Full Length Research Paper

Screening of aqueous plant extracts against *Beauveria* Bassiana infection to 5th instar larvae of Bombyx mori L.

Jyoti Ashok Chavan*, Santosh Jalindar Patil and Ganesh Parashram Bhawane

Department of Zoology, Shivaji University, Kolhapur-416 004, MS, India.

Accepted 17 May, 2011

Use of plant based drugs and chemicals for curing various ailments and personal adornment is as old as human civilization. The aqueous extract of *Argemone mexicana*, *Terminalia arjuna*, *Syzigium cumini* and *Annona squamosa* were tested for antifungal activity *in vitro*. The aim of study was to find out the effect of aqueous extracts on *Beauveria bassiana* infected *Bombyx mori* L. larvae. The aqueous extract of *A. Mexicana* shows effective results as compared to other botanicals. However it is clear that aqueous plant extracts showed decreased mortality with effective rate of rearing.

Key words: Bombyx mori, Beauveria bassiana, Argemone mexicana, Terminalia arjuna, Syzigium cumini, Annona squamosa, effective rate of rearing (ERR) percentage.

INTRODUCTION

Many Indian plants have been used from time immemorial to treat various diseases and infections in traditional medicinal system of India such as Siddha, Ayurvedha and Nataropathy (Shekhawat and Prasad, 1971; Khanna and Chandra, 1972). Pharmacological studies have acknowledged the value of medicinal plants as potential source of bioactive compounds (Prusti et al., 2008). Medicinal plants are rich source of novel drugs that forms the ingredients in traditional systems of medicine, modern medicine, food supplements, folk medicines, pharmaceutical intermediates, bioactive principles and lead compounds in synthetic drugs. Phytochemicals from medicinal plants serve as lead compounds in drug discovery and design (Ebi and Ofoefule, 2000). The traditional method of extraction involves a number of solvents that are used in various cases (Ganesan and Krishnaraju, 1995). Among the fungal disease of silkworm, white muscardine is found to be serious caused by Beauveria bassiana (Bassi, 1835). Cocoon crop loss in different contries of about 5 to 50% reported by Jhansi Lakshmi (2003).

A wide variety of plant secondary metabolites have been identified as active principles for the treatment of various ailments (Taylor et al., 2001; Ncube et al., 2008). Earlier studies have indicated the possibility of using botanical extracts for reducing white muscardine disease and effective in inhibiting the germination of spores. An aqueous extracts of *Terminalia chebula* fruit exhibit antifungal activity against a number of dermatophytes and yeast (Dutta et al., 1998). The broad spectrum of antibacterial activity was reported for Terminalia arjuna (Singh et al., 2008). Bark of T. arjuna has been attributed to possesess cardio protective properties as described by Vagbhatta, in 'Astang Hridayam' (Dwivedi, 2007). Argemone Mexicana seed extract are effective against cutaneous infection and skin diseases (Singh et al., 2009). Plant Annona squamosa is commonly found in Malaysia and used in folk medicine against skin infections. Different parts of *Syzygium cumini* such as seeds, bark, fruit and leaves have been used in traditional medicine in many countries (Rahman et al., 1989, Teixeira et al., 1997).

So in the present study the efficiency of aqueous extract of *A. mexicana*, *T. arjuna*, *S. cumini*, *Asquamosa*. is tested against larval characters, effective rate of rearing (ERR) and mortality in silkworm race CSR2 infected with *B. Bassiana*.

^{*}Corresponding author. E-mail: jyotijeevanjyoti@rediffmail.com. Tel: 956107686

Table 1. List of plants.

Botanical name	Local name	Family	Part used	Traditional uses
Argemone mexicana L.	Pivala Dhotara	Papaveraceae	Seeds	Skin infection, warts, sores, dropsy, jaundice and antibacterial.
<i>Terminalia arjuna</i> (Roxb ex DC) Wight	Arjun	Combretaceae	Bark	Cardiovascular diseases, myocardial, degenerative neurological diseases, cancer, amyloidosis, acute pancreatitis, anthritis, antherosclerosis, inflammatory bowl diseases, diabetes, senile dementia, retinal degeneration and senile cataract.
<i>Syzygium cumini</i> L. Skeel	Jamun	Myrtaceae	Seeds	Diabetes, hypoglycemia, antioxidant, antimicrobial astringent, used for sore throat, diarrhea, dysentery.
Annona squamosa L.	Sitafal	Annonaceae	Seeds	Antibacterial, antifungal, antiviral, sedative to heart and relieves vomiting.

MATERIALS AND METHODS

Selection of medicinal plants to study

In the present work few selected medicinal plants were screened for potential antifungal activity. These plants are listed in Table 1.

Identification and preservation of plant material

Fresh plant parts were collected from adjacent area of Kolhapur District shown in Plate 1. The taxonomic identification of the plants was made with available literature (Yadav and Sardesai 2002).

