm SEM, (n=5). Ranitidine (50 mg/kg) and the other drugs (100 mg/kg).

studies demonstrated that various flavonoids especially quercetin possesses anti-diabetic activities (Vessal et al., 2003; Hif and Howell, 1985). Therefore, the highest anti-diabetic activity, which was shown by the EtOAc (58%) fraction could be attributed to its content of flavonoids. All results are listed in Table 8. Our findings are in line with previous studies in family Verbenaceae (Rohit et al., 2012; Zanatta et al., 2007; Villasenor and Lamadrid, 2006).

Conclusion

Investigation of TMEL of *C. quadrangulare* afforded three compounds, which were reported for the first time in the genus in addition to six known compounds. The TMEL and its fractions

showed important pharmacological activities such as anti-inflammatory, anti-pyretic, gastroprotective and anti-diabetic. Therefore, it could have a supportive role in the pharmaceutical field towards the development of new drugs.

Conflict of interests

The authors have not declared any conflict of

Table 8. Anti-diabetic activities of C. quadrangulare.

0		Blood g	glucose level (r	ng/dl)/h	
Group	0	1	2	3	4
Control	205±1.0	205±0.7	201±5.0	201±5.3	199±7.3
%	100.00	100.00	98.04	98.04	97.07
Glibenclamide	231±5.6	220±15.0	195±13.6	179±12.3	171±11.4**
%	100.00	95.23	84.41	77.49	74.03
TMEL	221±13.3	256±13.9	359±20.6	316±20.8	187±12.3*
%	100.00	115.84	162.44	142.99	84.62
Petroleum ether fr.	414±30.0	541±4.7	508±7.7	485±25.4	470±20.9
%	100.00	130.68	122.71	117.15	113.53
CHCl₃ fr.	235±10.7	304±17.2	295±9.1	257±11.7	203±13.2*
%	100.00	129.36	125.53	109.36	86.38
EtOAc fr.	512±6.6	546±15.7	426±22.3	333±19.0	219±9.7***
%	100.00	106.64	83.20	65.04	42.77
Aqueous fr.	209±4.1	219±8.0	222±8.7	205±6.3	202±6.1
%	100.00	100.00	106.22	98.09	96.65

Values represent Mean±SEM, (n=5). Significant difference (**P*<0.05, ***P*<0.01 and ****P*<0.001). Glibenclamide (0.5 mg/kg) and the other drugs (100 mg/kg).

interest.

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REFERENCES

- Abena AA, Diatewa M, Gakosso G, Gbeassor M, Hondi-Assah T, Ouamba JM (2003). Analgesic, antipyretic and anti-inflammatory effects of essential oil of *Lippia multiflora*. Fitoterapia 74(3):231-236.
- Achuta VN, Mannangati V, Panda S, Usha Rani E, Prasanthi P (2011). Analgesic, antipyretic and anti-inflammatory studies on methanolic extract of *Jasminum trichotonum* leaves. Int. J. Res. Ayurv. Pharm. 2(2):637-639.
- Amir F, Yam WS, Chin KY (2011). Chemical constituents and biological applications of *Lippia nodiflora*. Arch. Pharm. Pract. 2:101-105.
- Arthur FKN, Woode E, Terlabi EO, Larbie C (2011). Evaluation of acute and subchronic toxicity of *Annona muricata* (Linn.) aqueous extract in animals. Eur. J. Exp. Biol. 1(4):115-124.
- Bahgat M, Shalaby NMM, Ruppel A, Maghraby AS (2005). Humoral and cellular immune responses induced in mice by purified iridoid mixture that inhibits penetration of *Schistosoma mansoni* cercariae upon topical treatment of mice tails. J. Egypt. Soc. Parasitol. 35:597-613.
- Balazs B, Toth G, Duddeck H, Soliman, Hesham SM (2006). Iridoid and lignan glycosides from *Citharexylum spinosum* L. Nat. Prod. Res. 20:201-205.
- Bedevian AK (1994). Illustrated polyglottic dictionary of plant name, Madbouly library, Cairo.
- Boakye-Gyasi E, George KA, Wonder KMA (2011). Anti-inflammatory, antipyretic and antioxidant properties of a hydroalcoholic leaf extract of *Palisota hirsuta* K. Schum. (Commelinaceae). West Afr. J. Pharm.

