

Full Length Research Paper

***In vitro* modulation of tracheal smooth muscle reactivity by extracts of some Senegalese medicinal plants**

Fatou B. Sarr¹, Mamadou Sarr^{1,2*}, Doudou Diop³, Modou Oumy Kane², Abdoulaye Ba¹, Bocar Sarr², Lamine Guèye¹, Aminata Sall Diallo², Abdoulaye Samb¹, Ramaroson Andriantsitohaina⁴ and Fallou Cissé¹

¹Laboratoire de Physiologie et Explorations fonctionnelles, FMPOS, UCAD, Dakar et Unité Mixte Internationale de Recherches (UMI 3189) 'Environnement, Santé, Sociétés' CNRS-UCAD-CNRST-UNIV.BAMAKO.

²Laboratoire de Physiologie Pharmaceutique, FMPOS, UCAD, Dakar, Senegal.

³Laboratoire de Botanique, IFAN, UCAD, Dakar, Senegal.

⁴Biologie Neuro-Vasculaire Intégrée, UMR-CNRS 6214, INSERM 771, Faculté de Médecine, Université d'Angers, 49045 Angers Cedex, France.

Accepted 23 October, 2009

Scientific validation of pharmacological actions of 7 Senegalese plants used in traditional medicine for the treatment of respiratory illness was carried out. Changes in contractility of isolated rat trachea were assessed in organ chambers. Rings were allowed to equilibrate for 60 min before experiments were carried out, during which time the resting tension was adjusted, as required. Rings were first exposed to methanolic plants extracts (10^{-2} and 10^{-1} mg/ml) or solvent. After a 30 min incubation period, they were contracted with acetylcholine in a cumulative manner (10^{-9} to 10^{-3} M). Extracts from leaves of *Guiera senegalensis*, *Melaleuca leucodendron* and *Hymenocardia acida* elicited a significant dose-dependant inhibition of the contractile agonist, suggesting further investigations on its chemical composition, the underlying mechanisms involved and its potential health value. However, in rings pre-incubated with extracts from *Cymbopogon giganteus* leaves and *Salvadora persica* roots, effect of the contractile agonist was not affected. Moreover, a hypereactivity was observed with extracts from *Gossypium barbadense* leaves and *Cassia occidentalis* seeds. These results provide evidence that Senegalese herbals may be of interest as valuable source of information for the selection of plants for focussed screening programmes and for therapeutically useful products.

Key words: Tracheal contractility; medicinal plants; respiratory illness

INTRODUCTION

Airway hyper-responsiveness and lung inflammation are reported to be the two major factors involved in respiratory illness (del Giudice et al., 2004; Futrakul et al., 2005; Fahy 2006). According to the World Health Organization (WHO), they constitute a public health priority. In fact, according to recent estimates from WHO, 300 million people suffer from asthma, this number could reach 400 million in 2025 and is responsible for over 200,000 deaths per year (Meetoo, 2008). These respiratory diseases, in addition to inflammation (Gerald et al., 2009; Yang et al., 2009), are often associated with bron-

chial hyperreactivity (Hamelmann, 1999). Bronchial responsiveness is the ability of airway response to bronchoconstrictor stimuli, be they physical, chemical or pharmacological. In response to these stimuli, the airways have the ability to reduce their diameter with bronchial responsiveness increased in many lung diseases, allergic or not. The bronchial hyperreactivity leads to abnormal sensitivity of the bronchi to contract excessively in response to stimuli that are ineffective in a normal subject. The prevention and treatment of bronchial hyperreactivity is a goal pursued intensively (Nie et al., 2005; Peters et al., 2006; Kim et al., 2009) and use of medicinal plants is a common practice in our traditional society.

Numerous *in vitro* studies have shown beneficial effects of phytotherapy (del Giudice et al., 2004; Govindan et al.,

*Corresponding author, E-mail: mamadousarr@ucad.sn. Tel: +221 77 506 86 36, Fax: +221 33 865 23 41

2004; Spasov et al., 2004; Schmiedel and Klein 2006; Boskabady et al., 2007) and reported Scientific validation of pharmacological actions of plants used in traditional medicine for the treatment of these pathologies have been reported (Borchers et al., 2000; Loi et al., 2004; Gazzaneo et al., 2005; Kufer et al., 2005; Leporatti and Impieri 2007; Redzic 2007; Nanyingi et al., 2008). However, none of such works cover plants used in Senegalese traditional medicine. Consequently, seven local plants (*Guiera senegalensis*, *Melaleuca leucodendron*, *Hymenocardia acida*, *Cymbopogon giganteus*, *Salvadora persica*, *Gossypium barbadense* and *Cassia occidentalis*) used by traditional health practitioners in Senegal for treatment of respiratory tract that have never been described were investigated.