Preparation of plant extract

The collected plant material was washed with distilled water and shade dried at room temperature. The materials were grinded to fine powder with the help of mixer grinder. Then these powdered materials were used for preparation of aqueous extracts by using 50 g powder in 100 ml double distilled water for 72 h. The mixture was stirred every 24 h using a sterile glass rod. At the end of extraction each extract was concentrated in vacuum at 30° C and stored at 4° C until further use.

Preparation of pathogenic suspension

The pure fungal spores grown on the broth were harvested on the tenth day of inoculation with sterile distilled water using a sterile glass rod and the suspension was filtered through double layered muslin cloth. The original conidial stock suspension was serially diluted in sterile distilled water of atleast 1x10⁹ spores/ml.

Artificial induction of fungal spores and plant extract to *Bombyx mori*

The B. bassiana infection was given to freshly moulted five instar

larvae after two h starvation. The 60 larvae were taken in each tray and sprayed with the LD_{50} concentration of fungal spores. After 6 h of treatment the larvae were segregated to appropriate group and deeped in 5000 ppm concentration of different plant extract. One group was kept as inoculated control in which larvae not deeped in any plant extract and another group was kept as normal control in which larvae reared on fresh mulberry leaves without any spraying. The treatment of plant extracts are given for three days at morning time.

RESULTS AND DISCUSSION

The botanical extracts have exhibited significant results on larval character, effective rate of rearing and extent of mortality. The larval character showed significant results in increasing larval weight from all the botanical extract treated groups as per with normal control than the inoculated control (Table 2). Patil et al. (1997) reported that extrafoliation leaf water extract of Parthenium sp. has increased the larval weight, ERR of B. mori. Murugan et al. (1994) observed that extra foliation of aqueous extract of leaves of Tribulus terrestris and Phylanthus niruri increased the total body weight of *B. mori.* These reports are in agreement with the present finding where the aqueous plant extract showed increase in larval weight gram per 10 larvae. In the present study all the plant extracts shows significant ERR and among these four plant extract A. mexicana recorded maximum ERR that is 71.66% and minimum was recorded for S. cumini that is 60.66 % (Tables 3 to 4). Phytochemical compounds of these plants suppressed the spore formation of white muscardine. These results are in accordance or same line with the extrafoliation of Psoralea coryleifolia and T. terrestris at 800 ppm to mulberry leaves once during 3rd instar suppressed the grasserie disease by 80% (Sivaprakasam and Rabindra, 1996). Raghavaiah et al.,



Argemone mexicana L. (Papaveraceae)

Terminalia arjuna (Roxb ex DC) Wight (Combretaceae)



Syzygiumcumini L. Skeel (Myrtaceae)



Annona squamosa L. (Annonaceae)

Plate 1. Plant parts collected from Kolhapur District.

Table 2. Effect of plant extracts on larval weight of Bombyx mori L.

Groups	Days						
Groups	1	2	3	4	5	6	7
Normal control	8.63	14.92	26.63	31.47	37.14	42.3	44.83
Inoculated control	8.67	10.87	21.58	28.27	32.8	37.68	39.44
A. mexicana	8.61	13.09	22.43	29.21	33.76	41.14	42.71
T. arjuna	8.69	13.74	22.29	29.49	34.69	37.36	37.69
S. cumuni	8.7	13.05	22.28	29.63	35.91	40.2	40.69
A. squmqsa	7.61	17.49	20.49	31.55	35.1	40.48	40.35

Table 3. Effect of plant extracts on mortality and mortality in percentage of Bombyx mori L.

Groups	Mortality	Mortality (%)
Normal control	5	8.33
Inoculated control	34	56.66
A. mexicana	17	28.33
T. arjuna	21	35
S. cumuni	24	40
A. squmqsa	22	36.66

Table 4. Effect of plant extracts on cocoon yield and cocoon yield % of Bombyx mori L

Groups	Cocoon yield	Cocoon yield (%)
Normal control	55	91.66
Inoculated control	24	43.33
A. mexicana	43	71.66
T. arjuna	39	65.66
S. cumuni	36	60.66
A. squmqsa	38	63.33

(1987) concluded that aqueous extract of Allium sativum (50%) was most effective in inhibiting the germination of spores of B. bassiana. All these results confirms the aqueous plant extract showed the disease reducing properties. This may be due to the presence of chemical constituent like, Alkaloids, Flavonoids, Glycosides, Steroids, Tannins, Eugenol, Farnesol, Geraniol, Rutin, Stigmasterol. Limonine, linalool. Hexacantanol, Jambosine, Arjunine, Lactone and Arjunetin, in A. Mexicana, A. sgamosa, S. cumini and T. arjuna which helped in preventing the germination of spores of B. bassiana which indirectly helped in ERR (Singh et al., 2009: Patel and Kumar 2008: Ahmad and Beg. 2001). Further these results are agreement with Barnabas and Nagrajan (1958) they reported that the flavonoid rich extracts prepared from *Eucalyptus teriticornis* exhibited antifungal and antibacterial activity against several test organisms including Aspergillus flavus and Candida albicam.