22(1):10-18.

- Chang SW, Kim KH, Lee IK, Choi SU, Lee KI (2009). Phytochemical constituents of *Geranium eriostemon*. Nat. Prod. Sci. 15:151-155.
- Chellappan DR, Pemiah B (2014). Pharmacognostical, phytochemical and *in vivo* gastro-protective investigation of *Gmelina arborea*. Int. J. Pharm. Pharm. Sci. 6(4):153-157.
- Choudhary M, Kumar V, Singh S (2014). Gastric anti-secretory and cytoprotective effects of hydroalcoholic extracts of *Plumeria alba* Linn. leaves in rats. J. Integr. Med. 12(1):42-51.
- Clause EP (1961). Pharmacognosy. Henry Krimpton, London 4th Ed. III. P 121.
- Dahiya BS (1979). Systematic Botany (Taxonomy of Angiosperms), Kalyani publishers, Luohiana, printed in India. pp. 243-247.
- Datta SC (1988). Systematic Botany, New age international (p) limited publishers, New Delhi, Bangalore, Chennai, Guwahati, Hyderabad, Kolkata, Lucknow, Mumbai. pp. 425-428.
- Datta SC (1970). A Hand Book of Systematic Botany, Asia publishing house. 2nd Ed. pp. 310-312.
- Evans WC (1996). Trease and Evan's Pharmacognosy, WB Saunders Company Ltd, London, Philadelphia, Toronto, Sydney, Tokyo. 14th Ed. pp. 47-48.
- Ferreira SH (1979). A new method for measuring variations of rat paw volume. J. Pharm. Pharmacol. 31:648.
- Garrik T, Buack S, Bass P (1986). Gastric motility is a major factor in cold restraint-induced lesion formation in rats. Am. J. Physiol. 250:191-199.
- Gege-Adebayo GI, Bassi AS, Igbokwe VU, Shafe MO (2013). Antipyretic effect of *Ocimum gratissium* on brewer's yeast induced fever in wistar rats. J. Med. Med. Sci. 4(6):247-251.
- Hall MA (1976). Plant Structure, Function and Adaptation, The English language book society and Macmillan press LTD, London and Basingstoke associated companies in Delhi, Hong Kong, Lagos and Singapore P 362.
- Hano J, Bugajaski J, Danek L (1976). Effect of adrenergic blockade on gastric secretion altered by catecholamines in rats. Ther. Exp. (Warsz). 24(4):507-524.
- Harborne JB (1973). Phytochemical methods. Chapman and Hall Ltd., London. pp. 49-188.