This study also investigates the relationship between these plants extracts and the cholinergic system, after *in vitro* stimulation of rat trachea with a major neurotransmitter (acetylcholine) that activates smooth muscles in the respiratory tract.

MATERIALS AND METHODS

Materials

Acetylcholine was purchased from Sigma Chemical Co (Saint Quentin-Fallavier, France). Adrenaline was a generous gift from 'Pharmacie Nationale d'Approvisionnement', Dakar, Senegal). All reagent and extract were diluted in Krebs bicarbonate solution before use.

Preparation of methanol extract

Leaves of *G. senegalensis*, *M. leucodendron*, *H. acida*, *C. giganteus* and *G. barbadense*, roots of *S. persica* and seeds of *C. occidentalis* were shade dried and subjected to size reduction to get a coarse powder (Mortar grinder RM100). The powdered material (75 g) was subjected to extraction for 2 h using a 60% methanol/water solution (300 ml/ 200 ml, respectively) as solvent as previously described (Ghayur and Gilani, 2006). The extract was then filtered in vacuum conditions (Vacuum pump V-700, Büchi®) by means of the phial of Kitassato and dried on a rotary evaporator (Büchi®). Methanolic extract evaporation was realized during 3 successive days until the obtaining of a dry crude extract. Evaporation conditions were the following ones: Temperature: +40°C; Cooling: +21°C; Rotation: 4000 tr. / min.

Tracheal reactivity studies

Male Wistar rats (12 weeks old), weighing 150-200 g were procured from local Institute, Dakar, Senegal. They were fed on standard rat feed and given free access to water. Tracheas were removed from rats after anaesthesia with pentobarbital (60 mg/kg, i.p.) and cleaned of connective tissue and cut into rings (3 - 4 mm in length).

Rings were suspended in organ baths (Panlab-TRI 202P) containing oxygenated (95% O₂; 5% CO₂) Krebs bicarbonate solution (mM: NaCl 119, KCl 4.7, KH₂PO₄ 1.18, MgSO₄ 1.18, CaCl₂ 1.25, NaHCO₃ 25 and D-glucose 11, pH 7.4, 37°C) for determination of changes in isometric tension as previously described and adapted (Boskabady and Shaikhi, 2000). Following equilibration for 60 minutes under a resting tension of 2 g, rings were contracted

with acetylcholine (1 µM) and relaxation to phenylephrine (1 µM) was determined. After washout and a 30 min equilibration period, rings were incubated with methanolic plants extracts (10⁻² and 10⁻¹ mg/ml) or solvent for 30 min before a concentration-contraction curve to acetylcholine (10⁻⁹ to 10⁻³ M) was constructed.

Statistical analysis

Values are expressed as mean ± SEM. Statistical evaluation was performed with Student's t test for paired data or ANOVA. Values of p < 0.05 were considered statistically significant.

RESULTS AND DISCUSSION

Several studies have reported the effects of traditional medicine plants in an attempt to rationalize some of its medical uses (Ko et al., 2003; Li et al., 2006; Castro et al., 2006; Ghayur et al., 2007) . This present study provides pharmacological evidence for the medicinal use of Senegalese plants in respiratory disorders.