Hence in the present study some of the plant extracts having antifungal properties were identified which could be an effective tool in improving the general health of the silkworm and they might be used in reducing mortality and improving their economic parameters in muscardine infected larvae under *in vivo* condition by decreasing the disease incidence.

ACKNOWLEDGEMENTS

The authors are grateful to the University Grants Commission (UGC), New Delhi for their financial support through the scheme no. 33-367/2007 (SR) dated 10th march 2008. Thanks also to Department of Zoology, Shivaji University, Kolhapur for providing the facilities for research work.

REFERENCES

- Ahmad I, Beg AZ (2001). Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multi-drugs resistant human pathogens. J. Ethnopharmacol., 74: 113-123.
- Barnabas CG, Nagrajan S (1988). Antimicrobial activity of flavonoids of some medicinal plants. Fitoterapia, 59(6): 508-510.

Bassi A (1835). And the sign of the plaster or disease that dormice afflicts bugs daseta Part-I, Tip Terica Orcesi, Lod, pp. 1-67.

Dutta BK, Rahman I, Das TK (1998). Antifungal activity of Indian plant

extracts, Mycoses, 41(11-12): 535-536.

- Dwivedi S (2007). *Terminalia arjuna* Weight and Arn- a useful drug for cardiovascular disorders. J. Ethnopharmacol., 114(2): 114-129.
- Ebi GC, Ofoefule SI (2000). Antimicrobial activity of *Pterocarpus osun* stems. Fitoterapia, 71: 433-435.
- Ganesan T, Krishnaraju JC (1995). Antifugal property of wild plants. Adv. Plant Sci., 8: 194-196.
- Jhansi LVVNS (2003). Ultrastructural studies on tissue of the silkworm, Bombyx mori L. infected with Beauveria bassiana (Balsmo) Vuillemin Ph. D. thesis submitted to Sri Padmavathi Vishvavidyalayam, Tirupati.
- Khanna KK, Chanrda S (1972). Antifungal activity of some plant extracts. Proc. Nat. Acd. Sci. India, 42(8): 111.
- Murugan K, Jeyabalan D, Sivaprakasan N, Senthil VR (1994). Influence of certain botanicals on the biochemical changes in mulberry silkworm, *Bombyx mori* L. (Lepidoptera:Bombycidae). Proceeding of the Second National Seminar on "Prospects and Problems of Sericulture in India", held at Madras University, Vellore during March, pp. 125-130.
- Ncube NS, Afolayan AJ, Okohs AL (2008). Assessment techniques of antimicrobial properties of natural compounds of plant origin: Current methods and future trends. Afr. J. Biotechnol., 7: 1797-1806.
- Patel JD, Kumar V (2008). *Annona squamosa* L.: Phytochemical analysis and antimicrobial screening. J. Pharm. Res., 1(1): 34-38.
- Patil RR, Mahadevappa M, Mahesha HM, Patil VC (1997).
 Phagostimulant effects of Parthenium on mulberry silkworm (Bombyx mori L.). "First International Conference on Parthenium Management", Dharwad, Karnataka, India, Oct. 6-8, pp. 81-85.
- Prusti A, Mishra SR, Sahoo S, Mishra SK (2008). Antimicrobial activity of some Indian Medicinal Plants. Ethnobot. Leaflets, 12: 227-230.
- Raghavaiah G, Jayramaiah M (1987). Antifungal activity of selected plant extracts against the white muscardine fungus. Curr. Res., 17: 62-64.
- Rahman AU, Zaman K (1989). Medicinal plants with hypoglycemic activity. J. Ethnopharmacol., 26: 2720-2722.
- Shekhawat PS, Prasad R (1971). Antifungal properties of some plant extracts and their inhibition of spore germination. Indian phytopathol., 24: 800-802.
- Singh DV, Gupta MM, Santha KTR, Saikia D, Khanuja SPS (2008). Antimicrobial principles from the bark of *Terminalia arjuna*. Curr. Sci., 94(1): 27-29.
- Singh SK, Pandey VD, Singh A, Singh C (2009). Anti-bacterial activity of seed extracts of Argemone mexican L on some pathogenic bacterial strains. Afr. J. Biotechnol., 8(24): 7077-7081.
- Sivaprakasam N, Rabindra RJ (1996). Integrated disease management methods for grasserie in silkworm, *B. mori* L. Indian J. Seric., 35: 122-127.
- Taylor JLS, Rabe T, Mcgaw LJ, Jayer AK, Van SJ (2001). Towords the scientific validation of traditional medicinal plants. Plant growth regul., 34: 23-37.
- Teixeira CC, Pinto LP, Kessler FHP, Knijnik L, Pinto CP, Gastaldo GJ, Fuchs FD (1997). The effect of *Syzygium cumini* L. Skeels on post – prandial blood glucose level in non-diabetic rats and rats with streptozotocin-induced diabetes mellitus. J. Ethnopharmacol., 56: 209-213.
- Yadav SR, Sardesai MM (2002). Flora of Kolhapur District, Shivaji Univerity Publication, pp. 32, 39, 191, 193.