- Hazra M, Kundusen S, Battacharya S, Haldar PK, Gupta M, Mazumder UK (2011). Evaluation of hypoglycemic and antihyperglycemic effects of *Luffa cylindrical* fruit extract in rats. J. Adv. Pharm. Educ. Res. 2:138-146.
- Hif CS, Howell SL (1985). Effects of flavonoids on insulin secretion and ⁴⁵Ca⁺² handling in rat islets of langerhans. J. Endocrinol. 107:1-8.
- Hossain E, Mandal SC, Gupta J (2011). Phytochemical screening and *In-vivo* anti-pyretic activity of the methanol leaf-extract of *Bombax malabaricum* DC. (Bombacaceae). Trop. J. Pharm. Res. 10(1):55-60.
- Inas ZAA, Hala AHK, Gehan HH (2011). Gastroprotective effect of *Cordia myxa* L. fruit extract against indomethacin-induced gastric ulceration in rats. Life Sci. J. 8(3):433-445.
- Jothi ET, Karthikeyan R, Suryalakshmi PV, Srinivasababu P (2010). Gastroprotective potential of *Premna serratifolia* Linn. leaves against aspirin induced ulcer in albino rats. Pharmacologyonline 3:189-198.
- Kang JY, Khan MNA, Park NH, Cho JY, Lee MC, Fujii H, Hong YK (2008). Antipyretic, analgesic and anti-inflammatory activities of the seaweed Sargassum fulvellum and Sargassum thunbergii in mice. J. Ethnopharmacol. 116:187-190.
- Kenner D, Requena Y (1996). Botanical Medicine: A European Professional Perspective, Paradigm publications, Brookline, Massachusetts, USA. pp. 116-240.
- Khalifa TI, El-Gindi OD, Ammar HA, El-Naggar DM (2002). Iridoid glycosides from *Citharexylum quadrangular*. Asian J. Chem. 14:197-202.
- Krishnaraju AV, Rao CBM, Sundararaju D, Sengupta K, Trimurtulu G (2009). Anti-Inflammatory Activity of *Vitex leucoxylon* L. Bark Extracts Against Freund's Complete Adjuvant Induced Arthritis in Sprague Dawley Rat. Am. J. Infect. Dis. 5:68-73.
- Lalitha P, Gayathiri P (2013). In vitro anti-inflammatory and phytochemical properties of crude ethyl acetate extract of Baliospermum montanum leaf (Muell.-Arg.). Afr. J. Biotechnol. 12(39):5743-5748.
- Maima AO, Thotthi GN, Ndwigah SN, Kamau FN, Kibwage IO (2008). Phytosterols from the stem bark of *Combretum fragrans* F. Hoffin. East Cent. Afr. J. Pharm. Sci. 11:52-54.
- Mishra A, Arora S, Gupta R, Manvi, Punia RK, Sharma AK (2009). Effect of *Feronia elephantum* (Corr.) fruit pulp extract on indomethacin-induced gastric ulcer in albino rats. Trop. J. Pharm. Res. 8(6):509-514.
- Monteiro MVB, Leite AKR, Bertini LM, Morais SM, Nunes-Pinheiro DCS (2007). Topical anti-inflammatory, gastro-protective and antioxidant effects of the essential oil of *Lippia sidoides* Cham. leaves J. Ethnopharmacol. 111:378-382.
- Panthong A, Kanjanapothi D, Taesotikul T, Wongcome T, Reutrakul V (2003). Anti-inflammatory and antipyretic properties of *Clerdendrum petasites* S. Moore. J. Ethnopharmacol. 85:151-156.
- Penido C, Costa KA, Futuro DO, Paiva SR, Kaplan MA, Figueiredo MR, Henriques MG (2006). Anti-inflammatory and anti-ulcerogenic properties of *Stachytarpheta cayennensis* (L.C. Rich) Vahl. J. Ethnopharmacol. 104(1-2):225-33.
- Perez G (2001). Anti-inflammatory activity of compounds isolated from plants. Sci. World J. 1:713-784.
- Petrovic SD, Dobric S, Bokonjic D, Niketic M, Garcia-Piñeres A, Merfort I (2003). Evaluation of *Tanacetum larvatum* for an anti-inflammatory activity and for the protection against indomethacin-induced ulcerogenesis in rats. J. Ethnopharmacol. 87(1):109-113.
- Raji Y, Oyeyemi WA, Shittu ST, Bolarinwa AF (2011). Gastroprotective effect of methanol extract of *Ficus asperifolia* bark on indomethacininduced gastric ulcer in rats. Niger. J. Physiol. Sci. 26(1):43-48.
- Rizk AM, Al-Nowaishi AS (1989). The Phytochemistry of the Horticultural Plants of Qatar, The Alden press LTD, UK. pp. 198-207.
- Rizk AM (1986). The Phytochemistry of the Flora of Qatar. Kingprint of Richmond, Great Britain. pp. 416-418.
- Rohit K, Vaibhav P, Manodeep C, Jagadish VK (2012). Phytochemical and pharmacological profile of *Gmelina arborea*. Int. Res. J. Pharm. 3:61-64.
- Sawadogo WR, Boly R, Lompo M, Some N, Lamien CE, Guissou IP, Nacoulma OG (2006). Anti-inflammatory, analgesic and anti-pyretic activities of *Dicliptera verticillata*. Int. J. Pharmacol. 2(4):435-438.
- Schapoval EES, Winter de Vargas MR, Chaves CG, Bridi R, Zuanazzi

JA, Henriques AT (1998). Anti-inflammatory and antinociceptive activities of extracts and isolated compounds from *Stachytarpheta cayennensis*. J. Ethnopharmacol. 60:53-59.