One major finding of this study is that extracts from *G. senegalensis*, *M. leucodendron* and *H. acida* inhibited the contractions of rat tracheal smooth muscles induced by acetylcholine. To investigate the effects of these medicinal plants, rat trachea were incubated with hydroalcoholic extracts of *G. Senegalensis* leaves, *M. Leucodendron* leaves and *H. acida* leaves. As shown in Figure 1, after a 30 min incubation period, the contractions induced by cumulative concentrations of acetylcholine were significantly inhibited by these extracts when administrated at 10⁻¹ mg/ml, in comparison to control rings not exposed (Emax: 2.142g ± 0.131). These hyporesponsiveness to the contractile agonist was not observed at 10⁻² mg/ml. Moreover, the inhibitory effect evoked by *H. acida* (Emax: 0.675 g ± 0.191; Figure 1F) was more potent than those with either *M. leucodendron* (Emax: 0.972g ± 0.085; Figure 1D) or *G. senegalensis* (Emax: 1.677 g ± 0.408; Figure 1B) which appeared to be the least potent among these 3 extracts. With regards to *G. senegalensis*, an anti-tissue action has been reported. The leaf and roots which contain total alkaloids (Fiot et al., 2006) appears to be the most frequently used plant parts (Silva et al., 2008). No organic toxicity in relation with renal and hepatic functions, heart, lungs, brain, cerebellum, spleen and haematological features in relation with erythropoiesis, haemoglobinogenesis and leucopoiesis was found (Diouf et al., 2000). *H. acida* also contains alkaloids and phenolic contents (Sofidiya et al., 2009). This plants is known to have *in vitro* anti-trypanosomal (De Vooght et al., 2009), antisickling (Farag et al., 2004; Mpiana et al., 2007), antioxidant and antibacterial activities (Sofidiya et al., 2009), but no data is available for its bronchodilatory effect. Melaleuca tree appears not to be a significant source of aeroallergen and the Melaleuca odour is not a respiratory irritant (Stablein et al., 2002).

The results with *C. giganteus* and *S. persica* are not surprising. No data is available about their efficacy against respiratory disorders. As shown in Figure 2, no

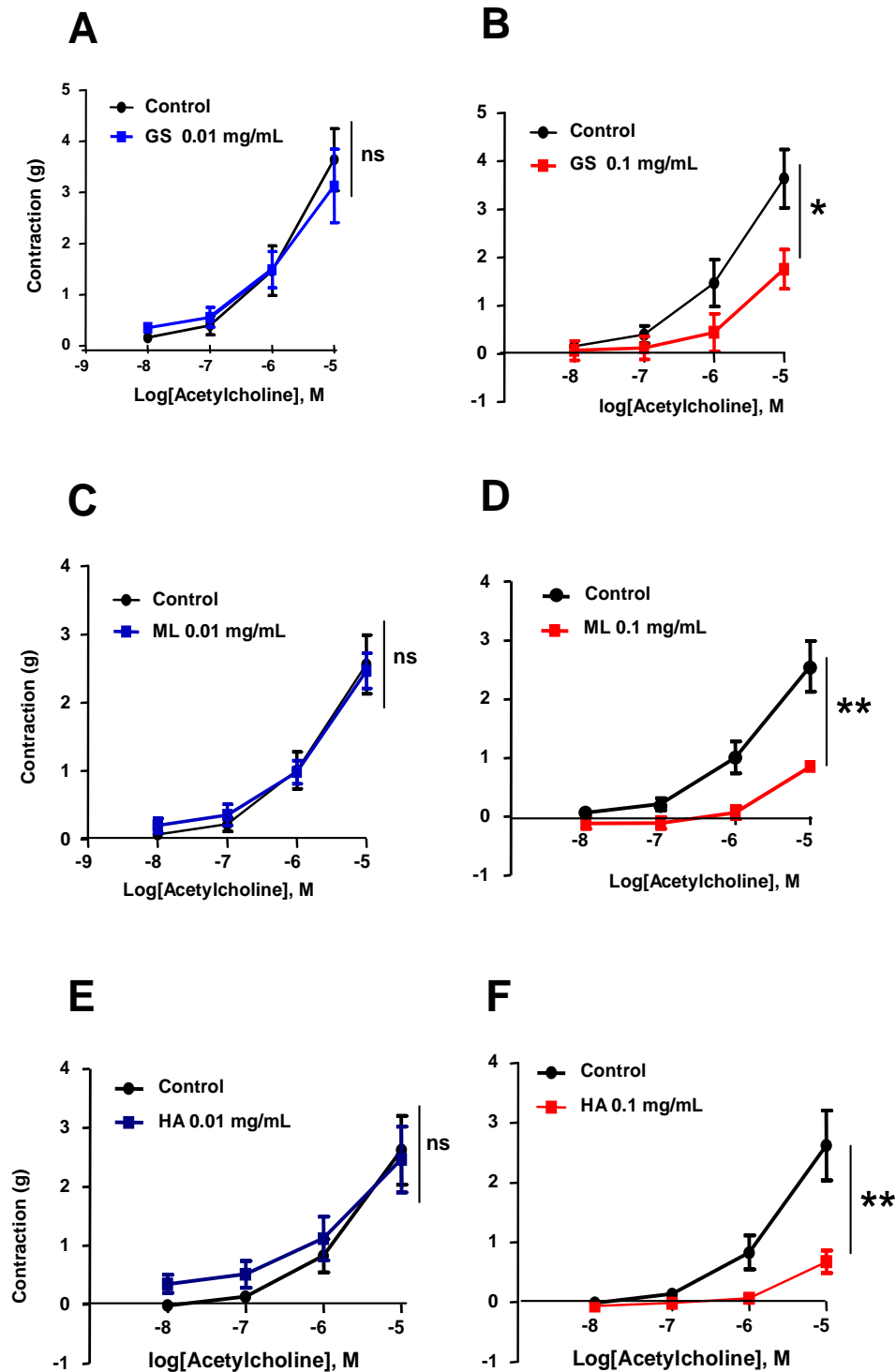


Figure 1. Effect of *in vitro* exposure of the leaves of *Guiera senegalensis*, GS (A and B), *Melaleuca leucodendron*, ML (C and D) and *Hymenocardia acida*, HA (E and F) at 0.01 mg/ml (left panel) or 0.1 mg/ml for 30 min (right panel) on acetylcholine-induced contraction of isolated rat trachea. Each values represents the mean \pm S.E.M, (n = 5). ns = not significant ; *p < 0.05, ** p < 0.01, one-way ANOVA).

significant differences were seen between rings incubated with extracts from *C. Giganteus* leaves (Figures 2A and B) and *S. persica* roots (Figures 2C and D) regarding

tracheal reactivity in comparison to control rings. However, these plants have some efficacy in other biological systems. Indeed, *C. giganteus* was used against chloro-

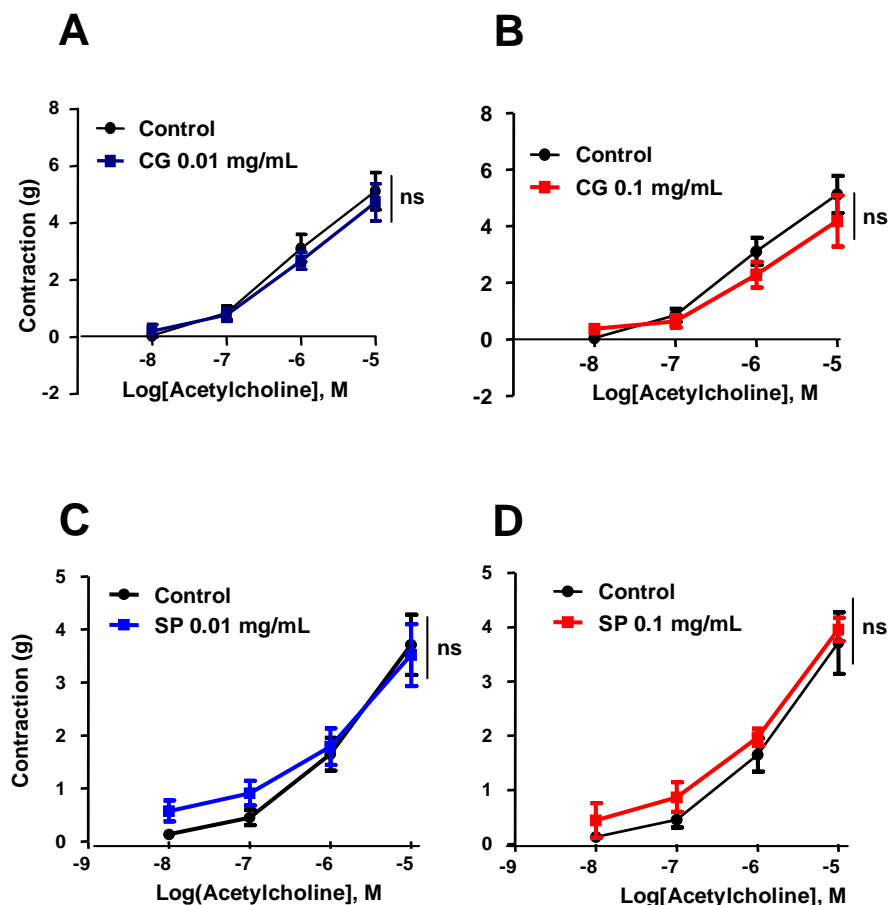


Figure 2. Effect of *in vitro* exposure of *Cymbopogon giganteus*, CG leaves (A and B) and *Salvadora persica*, SP roots (C and D) at 0.01 mg/ml (left panel) or 0.1 mg/ml for 30 min on acetylcholine-induced contraction of isolated rat trachea. Each values represents the mean \pm S.E.M, (n = 5). ns = not significant ; *p < 0.05, one-way ANOVA).