- Schmidt J (1964). Organic Chemistry, Oliver and Boyd, Edinburgh and London, 8th Ed. pp. 318-673.
- Silva GN, Martins FR, Matheus ME, Leitao SG, Fernandes PD (2005). Investigation of anti-inflammatory and antinociceptive activities of *Lantana trifolia*. J. Ethnopharmacol. 100:254-259.
- Shalaby NMM, Bahgat M (2003). Phytochemical and some biological studies of *Citharexylum quadrangular* Jacq. Chem. Nat. Microb. Prod. 4:219-228.
- Shukla P, Mishra SB, Gopalakrishna B (2011). Screening of antiinflamatory and antipyretic activity of *Vitex leucoxylon* Linn. Indian J. Pharmacol. 42:409-411.
- Sofowora A (1993). Medicinal plants and traditional medicines in Africa spectrum book Ltd. Ibadan, Nigeria. P 289.
- Stahl E (1970). Thin Layer Chromatography, 2nd Ed. Springer Verlag, Berlin, Heidelberg, New York. pp. 750-810.
- Subhadhirasakul S, Pechpongs P (2005). A terpenoid and two steroids from the flowers of *Mammea siamensis*. Songklanakarin J. Sci. Technol. 27(2):555-561.
- Tajik J, Kheirandish R, Amanollahi R, Shahabi A (2015). Gastroprotective effect of aqueous extracts of *Lippia citriodora*, ajowan (*Trachyspermum copticum*), and *Dracocepalum polychaetum* on induced gastric ulcer in rats. Comp. Clin. Pathol. 24(6):1605-1610.
- Takeda Y, Morimoto Y, Matsumoto T, Ogimi C, Hirata E, Takushi A, Otsuka H (1995). Iridoid glucosides from the leaves and stems of *Duranta erecta*. Phytochemistry 39:829-833.
- Teotino UM, Friz LP, Ganduni A, Bella DD (1963). Thio derivatives of 2,3-dihydro-4H-1,3-benzoazin-4-one synthesis and pharmacological properties. J. Med. Chem. 6:248-250.
- Tovey FI, Capanoglu D, Langley GJ, Herniman JM, Bor S, Ozutemiz O, Hobsley M, Bardhan KD, Linclau B (2011). Dietary phytosterols protective against peptic ulceration. Gastroenterol. Res. 4(4):149-156.
- Trease GE, Evans WC (1985). Pharmacognosy. W.B. Sanders Company, London, 14th Ed.
- Vogel GH, Gang W (2002). Drug Discovery and Evaluation Pharmacological Assay, In methods to induce experimental diabetes mellitus. Heidelberg, Springer Verlag. P 950.
- Vogel HG, Vogel WH (1997). Drug Discovery and Evaluation, Pharmacological Assays, Springer-Verlag, Berlin, Heidelberg, New York. pp. 406-407.
- Vessal M, Hemmati M, Vasei M (2003). Antidiabetic effects of quercetin in streptozocin induced diabetic rats. Comp. Biochem. Physiol. C 135:357-364.
- Villasenor IM, Lamadrid MRA (2006). Comparative anti-hyperglycemic potentials of medicinal plants. J. Ethnopharmacol. 104:129-131.
- Wagner H, Wolf P (1977). New natural products and plant drugs with pharmacological and biological or therapeutic activity, Springer Verlag, Berlin. P 231.
- Wagner WL, Herbst DR, Sohmer SH (1999). Manual of the Flowering Plants of Hawai'i, University of Hawai'i and Bishop Museum Press, Honolulu, HI.
- Winter CA, Risley EA, Nuss CW (1962). Carrageenan-induced edema in hind paw of the rat as an assay for antiinflammatory drugs. Proc. Soc. Exp. Biol. Med. 111:544-547.
- Yalcin FN, Ersoz T, Avci K, Gotfredsen CH, Jensen SR, Calis I (2007). New iridoid glycoside from *Lamium eriocephalum* subsp. *eriocephalum*. Helv. Chim. Acta 90(2):332-336.
- Zanatta L, Sousa E, Cazarolli LH, Junior AC, Pizzolatti MG, Szpoganicz B, Silva FRMB (2007). Effect of crude extract and fractions from *Vitex megapotamica* leaves on hyperglycemia in alloxan-diabetic rats. J. Ethnopharmacol. 109:151-155.