quine resistant *Plasmodium* (Almas and Al-Zeid, 2004; Almas et al., 2005) while an antimicrobial effects (Almas, 1999), a partial removal of smear layer and occlusion of dentinal tubules (Almas, 2001) or as toothbrush tree (Boskabady et al., 2004) used as 'persica mouthwash' resulting in improved gingival health and lower carriage rate of carcinogenic bacteria (Khalessi et al., 2004), a hypolipidemic activity on experimental hypercholesterolemia (Galati et al., 1999) and an anticonvulsive and sedative (Monforte et al., 2002) effect was attributed to *Salvadora persica*. By contrast, extracts from *G. barbadense* leaves and *C. occidentalis* seeds enhanced the contractility of tracheal SMCs. Incubation of rat trachea with extracts from *G. barbadense* leaves and *C. occidentalis* seeds caused a significant hyperactivity. As shown in Figures 3B and D, the contractions induced by acetylcholine at low concentrations (10^{-8} to 10^{-7} M) were significantly increased by these extracts when administered at 10^{-1} mg/ml, in comparison to controls ring, not exposed. However, the maximal responses remain unchanged for the 2 extracts ($E_{max} = 6.09 \text{ g} \pm 0.75$ for *G.*

barbadense; $5.35 \text{ g} \pm 1.48$ for *C. occidentalis* and $6.142 \text{ g} \pm 0.131$ for controls). Perhaps, this might be responsible for *G. barbadense* being useful for the treatment of animal mycoses (Mace et al., 1993). Similarly, *C. occidentalis* appears to be equally highly toxic (Vashishtha et al., 2007; Gupta, 2008; Panwar and Kumar, 2008). This plant (*C. Occidentalis*) is known to cause toxic cardiomyopathy, toxic peripheral neuropathy and muscle atrophy. Histopathology revealed that the heart and liver were the main organs affected, with myocardial necrosis and centrilobular degeneration (Tasaka et al., 2000). This result is similar to the effects reported on aqueous extract of cotton which contains substances that can rapidly cause damage to testicular, liver, kidney and muscular tissues (Ye et al., 2007).

CONCLUSION

Altogether, these results justified the use of some selected plants in the management of respiratory disorders by

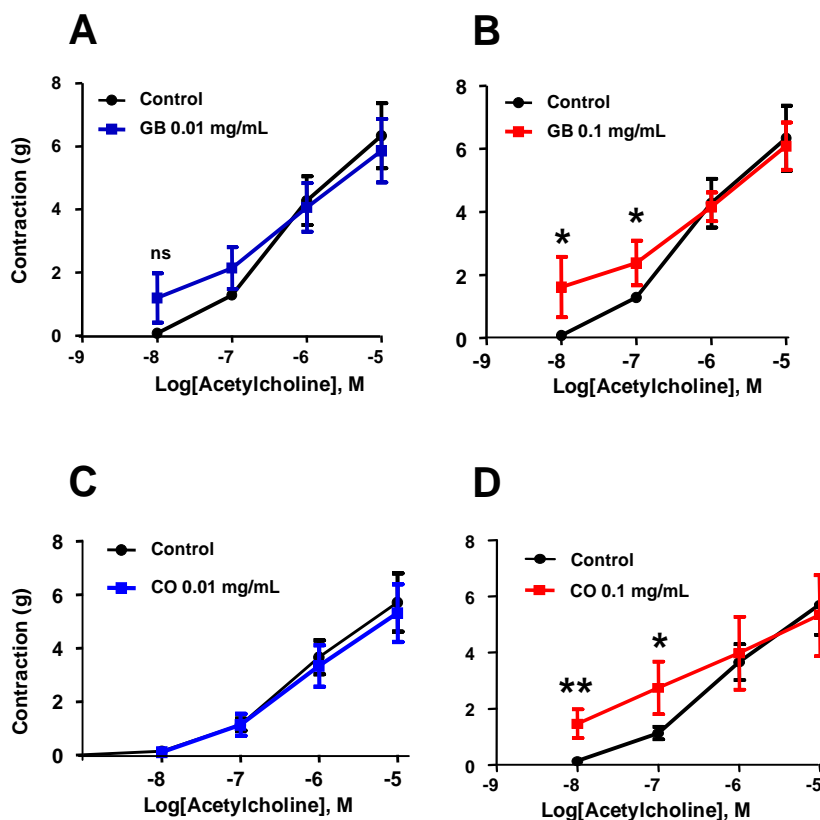


Figure 3. Effect of *in vitro* exposure of *Gossypium barbadense*, GB leaves (A and B) and *Cassia occidentalis*, CO seeds (C and D) at 0.01 mg/ml (left panel) or 0.1 mg/ml for 30 min (right panel) on acetylcholine-induced contraction of isolated rat trachea. Each values represents the mean \pm S.E.M, (n = 5). ns = not significant ; *p < 0.05, one-way ANOVA).

traditional oral health practices which constitute an important part of lifestyle in Senegalese population. The mechanisms by which these plants interact with SMCs contractions remains to be elucidated. However, we can not exclude the involvement of the cholinergic system (stimulated by acetylcholin) and the interaction with calcium signalling.

ACKNOWLEDGMENTS

This work was supported in part by grants from 'Coopération Française/Initiative Codéveloppement'. Dr. Fatou Bintou SAR is a recipient of doctoral fellowship (ED-SEV) supported by Asyilia Gum Company.

REFERENCES

- Almas K (1999). The antimicrobial effects of extracts of *Azadirachta indica* (Neem) and *Salvadora persica* (Arak) chewing sticks. *Indian J. Dent. Res.* 10: 23-26.
- Almas K (2001). The effects of extracts of chewing sticks (*Salvadora persica*) on healthy and periodontally involved human dentine: a SEM study. *Indian J. Dent. Res.* 12: 127-132.
- Almas K, Al-Zeid Z (2004). The immediate antimicrobial effect of a toothbrush and miswak on cariogenic bacteria: a clinical study. *J. Contemp. Dent. Pract.* 5: 105-114.
- Almas K, Skaug N, Ahmad I (2005). An *in vitro* antimicrobial comparison of miswak extract with commercially available non-alcohol mouthrinses. *Int. J. Dent. Hyg.* 3: 18-24.
- Borchers AT, Keen CL, Stern JS, Gershwin ME (2000). Inflammation and Native American medicine: the role of botanicals. *Am. J. Clin. Nutr.* 72: 339-347.
- Boskabady MH, Javan H, Sajady M, Rakhshandeh H (2007). The possible prophylactic effect of *Nigella sativa* seed extract in asthmatic patients. *Fundam. Clin. Pharmacol.* 21: 559-566.
- Boskabady MH, Shaikhi J (2000). Inhibitory effect of *Carum copticum* on histamine (H1) receptors of isolated guinea-pig tracheal chains. *J. Ethnopharmacol.* 69: 217-227.
- Boskabady, MH, Shirmohammadi B, Jandaghi P, Kiani S (2004). Possible mechanism(s) for relaxant effect of aqueous and macerated extracts from *Nigella sativa* on tracheal chains of guinea pig. *BMC Pharmacol.* 4: 3.
- Castro SM, Guerrero-Plata A, Suarez-Real G, Adegboyega PA, Colasurdo GN, Khan AM, Garofalo RP, Casola A (2006). Antioxidant treatment ameliorates respiratory syncytial virus-induced disease and lung inflammation. *Am. J. Respir. Crit. Care Med.* 174: 1361-1369.
- De Vooght V, Vanoirbeek JA, Haenen S, Verbeken E, Nemery B, Hoet PH (2009). Oropharyngeal aspiration: an alternative route for challenging in a mouse model of chemical-induced asthma. *Toxicol.* 259: 84-89.
- Giudice MM, Brunese FP, Piacentini GL, Pedulla M, Capristo C, Decimo F, Capristo AF (2004). Fractional exhaled nitric oxide (FENO), lung function and airway hyperresponsiveness in naive atopic asthmatic

- children. *J. Asthma* 41: 759-765.
- Diouf A, Cisse A, Gueye SS, Mendes V, Siby T, Diouf Diop RM, Bassene E (2000). [Toxicological study of *Guiera senegalensis* Lam (Combretaceae)]. *Dakar Med.* 45: 89-94.
- Fahy JV (2006). Anti-IgE: lessons learned from effects on airway inflammation and asthma exacerbation. *J. Allergy Clin. Immunol.* 117: 1230-1232.
- Farag RS, Shalaby AS, El-Baroty GA, Ibrahim NA, Ali MA, Hassan EM (2004). Chemical and biological evaluation of the essential oils of different *Melaleuca* species. *Phytother. Res.* 18: 30-35.
- Fiot J, Sanon S, Azas N, Mahiou V, Jansen O, Angenot L, Balansard G, Ollivier E (2006). Phytochemical and pharmacological study of roots and leaves of *Guiera senegalensis* J. F. Gmel (Combretaceae). *J. Ethnopharmacol.* 106: 173-178.
- Putrakul S, Deerojanawong J, Prapphal N (2005). Risk factors of bronchial hyperresponsiveness in children with wheezing-associated respiratory infection. *Pediatr. Pulmonol.* 40: 81-87.
- Galati EM, Monforte MT, Forestieri AM, Miceli N, Bader A, Trovato A (1999). *Salvadora persica* L.: hypolipidemic activity on experimental hypercholesterolemia in rat. *Phytomedicine* 6: 181-185.
- Gazzaneo LR, de Lucena RF, de Albuquerque UP (2005). Knowledge and use of medicinal plants by local specialists in an region of Atlantic Forest in the state of Pernambuco (Northeastern Brazil). *J. Ethnobiol. Ethnomed.* 1: 9
- Geraldes L, Todo-Bom A, Loureiro C (2009). [Airways inflammation evaluation. Upper and lower airways]. *Rev. Port. Pneumol.* 15: 443-460.
- Ghayur MN, Gilani AH (2006). Studies on cardio-suppressant, vasodilator and tracheal relaxant effects of *Sarcococca saligna*. *Arch Pharm. Res.* 29: 990-997.
- Ghayur MN, Khan H, Gilani AH (2007). Antispasmodic, bronchodilator and vasodilator activities of (+)-catechin, a naturally occurring flavonoid. *Arch. Pharm. Res.* 30: 970-975.
- Govindan S, Viswanathan S, Vijayasekaran V, Alagappan R (2004). Further studies on the clinical efficacy of *Solanum xanthocarpum* and *Solanum trilobatum* in bronchial asthma. *Phytother. Res.* 18: 805-809.
- Gupta AK (2008). *Cassia occidentalis* poisoning causes fatal coma in children in western Uttar Pradesh. *Indian Pediatr* 45-424.
- Hamelmann E (1999). [Genesis of airway inflammation and hyper-reactivity: insight into the asthma mouse model]. *Pneumologie* 53: 307-312.
- Henneicke-von Zepelin H, Hentschel C, Schnitker J, Kohnen R, Kohler G, Wustenberg P (1999). Efficacy and safety of a fixed combination phytomedicine in the treatment of the common cold (acute viral respiratory tract infection): results of a randomised, double blind, placebo controlled, multicentre study. *Curr. Med. Res. Opin.* 15: 214-227.
- Khalessi AM, Pack AR, Thomson WM, Tompkins GR (2004). An in vivo study of the plaque control efficacy of *Persica*: a commercially available herbal mouthwash containing extracts of *Salvadora persica*. *Int. Dent. J.* 54: 279-283.
- Kim D, Kim SH, Park EJ, Kang CY, Cho SH, Kim S (2009). Anti-allergic effects of PG102, a water-soluble extract prepared from *Actinidia arguta*, in a murine ovalbumin-induced asthma model. *Clin. Exp. Allergy* 39: 280-289.
- Ko WC, Liu PY, Chen JL, Leu IJ, Shih CM (2003). Relaxant effects of flavonoids in isolated guinea pig trachea and their structure-activity relationships. *Planta. Med.* 69: 1086-1090.
- Kufer J, Forther H, Poll E, Heinrich M (2005). Historical and modern medicinal plant uses--the example of the Ch'orti' Maya and Ladinos in Eastern Guatemala. *J. Pharm. Pharmacol.* 57: 1127-1152.
- Leporatti ML, Impieri M (2007). Ethnobotanical notes about some uses of medicinal plants in Alto Tirreno Cosentino area (Calabria, Southern Italy). *J. Ethnobiol. Ethnomed* pp.3-34.
- Li PB, Ma Y, Wang YG, Su WW (2006). [Experimental studies on anti-tussive, expectorant and antiasthmatic effects of extract from *Citrus grandis* var. *tomentosa*]. *Zhongguo Zhong Yao Za Zhi* 31: 1350-1352.
- Loi MC, Poli F, Sacchetti G, Selenu MB, Ballero M (2004). Ethnopharmacology of ogliastra (villagrande strisaili, sardinia, Italy). *Fitoterapia* 75: 277-295.
- Mace ME, Stipanovic RD, Bell AA (1993). Toxicity of cotton phytoalexins to zoopathogenic fungi. *Nat Toxins* 1: 294-295.
- Meetoo D (2008). Chronic diseases: the silent global epidemic. *Br. J. Nurs.* 17: 1320-1325.
- Monforte MT, Trovato A, Rossitto A, Forestieri AM, D'Aquino A, Miceli N, Galati EM (2002). Anticonvulsant and sedative effects of *Salvadora persica* L. stem extracts. *Phytother Res.* 16: 395-397.
- Mpiiana PT, Tshibangu DS, Shetonde OM, Ngbolua KN (2007). In vitro antitrepanocytary activity (anti-sickle cell anemia) of some congolese plants. *Phytomedicine* 14: 192-195.
- Nanyingi MO, Mbaria JM, Lanyasanya AL, Wagate CG, Koros KB, Kaburia HF, Munenge RW, Ogara WO (2008). Ethnopharmacological survey of Samburu district, Kenya. *J. Ethnobiol. Ethnomed.* 4-14.
- Nie X, Li Q, Cai G, Dai Y, Zhang J (2005). The effect of N-acetylcysteine on Clara cells and Clara cell 16 kDa protein in a murine model of allergen-induced airway inflammation. *Respirol.* 10: 157-163.
- Panwar RS, Kumar N (2008). *Cassia occidentalis* toxicity causes recurrent outbreaks of brain disease in children in Saharanpur. *Indian J. Med. Res.* 127: 413-414.
- Peters M, Kauth M, Schwarze J, Korner-Rettberg C, Riedler J, Nowak D, Braun-Fahrlander C, von Mutius E, Bufe A, Holst O (2006). Inhalation of stable dust extract prevents allergen induced airway inflammation and hyperresponsiveness. *Thorax* 61: 134-139.
- Redzic SS (2007). The ecological aspect of ethnobotany and ethnopharmacology of population in Bosnia and Herzegovina. *Coll Antropol.* 31: 869-890.
- Schmiedel V, Klein P (2006) A complex homeopathic preparation for the symptomatic treatment of upper respiratory infections associated with the common cold: An observational study. *Explore (NY)* 2: 109-114.
- Silva O Serrano R, Gomes ET (2008). Botanical characterization of *Guiera senegalensis* leaves. *Microsc Microanal* 14: 398-404.
- Sofidiya MO, Odukoya OA, Afolayan AJ, Familoni OB (2009). Phenolic contents, antioxidant and antibacterial activities of *Hymenocardia acida*. *Nat. Prod. Res.* 23: 168-177.
- Spasov AA, Ostrovskij OV, Chernikov MV, Wikman G (2004). Comparative controlled study of *Andrographis paniculata* fixed combination, Kan Jang and an Echinacea preparation as adjuvant, in the treatment of uncomplicated respiratory disease in children. *Phytother Res.* 18: 47-53.
- Stablein JJ, Bucholtz GA, Lockey RF (2002). *Melaleuca* tree and respiratory disease. *Ann Allergy Asthma Immunol.* 89: 523-530.
- Tasaka AC, Weg R, Calore EE, Sinhorini IL, Dagli ML, Haraguchi M, Gorniak SL (2000). Toxicity testing of *Senna occidentalis* seed in rabbits. *Vet. Res. Commun.* 24: 573-582.
- Vashishtha VM, Kumar A, John TJ, Nayak NC (2007). *Cassia occidentalis* poisoning as the probable cause of hepatomyoencephalopathy in children in western Uttar Pradesh. *Indian J. Med. Res.* 125: 756-762.
- Yang M, Kumar RK, Foster PS (2009). Pathogenesis of steroid-resistant airway hyperresponsiveness: interaction between IFN-gamma and TLR4/MyD88 pathways. *J. Immunol.* 182: 5107-5115.
- Ye W, Giblin-Davis RM, Davies KA, Purcell MF, Scheffer SJ, Taylor GS, Center TD, Morris K, Thomas WK (2007) Molecular phylogenetics and the evolution of host plant associations in the nematode genus *Fergusobia* (Tylenchida: *Fergusobiinae*). *Mol. Phylogenet. Evol.* 45: 123-141.